

PCT

WORLD INTELLECTUAL PROPERTY ORGANIZATION
International Bureau



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification: C12N 15/12, 15/62, C07K 14/005, 14/51, 14/52, 14/54, 16/46, 19/00
(11) International Publication Number: WO 98/08949
(53) International Publication Date: 5 March 1998 (05.03.98)

(21) International Application Number: PCT/US97/14859

(22) International Filing Date: 29 August 1997 (29.08.97)

(30) Priority Data:
08 713,556 30 August 1996 (30.08.96) US

(71) Applicant: GENETICS INSTITUTE, INC. (US/US); 87 CambridgePark Drive, Cambridge, MA 02140 (US).

(72) Inventors: LARSEN, Glenn, R., 112 Maynard Farm Road, Sudbury, MA 01776 (US); SAKO, Dianne, S., 50 Jamaica Way #6, Boston, MA 02130 (US); CHANG, Xiao-Jia; 59 Commonwealth Park West, Newton Centre, MA 02159 (US); VELDMAN, Geertruida, M., 60 Woodmere Drive, Sudbury, MA 01776 (US); CUMMING, Dale, 57 Hammond Street, Acton, MA 01720 (US); KUMAR, Ravindra; 533 School Street, Belmont, MA 02178 (US); SHAW, Gray; 133 Irving Street, Cambridge, MA 02140 (US).

(74) Agent: SPRUNGER, Suzanne, A., Genetics Institute, Inc., 87 CambridgePark Drive, Cambridge, MA 02140 (US).

(81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, GH, HU, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, UZ, VN, YU, ZW, ARIPO patent (GH, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).

Published

With international search report.
Before the expiration of the time limit for amending the claims and to be republished in the event of no amendments.

(54) Title: P-SELECTIN LIGAND PROTEINS

		Selectin Binding		
		P	E	L
YYYD 19.Fc		+++	+	-
FFYD 19.Fc		++	+	-
FFYD 19.Fc		+	+	-
FFFD 19.Fc		+	+	-
FFYDN1619.Fc		-	-	-

(57) Abstract

A novel P-selectin ligand glycoprotein is disclosed, comprising the amino acid sequence set forth in SEQ ID NO 2 or by the amino acid sequence set forth in SEQ ID NO 4. DNA sequences encoding the P-selectin ligand protein are also disclosed, along with vectors, host cells, and methods of making the P-selectin ligand protein. Pharmaceutical compositions containing the P-selectin ligand protein and methods of treating inflammatory disease states characterized by P-selectin- and E-selectin-mediated intercellular adhesion are also disclosed.

FOR THE PURPOSES OF INFORMATION ONLY

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT

AL	Albania	ES	Spain	LS	Lesotho	SI	Slovenia
AM	Armenia	FI	Finland	LT	Lithuania	SK	Slovakia
AT	Austria	FR	France	LU	Luxembourg	SN	Senegal
AU	Australia	GA	Gabon	LV	Latvia	SZ	Swaziland
AZ	Azerbaijan	GB	United Kingdom	MC	Monaco	TD	Chad
BA	Bosnia and Herzegovina	GE	Georgia	MD	Republic of Moldova	TG	Togo
BB	Barbados	GH	Ghana	MG	Madagascar	TJ	Tajikistan
BE	Belgium	GN	Guinea	MK	The former Yugoslav Republic of Macedonia	TM	Turkmenistan
BF	Burkina Faso	GR	Greece	ML	Mali	TR	Turkey
BG	Bulgaria	HU	Hungary	MN	Mongolia	TT	Trinidad and Tobago
BJ	Benin	IE	Ireland	MR	Mauritania	UA	Ukraine
BR	Brazil	IL	Israel	MW	Malawi	UG	Uganda
BY	Belarus	IS	Iceland	MX	Mexico	US	United States of America
CA	Canada	IT	Italy	NE	Niger	UZ	Uzbekistan
CF	Central African Republic	JP	Japan	NL	Netherlands	VN	Viet Nam
CG	Congo	KE	Kenya	NO	Norway	YU	Yugoslavia
CH	Switzerland	KG	Kyrgyzstan	NZ	New Zealand	ZW	Zimbabwe
CI	Côte d'Ivoire	KP	Democratic People's Republic of Korea	PL	Poland		
CM	Cameroun	KR	Republic of Korea	PT	Portugal		
CN	China	KZ	Kazakhstan	RO	Romania		
CU	Cuba	LC	Saint Lucia	RU	Russian Federation		
CZ	Czech Republic	LI	Liechtenstein	SD	Sudan		
DE	Germany	LK	Sri Lanka	SE	Sweden		
DK	Denmark	LR	Liberia	SG	Singapore		
EE	Estonia						

P-SELECTIN LIGAND PROTEINS

This application is a continuation-in-part of depending application U.S. Serial No. 08/428,734, filed April 25, 1995, which was a continuation-in-part of depending application U.S. Serial No. 08/316,305, filed September 30, 1994, which was a continuation-in-part of depending application U.S. Serial No. 08/235,398, filed April 28, 1994, which was a continuation-in-part of depending application U.S. Serial No. 08/112,608, filed August 26, 1993, which was a continuation-in-part of U.S. Serial No. 07/965,662, filed October 23, 1992, now abandoned. This application also claims priority from International Application No. PCT/US93/11085, filed October 22, 1993.

BACKGROUND OF THE INVENTION

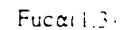
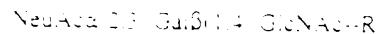
The present invention relates to the field of anti-inflammatory substances which act by inhibiting leukocyte adhesion to endothelial cells. More particularly, the present invention is directed to novel ligands for the mammalian adhesion proteins known as selectins.

During inflammation leukocytes adhere to the vascular endothelium and enter subendothelial tissue, an interaction which is mediated by specific binding of the selectin or LEC-CAM class of proteins to ligands on target cells. Such selectin-mediated cellular adhesion also occurs in thrombotic disorders and parasitic diseases and may be implicated in metastatic spread of tumor cells.

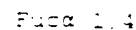
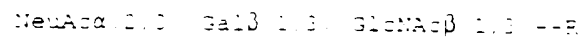
The selectin proteins are characterized by a N-terminal lectin-like domain, an epidermal growth factor-like domain, and regions of homology to complement binding proteins. Thus far three human selectin proteins have been identified, E-selectin (formerly ELAM-1), L-selectin (formerly LAM-1) and P-selectin (formerly PADGEM or GMP-140). E-selectin is induced in endothelial cells several hours after activation by cytokines, mediating the calcium-dependent interaction between neutrophils and the endothelium. L-selectin is the lymphocyte homing receptor, and P-selectin rapidly appears on the cell surface of platelets when they are activated, mediating calcium-dependent adhesion of neutrophils or monocytes to platelets. P-selectin is also found in the Weibel-Palade bodies of endothelial cells; upon its release from these vesicles P-selectin mediates early binding of neutrophils to histamine- or thrombin-stimulated endothelium.

Selectins are believed to mediate adhesion through specific interactions with ligands present in the surface of target cells. Generally the ligands of selectins are comprised of at least in

and also to carbohydrates having the terminal structure



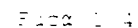
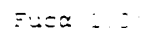
and also to carbohydrates having the terminal structure



where R = the remainder of the carbohydrate chain. These carbohydrates are known blood group antigens and are commonly referred to as sialyl Lewis^x and sialyl Lewis^y, respectively. The presence of the sialyl Lewis^x antigen alone on the surface of an endothelial cell may be sufficient to promote binding to an E-selectin expressing cell. E-selectin also binds to carbohydrates having the terminal structures



and



As with E-selectin, each selectin appears to bind to a range of carbohydrates with varying affinities. The strength of the selectin mediated adhesive event (binding affinity) may also depend on the density of the carbohydrate and on the density of the selectin on the cell surface.

P-selectin binds to carbohydrates containing the non-sialated form of the Lewis^x blood group antigen and with higher affinity to sialyl Lewis^x. P-selectin may also recognize sulfatides, which are heterogeneous 3-sulfated galactosyl ceramides, isolated from myeloid and tumor cells by lipid extraction. However, the binding of cells bearing P-selectin to cells bearing P-selectin ligands is abolished when the ligand-bearing cells are treated with proteases, indicating that the P-selectin ligand may be a glycoprotein.

Two putative soluble protein ligands for P-selectin have recently been identified, one of which has been partially purified. Moore et al. (1) Cell Biol. 135: 443-456 (1992). However, neither amino acid composition, nor the amino acid sequence of these glycoproteins are disclosed.

SUMMARY OF THE INVENTION

10 In one embodiment, the present invention provides a composition comprising an isolated DNA encoding a P-selectin ligand protein, said protein comprising the amino acid sequence set forth in SEQ ID NO 2 from amino acid 1 to amino acid 402. Also provided is a composition comprising an isolated DNA encoding a soluble P-selectin ligand protein, said protein comprising the amino acid sequence set forth in SEQ ID NO 2 from amino acid 1 to amino acid 310. The invention further provides a composition comprising an isolated DNA encoding a mature P-selectin ligand protein, said protein comprising the amino acid sequence set forth in SEQ ID NO 2 from amino acid 42 to amino acid 402. In another embodiment, the invention provides a composition comprising an isolated DNA encoding a soluble mature P-selectin ligand protein, said protein comprising the amino acid sequence set forth in SEQ ID NO 2 from amino acid 42 to amino acid 310. In another embodiment, the invention provides a composition comprising an isolated DNA encoding a P-selectin ligand protein, said protein comprising the amino acid sequence set forth in SEQ ID NO 4. The invention further provides a composition comprising an expression vector comprising any one of the isolated DNAs of the invention, said DNA being operably linked to an expression control sequence, a host cell transformed with the expression vector containing any one of the DNAs described above, and a process for producing the P-selectin ligand protein, which comprises:

- (a) culturing a host cell transformed with an expression vector containing any one of the DNAs of the invention in a suitable culture medium; and
- (b) purifying the P-selectin ligand protein from the culture medium.

30 In another embodiment, the invention provides a composition comprising a protein comprising the amino acid sequence set forth in SEQ ID NO 2 from amino acid 21 to amino acid 402, said protein being substantially free from other mammalian proteins. The invention further comprises a soluble P-selectin ligand protein comprising the amino acid sequence set forth in SEQ ID NO 2 from amino acid 21 to amino acid 310, said protein being substantially free from other mammalian proteins. In another embodiment, the invention comprises a P-selectin ligand protein comprising the amino acid sequence set forth in SEQ ID NO 2 from amino acid 1 to amino acid 402, said protein being substantially free from other mammalian proteins. The invention also provides a composition comprising a mature P-selectin ligand protein comprising the amino acid

sequence set forth in SEQ ID NO 2 from amino acid 42 to amino acid 402, said protein being substantially free from other mammalian proteins. Further provided is a composition comprising a soluble mature P-selectin ligand protein comprising the amino acid sequence set forth in SEQ ID NO 2 from amino acid 42 to amino acid 310, said protein being substantially free from other mammalian proteins. In another embodiment, the invention provides a composition comprising a protein comprising the amino acid sequence set forth in SEQ ID NO 4.

In yet another embodiment, the invention provides compositions comprising antibodies specific for P-selectin ligand proteins.

In another embodiment, the invention provides a method of identifying an inhibitor of P-selectin-mediated intercellular adhesion which comprises

- (a) combining a P-selectin protein with a P-selectin ligand protein comprising an amino acid sequence selected from the group consisting of the amino acid sequence set forth in SEQ ID NO 2 from amino acid 1 to amino acid 402, the amino acid sequence set forth in SEQ ID NO 2 from amino acid 42 to amino acid 402, the amino acid sequence set forth in SEQ ID NO 2 from amino acid 42 to amino acid 310, and the amino acid sequence set forth in SEQ ID NO 4, said combination forming a first binding mixture;
- (b) measuring the amount of binding between the P-selectin protein and the P-selectin ligand protein in the first binding mixture;
- (c) combining a compound with the P-selectin protein and the P-selectin ligand protein to form a second binding mixture;
- (d) measuring the amount of binding in the second binding mixture; and
- (e) comparing the amount of binding in the first binding mixture with the amount of binding in the second binding mixture;

wherein the compound is capable of inhibiting P-selectin-mediated intercellular adhesion when a decrease in the amount of binding of the second binding mixture occurs.

In another embodiment, the invention provides a method of identifying an inhibitor of E-selectin-mediated intercellular adhesion which comprises

- (a) combining a E-selectin protein with a P-selectin ligand protein comprising an amino acid sequence selected from the group consisting of the amino acid sequence set forth in SEQ ID NO 2 from amino acid 1 to amino acid 402, the amino acid sequence set forth in SEQ ID NO 2 from amino acid 42 to amino acid 402, the amino acid sequence set forth in SEQ ID NO 2 from amino acid 42 to amino acid 310, and the amino acid sequence set forth in SEQ ID NO 4, said combination forming a first binding mixture;

(c) measuring the amount of binding between the E-selectin protein and the P-selectin ligand protein in the first binding mixture;

(d) combining a compound with the E-selectin protein and the P-selectin ligand protein to form a second binding mixture;

(d') measuring the amount of binding in the second binding mixture; and

10 (e) comparing the amount of binding in the first binding mixture with the amount of binding in the second binding mixture,

wherein the compound is capable of inhibiting E-selectin-mediated intercellular adhesion when a decrease in the amount of binding of the second binding mixture occurs. These methods may also be used to look for L-selectin inhibitors by substituting L-selectin for E-selectin.

15 The invention also encompasses processes for producing P-selectin ligand proteins which comprise: (a) co-transforming a host cell with a DNA encoding a P-selectin ligand protein and a DNA encoding a fucosyltransferase capable of synthesizing sialyl Lewis X (sLe^x) or sialyl Lewis A (sLe^a) such as an $(\alpha 1,3)(\alpha 1,4)$ fucosyltransferase or an $(\alpha 1,3)$ fucosyltransferase, each of said DNAs being operably linked to an expression control sequence; (b) culturing the host cell in
20 suitable culture medium; and (c) purifying the P-selectin ligand protein from the culture medium. In certain other embodiments, the host cell is also co-transformed with a DNA encoding a paired basic amino acid converting enzyme and/or a DNA encoding a GlcNAc transferase (preferably a "core2 transferase"). In preferred embodiments, the P-selectin ligand protein is a full-length or soluble form.

25 In other embodiments, the present invention includes a P-selectin ligand protein having P-selectin ligand protein activity. In preferred embodiments, the ligand protein is a protein comprising the sequence from amino acid 42 to amino acid 60 of SEQ ID NO. 1, consisting essentially of the sequence from amino acid 42 to amino acid 60 of SEQ ID NO. 1, comprising the sequence from amino acid 42 to amino acid 88 of SEQ ID NO. 1, consisting essentially of the
30 sequence from amino acid 42 to amino acid 88 of SEQ ID NO. 1, consisting essentially of the sequence from amino acid 42 to amino acid 118 of SEQ ID NO. 1, or consisting essentially of the sequence from amino acid 42 to amino acid 159 of SEQ ID NO. 1. In other preferred embodiments, at least one of the asparagine residues at positions 65, 111 and 292 of SEQ ID NO. 1 have been deleted or replaced. Certain preferred embodiments of the ligand protein comprises
35 at least one of the tyrosine residues at positions 46, 48 and 51 of SEQ ID NO. 1. DNAs encoding these P-selectin ligand proteins, host cells transformed with such DNAs, process for producing protein by culturing such host cells, pharmaceutical compositions comprising the proteins, methods of identifying selectin binding inhibitors using the proteins, antibodies to the proteins

and methods of real time, electric mediated binding using the proteins are also provided by the invention.

In yet other embodiments the invention provides an isolated DNA encoding a fusion protein comprising: (a) a first amino acid sequence comprising amino acid 42 to amino acid 60 of SEQ ID NO:2, and (b) a second amino acid sequence derived from the sequence of a protein other than P-selectin ligand. Preferably, an expression control sequence is operably linked to the nucleotide sequence. Host cells transformed with such DNAs are also provided. The invention also provides a process for producing a fusion protein, which comprises: (a) culturing the host cell under condition suitable for expression of the fusion protein; and (b) purifying the fusion protein from the culture medium. Fusion proteins produced according to such process are also provided.

In certain preferred embodiments, the first amino acid sequence of such fusion protein comprises amino acid 42 to amino acid 402 of SEQ ID NO:2, amino acid 42 to amino acid 310 of SEQ ID NO:2, amino acid 42 to amino acid 88 of SEQ ID NO:2, amino acid 42 to amino acid 118 of SEQ ID NO:2, or amino acid 42 to amino acid 189 of SEQ ID NO:2.

In other preferred embodiments, the DNA comprises the nucleotide sequence of SEQ ID NO:35 from nucleotide 123 to nucleotide 939, the nucleotide sequence of SEQ ID NO:35, the nucleotide sequence of SEQ ID NO:37 from nucleotide 123 to nucleotide 807, the nucleotide sequence of SEQ ID NO:37, the nucleotide sequence of SEQ ID NO:39 from nucleotide 123 to nucleotide 1311, the nucleotide sequence of SEQ ID NO:39, the nucleotide sequence of SEQ ID NO:41 from nucleotide 123 to nucleotide 792, or the nucleotide sequence of SEQ ID NO:41.

The present invention also provides a fusion protein comprising: (a) a first amino acid sequence comprising amino acid 42 to amino acid 60 of SEQ ID NO:2, and (b) a second amino acid sequence derived from the sequence of a protein other than P-selectin ligand. Preferably, the first amino acid sequence comprises amino acid 42 to amino acid 402 of SEQ ID NO:2, amino acid sequence comprises amino acid 42 to amino acid 310 of SEQ ID NO:2, amino acid 42 to amino acid 88 of SEQ ID NO:2, amino acid 42 to amino acid 118 of SEQ ID NO:2, or amino acid 42 to amino acid 189 of SEQ ID NO:2.

In certain particularly preferred embodiments, the fusion protein comprises the amino acid sequence of SEQ ID NO:36 from amino acid 42 to amino acid 313, the

amino acid sequence of SEQ ID NO 36, the amino acid sequence of SEQ ID NO 38 from amino acid 42 to amino acid 264, the amino acid sequence of SEQ ID NO 38, the amino acid sequence of SEQ ID NO 40 from amino acid 42 to amino acid 437, the amino acid sequence of SEQ ID NO 40, the amino acid sequence of SEQ ID NO 42 from amino acid 42 to amino acid 264, or the amino acid sequence of SEQ ID NO 42.

In other preferred embodiments, the second amino acid sequence is linked to the C-terminus or the N-terminus of the first amino acid sequence, with or without being linked by a linking sequence.

In yet other embodiments, the second amino acid sequence is derived from a protein selected from the group consisting of an antibody, a cytokine, a growth factor, a differentiation factor, a hormone, an enzyme, a receptor or fragment thereof and a ligand. Preferably, the second amino acid sequence is derived from the sequence of an antibody from the Fc portion of an antibody, or is a mutation of a sequence derived from an antibody.

In yet further embodiments, the present invention provides for a composition comprising (a) a first peptide comprising amino acid 42 to amino acid 60 of SEQ ID NO:2, and (b) a second peptide derived from the sequence of a protein other than P-selectin ligand, wherein the first peptide and the second peptide are chemically linked by a moiety other than a peptide bond. Any P-selectin ligand protein of the invention can be used in such a composition.

BRIEF DESCRIPTION OF THE FIGURES

Fig. 1 is a graph comparing the binding of P-selectin ligand proteins expressed with and without core2.

Fig. 2 is an autoradiograph of immunoprecipitations of P-selectin ligand protein expressed with and without core2.

Fig. 3 depicts the results of flow cytometry analysis of the binding of P-selectin ligand protein (expressed with and without core2) to P-selectin/IgG chimera (LEC-1), and anti-P-selectin ligand protein monoclonal antibody (MAb 275).

Fig. 4 is an autoradiograph of proteins, including P-selectin ligand protein, which bound to P- and E-selectin/IgG chimeras.

Fig. 5 is a schematic representation of structural features of the full length P-selectin ligand protein of SEQ ID NO:2.

Fig. 6 is a schematic representation of several P-selectin ligand protein fragments constructed for the purpose of examining the role of N-linked glycosylation sites in binding of the P-selectin ligand proteins to selectins.

Fig. 7 depicts the results of experiments to determine the role of N-linked glycosylation sites in binding of the P-selectin ligand proteins to selectins.

Fig. 8 is a schematic representation of several P-selectin ligand protein fragments constructed for the purpose of examining the role of sulfated tyrosine residues in binding of the P-selectin ligand proteins to selectins.

Figs. 9-11 depicts the results of experiments to determine the role of sulfated tyrosine residues in binding of the P-selectin ligand proteins to selectins.

Fig. 12 is a schematic representation of several P-selectin ligand protein fragments constructed for the purpose of examining the effects of various deletions on the binding of the P-selectin ligand proteins to selectins.

Fig. 13 is a schematic depiction of the quantitative plate binding assay of Example 4(c).

Figs. 14-17 depict the results of experiments comparing the binding of various deleted and altered P-selectin ligand proteins to selectins.

Fig. 18 is a schematic representation of several P-selectin ligand protein fragments constructed for the purpose of examining the effects of alteration of tyrosine residues in the anionic region of the P-selectin ligand proteins on selectin binding.

Figs. 19-21 depict the results of experiments comparing the binding of various deleted and altered P-selectin ligand proteins to selectins.

Fig. 22 depicts a proposed model for binding of P-selectin ligand proteins to P- and E-selectin.

Figs. 23 and 24 depict the results of experiments comparing the binding of various deleted and altered P-selectin ligand proteins to selectins.

Some of the foregoing figures employ a convention for numbering amino acids within the depicted constructs which is different than the residue numbering employed in SEQ ID NO. 2. In the figures, residues are numbered using the first amino acid of soluble mature P-selectin ligand as a starting point. Hence, the residue numbers used in the figures are 41 less than those of SEQ ID NO. 2. For example, residue 19 in the figures corresponds to residue 60 in SEQ ID NO. 2.

Fig. 25 is an analysis of the expression products of CHO cells, already expressing 3/4 fucosyltransferase and Core2 transferase, which were transfected with psPSL.T7, ΔTM, I316 or psPSL.QC and amplified using methotrexate. Conditioned media was either analyzed directly or

not precipitated with LibS1, and then analyzed by SDS-PAGE under non-reducing and reducing conditions.

Fig. 26: SDS-PAGE separation of myeloid cell membrane proteins affinity captured by P- and E-selectin. Membrane lysates were prepared from U937 cells metabolically labeled with H-glucosamine and subjected to affinity precipitation with immobilized P- and E-selectin and control human IgG. Eluted proteins were treated with ("reduced") or without ("non-reduced") DTT prior to gel electrophoresis. Lanes: 1, affinity capture by human IgG; 2, affinity capture by P-selectin; 3, affinity capture by E-selectin.

Fig. 27: Sequential affinity capture experiments. ³H-labeled U937 lysate species were affinity captured by P- or E-selectin, eluted, and then subjected to immunoprecipitation with anti-PSGL-I antiserum Rb3443. Lanes: 1 and 2, control immunoprecipitations of fresh myeloid cell membrane lysates using pre-immune rabbit serum (lane 1) and Rb3443 (lane 2); 3-5, immunoprecipitation with Rb3443 of myeloid cell membrane lysates previously affinity captured and eluted from P-selectin (lane 3), E-selectin (lane 4), and human IgG (lane 5).

Fig. 28: Comparison of CD43 and PSGL-I content of myeloid cell membrane extracts. Labeled U937 cell extracts were immunoprecipitated with anti-PSGL-I rabbit polyclonal antibody Rb3443 or an anti-CD43 mouse MAb and then subjected to SDS-PAGE autoradiography. Lanes: 1, immunoprecipitation with control pre-immune rabbit serum; 2, immunoprecipitation with Rb3443; 3, immunoprecipitation with control isotype-matched mouse antibody; 4, immunoprecipitation with anti-CD43 antibody.

Fig. 29: COS transfection experiments. COS M6 cells transfected with plasmids encoding PSGL-I or CD43 as well as Fuc-TIII or Fuc-TVII were metabolically labeled with ³⁵S-methionine, and membranes were prepared for affinity capture experiments as described in Materials and Methods. The cDNAs employed in the transfections are indicated above the lanes. Precipitations were performed using (A) E-selectin, (B) P-selectin, and (C) anti-PSGL-I antiserum Rb3443 and anti-CD43 MAb.

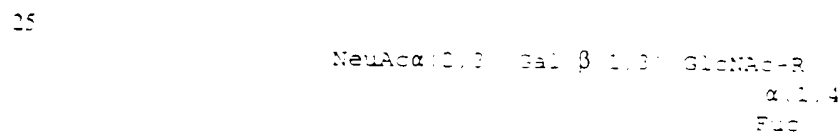
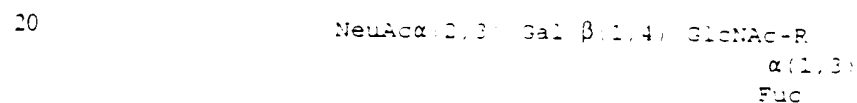
Fig. 30 summarizes the results of screening of various P-selectin ligand proteins for inhibition of P- and E-selectin binding (see Example 13).

DETAILED DESCRIPTION OF THE INVENTION

The present inventors have for the first time identified and isolated a novel DNA which encodes a protein which acts as a ligand for P-selectin on human endothelial cells and platelets. The sequence of the DNA is set forth in SEQ ID NO 1. The complete amino acid sequence of the P-selectin ligand protein (i.e., the mature peptide plus the leader sequence) is characterized by the amino acid sequence set forth in SEQ ID NO 2 from amino acid 1 to amino acid 402.

Hydroprobability analysis and comparison with known cleavage patterns predict a signal sequence of 20 to 22 amino acids, i.e. amino acids 1 to 20 or amino acids 1 to 22 of SEQ ID NO 1. The P-selectin ligand protein contains a PACE paired basic amino acid converting enzyme cleavage site (-Arg-Asp-Arg-Arg-) at amino acids 38-41 of SEQ ID NO 2. The mature P-selectin ligand protein of the present invention is characterized by the amino acid sequence set forth in SEQ ID NO 2 from amino acid 42 to amino acid 402. A soluble form of the P-selectin ligand protein is characterized by containing amino acids 21 to 310 of SEQ ID NO 2. Another soluble form of the mature P-selectin ligand protein is characterized by the amino acid sequence set forth in SEQ ID NO 2 from amino acid 42 to amino acid 310. The soluble form of the P-selectin ligand protein is further characterized by being soluble in aqueous solution at room temperature. Of course, the corresponding DNA sequences as set forth in SEQ ID NO 1 encoding these proteins are also included in the subject invention.

The P-selectin ligand of the invention is a glycoprotein which may contain one or more of the following terminal carbohydrates



where R= the remainder of the carbohydrate chain, which is covalently attached either directly to the P-selectin ligand protein or to a lipid moiety which is covalently attached to the P-selectin ligand protein. The P-selectin ligand glycoprotein of the invention may additionally be sulfated or otherwise post-translationally modified. As expressed in COS and CHO cells, full length P-selectin ligand protein (amino acids 1 to 402 of SEQ ID NO 2 or amino acids 42 to 402 of SEQ

4 SEQ ID NO 2 is a homodimeric protein having an apparent molecular weight of 120 kDa when analyzed by reducing SDS-polyacrylamide gel electrophoresis.

The structure of the full-length P-selectin ligand protein is schematically represented in Fig. 5. Three regions of the P-selectin ligand protein of SEQ ID NO 2 are: an extracellular domain (from about amino acid 21 to 310 of SEQ ID NO 2), a transmembrane domain (from
10 about amino acid 311 to 332 of SEQ ID NO 2), and an intracellular cytoplasmic domain (from about amino acid 333 to 402 of SEQ ID NO 2). The extracellular domain contains three consensus tripeptide sites (Asn-X-Ser/Thr) of potential N-linked glycosylation beginning at Asn residues 65, 111, and 292. The extracellular domain further contains three potential sites of tyrosine sulfation at residues 46, 48, and 51. The region comprised of residues 55-267 contains
15 a high percentage of proline, serine, and threonine including a subdomain of fifteen decameric repeats of the ten amino acid consensus sequence Ala-Thr-Met-Glu-Ala-Gln-Thr-Thr-X-Pro-Leu-Ala-Thr wherein X can be either Pro, Ala, Gln, Glu, or Arg. Regions such as these are characteristic of highly O-glycosylated proteins.

COS or CHO cells co-transfected with a gene encoding the P-selectin ligand protein and
20 a gene encoding fucosyltransferase (hereinafter FT), preferably an $(\alpha 1,3/\alpha 1,4)$ fucosyltransferase ("3/4FT"), are capable of binding to CHO cells expressing P-selectin on their surface, but are not capable of binding to CHO cells which do not express P-selectin on their surface. In order to bind to P-selectin, either in purified form or expressed on the surface of CHO cells, the gene encoding the P-selectin ligand protein must be co-transfected with the gene encoding an FT, since
25 transfection of either gene in the absence of the other either abolishes or substantially reduces the P-selectin binding activity. The binding of the P-selectin ligand protein of the invention to P-selectin can be inhibited by EDTA or by a neutralizing monoclonal antibody specific for P-selectin. The binding of the P-selectin ligand protein of the invention to P-selectin is not inhibited by a non-neutralizing monoclonal antibody specific for P-selectin or by an isotype control. These
30 results characterize the binding specificity of the P-selectin ligand protein of the invention.

For the purposes of the present invention, a protein is defined as having "P-selectin ligand protein activity" (i.e., variably referred to herein as a "P-selectin ligand protein" or as a "P-selectin ligand glycoprotein" or simply as a "P-selectin ligand" when it binds in a calcium-dependent
35 manner to P-selectin which is present on the surface of cells as in the CHO-P-selectin binding assay of Example 4, or to P-selectin which is affixed to another surface, for example, as the chimeric P-selectin-IgG1 protein of Example 4 is affixed to Petri dishes.

The glycosylation state of the P-selectin ligand protein of the invention was studied using a chimeric, soluble form of the P-selectin ligand protein, described in detail in Example 5(C) and

5 designated sPSL T7. The sPSL T7 protein produced from COS cells co-transfected with 3/4FT is extensively modified by post-translational glycosylation as described in detail in Example 60C. Thus, it is believed that both N- and O-linked oligosaccharide chains, at least some of which are sialated, are present on the P-selectin ligand protein of the invention.

The P-selectin ligand protein of the invention may also bind to E-selectin and L-selectin. 10 Conditioned medium from COS cells which have been co-transfected with the DNA encoding sPSL T7 or P-selectin ligand-Ig fusions and with the DNA encoding 3/4FT, when coated on wells of plastic microtiter plates, causes CHO cells which express E-selectin to bind to the plates; however CHO cells which do not express E-selectin do not bind to such plates. The binding of CHO cells which express E-selectin to microtiter plates coated with conditioned medium from 15 COS cells which have been co-transfected with the DNA encoding sPSL T7 and with the DNA encoding 3/4FT is abolished in the presence of EDTA or of a neutralizing antibody specific for E-selectin. Conditioned medium from COS cells transfected only with the sPSL T7 DNA does not cause binding of CHO cells which express E-selectin when coated on wells of microtiter plates. For these reasons, the P-selectin ligand protein of the invention is believed to be useful 20 as an inhibitor of E-selectin-mediated intercellular adhesion in addition to P-selectin-mediated intercellular adhesion.

Antibodies raised against COS-produced soluble P-selectin ligand protein are immunoreactive with the major HL-60 glycoprotein that specifically binds P-selectin as determined by affinity capture using an immobilized Fc chimera of P-selectin. U937 cells bear 25 a similar immunoreactive glycoprotein ligand. Thus, a single glycoprotein species is observed upon EDTA elution of immobilized P-selectin previously incubated with detergent extracts of H- glucosamine labeled U937 cells. This major species exhibits an apparent molecular weight by SDS-PAGE of 220 kD under non-reducing conditions and 100 kD under reducing conditions. As with the comparable species isolated from HL-60 cells, this U937 ligand is immunoreactive 30 with a polyclonal antibody raised against COS recombinant P-selectin ligand protein. In addition, affinity capture of E-selectin ligands from U937 cell and cell membrane preparations using an immobilized Fc chimera of E-selectin, yield a single major species with identical mass and electrophoretic behavior as the major U937 P-selectin ligand. Thus, E- and P-selectin recognize the same major glycoprotein ligand in U937 cells, a glycoprotein ligand immunoreactive with an 35 anti-P-selectin ligand protein antibody and possessing the same apparent mass and electrophoretic behavior as full length, recombinant P-selectin ligand protein.

Fragments of the P-selectin ligand protein which are capable of interacting with P-selectin or which are capable of inhibiting P-selectin-mediated intercellular adhesion are also encompassed

8 by the present invention. Such fragments comprise amino acids 27 to 54 of SEQ ID NO 2, a region of the P-selectin ligand protein having a low frequency of proline and threonine residue; amino acids 55 to 127 of SEQ ID NO 2, having a high frequency of proline, serine, and threonine in addition to two consensus sequences for asparagine-linked glycosylation (Asn-N-Ser-Thr); another larger fragment, amino acids 128 to 267 of SEQ ID NO 2, having both a high frequency
10 of proline, serine, and threonine and containing fifteen repeats of the following ten amino acid consensus sequence (Ala-Thr-Met)-Glu-Ala-Gln-Thr-Thr-Pro-Arg-Gln-Ala-Glu-Leu-Pro-(Ala-Thr); smaller fragments within this large fragment may also retain the capacity to interact with P-selectin or act as inhibitors of P-selectin-mediated intercellular adhesion; the region containing a consensus sequence for asparagine-linked glycosylation and comprising amino acids
15 268 to 303 of SEQ ID NO 2, the hydrophobic region of the protein represented by amino acids 309 to 333 of SEQ ID NO 2, and the amphiphilic region of the P-selectin ligand protein from amino acids 334 to 402 of SEQ ID NO 2. Additional fragments may comprise amino acid 43 to amino acid 56 of SEQ ID NO 2 or amino acid 42 to amino acid 60 of SEQ ID NO 2, with one or more sulfated or phosphorylated (Domcheck *et al.*, *Biochemistry* 31, 9865-9870 (1992)) tyrosines
20 at amino acid 46, amino acid 48, and/or amino acid 51. Fragments of the P-selectin ligand protein may be in linear form or they may be cyclized using known methods, for example, as described in H.U. Saragovi, *et al.*, *Bio/Technology* 10, 773-778 (1992) and in R.S. McDowell, *et al.*, *J. Amer. Chem. Soc.* 114, 9245-9253 (1992), both of which are incorporated herein by reference. For the purposes of the present invention, all references to "P-selectin ligand protein" herein
25 include fragments capable of binding to P-selectin.

Such fragments may be fused to carrier molecules such as immunoglobulins, to increase the valency of P-selectin ligand binding sites. For example, soluble forms of the P-selectin ligand protein such as the fragments from amino acid 42 to amino acid 295 or from amino acid 42 to amino acid 388 of SEQ ID NO 2 may be fused through "linker" sequences to the Fc portion of an
30 immunoglobulin (native sequence or mutated sequences for conferring desirable qualities, such as longer half-life or reduced immunogenicity) to the resulting chimera. For a bivalent form of the P-selectin ligand protein, such a fusion could be to the Fc portion of an IgG molecule as in Example 5(D) and in SEQ ID NO 6. Other immunoglobulin isotypes may also be used to generate such fusions. For example, a P-selectin ligand protein - IgM fusion would generate a
35 decavalent form of the P-selectin ligand protein of the invention.

Fusions of any of the P-selectin ligand proteins of the present invention to amino acid sequences derived from other proteins may also be constructed. Preferred P-selectin ligand proteins for such purpose include the fragments from amino acid 42 to amino acid 295 or from

5 amino acid 42 to amino acid 55 of SEQ ID No. 2. Desirable fusion proteins may include any amino acid sequence from proteins having a biological activity different from that of P-selectin ligand, such as, for example, cytokines, growth and differentiation factors, such as bone morphogenetic proteins (e.g., BMPs), hormones, enzymes, receptor components or fragments and other ligands. Also, P-selectin ligand protein can be chemically coupled to other proteins or
10 pharmaceutical agents. In such usage, the P-selectin ligand protein, by virtue of the ability to interact with selectin molecules, alters the pharmacokinetics and/or biodistribution of the fused or coupled agent thereby enhancing its therapeutic efficacy. For example, fusion of a P-selectin ligand protein sequence to a cytokine sequence can direct the cytokine's activity to an area of inflammation. In such instance, the P-selectin ligand protein portion of the fusion protein will
15 bind to selectins expressed at the site of inflammation. This binding will cause the cytokine portion of the fusion protein to become localized and available to bind its cognate receptor on any proximal cell surface. Other ligands could similarly be used in such fusions proteins to attract cells expressing their corresponding receptors to a site of P-selectin expression. Preferred examples of such fusions are described in Example 15.

20 In any fusion protein incorporating a P-selectin ligand protein, the amino acid sequence derived from a protein or proteins other than P-selectin ligand can be linked to either the C-terminus or N-terminus of the P-selectin ligand-derived sequence. The linkage may be direct (i.e., without an intervening linking sequence not derived from either protein) or through a linking sequence.

25 Methods of treating a mammalian subject using such fusion proteins are also contemplated by the present invention. In such instances, the fusion protein is used to treat a condition which is affected by the protein to which the P-selectin ligand protein is fused. For example, a fusion of a P-selectin ligand protein to IL-11 could be used to localize the activity of IL-11 to bone marrow endothelial cells which express selectins on their surface. Once localized,
30 the IL-11 portion of the fusion protein will stimulate megakaryocyte progenitors. Similarly, a fusion of a P-selectin ligand protein to a BMP could be used to stimulate bone or cartilage formation in an area of injury. Injured tissues express P-selectin, which will bind the fusion protein. Once localized, the BMP portion of the fusion protein will stimulate bone or cartilage production in the area of injury.

35 As detailed in the Examples below, the P-selectin ligand protein of the invention was initially obtained using an expression cloning approach (Clark *et al.*, U.S. 4,675,285). A cDNA library was constructed from the human promyelocytic cell line HL-60 (S.J. Collins, *et al.*, *Nature* 270, 347-349, 1977; ATCC No. CCL 240). This library was cotransfected into COS cells with

8 cDNA encoding a 34FT and the cotransfectants were screened for binding to a chimeric molecule consisting of the extracellular portion of P-selectin and the Fc portion of a human IgG monoclonal antibody. Cotransfectants which bound to the chimeric P-selectin were enriched for cDNAs encoding the P-selectin ligand protein. This screening process was repeated several times to enrich the plasmid population further for cDNAs encoding the P-selectin ligand protein. In a
10 second cloning stage, the enriched plasmid population was again cotransfected into COS cells with the 34FT gene and screened for binding to a fluorescently labeled CHO cell line which expressed P-selectin on the cell surface. A single cDNA clone was obtained from this approach and was designated pMT21 PL85. The pMT21 PL85 plasmid was deposited with the American Type Culture Collection on October 16, 1992 and given the accession number ATCC 69096.

15 One novel DNA of the present invention is set forth in SEQ ID NO 1. The DNA of the present invention may encode a variety of forms of the P-selectin ligand protein. For example, in one embodiment, the DNA of the invention encodes the entire P-selectin ligand protein having the amino acid sequence set forth in SEQ ID NO 2 from amino acid 1 to amino acid 402. In another embodiment, the DNA of the invention encodes a form of the P-selectin ligand protein
20 which lacks the signal sequence and which is characterized by the amino acid sequence set forth in SEQ ID NO 2 from amino acid 21 to amino acid 402. In yet another embodiment, the DNA of the invention encodes the mature P-selectin ligand protein characterized by the amino acid sequence set forth in SEQ ID NO 2 from amino acid 42 to amino acid 402. Another embodiment of the DNA of the invention encodes a soluble form of the P-selectin ligand protein characterized
25 by the amino acid sequence set forth in SEQ ID NO 2 from amino acid 1 to amino acid 310. The DNA of the invention is also embodied in a DNA encoding a soluble form of the mature P-selectin ligand protein, said protein being characterized by the amino acid sequence set forth in SEQ ID NO 2 from amino acid 42 to amino acid 310. The DNA of the invention is further embodied in a DNA sequence encoding a soluble form of the P-selectin ligand protein which
30 lacks the signal sequence, said protein being characterized by the amino acid sequence set forth in SEQ ID NO 2 from amino acid 21 to amino acid 310. The DNA of the present invention is free from association with other human DNAs and is thus characterized as an isolated DNA. As detailed above, DNAs which encode P-selectin ligand fragments which interact with P-selectin are also included in the present invention.

35 The expression of P-selectin ligand protein mRNA transcripts has been observed in a variety of human cell lines (HL-60, THP-1, U937) and in human monocytes and polymorphonuclear leukocytes by Northern analysis using a P-selectin ligand protein cDNA probe. In all of these cell lines, a major transcript of 2.5 kb was observed. A minor species of

approximately 4 kb was observed in the HL60 and U937 cell lines and in polymorphonuclear leukocytes. In contrast, no P-selectin ligand mRNA expression was detected in the human hepatoblastoma cell line HepG2.

The P-selectin ligand protein of the invention is encoded by a single copy gene and is not part of a multi-gene family, as determined by Southern blot analysis. The genomic form of the P-selectin ligand protein of the invention contains a large intron of approximately 9 kb located at nucleotide 54 in the 5' untranslated region. In polymorphonuclear leukocytes and monocytes, the P-selectin ligand protein of the invention is encoded by the DNA sequence set forth in SEQ ID NO 3. In this embodiment, the P-selectin ligand protein contains sixteen repeat regions. The isolated DNA of the invention is correspondingly also embodied in the DNA sequence set forth in SEQ ID NO 3 and is contained on plasmid pPL85R16 which was deposited with the American Type Culture Collection on October 22, 1993 and given the Accession Number ATCC 75577.

The invention also encompasses allelic variations of the isolated DNA as set forth in SEQ ID NO 1 or of the isolated DNA as set forth in SEQ ID NO 3, that is, naturally-occurring alternative forms of the isolated DNA of SEQ ID NO 1 or SEQ ID NO 3 which also encode proteins having P-selectin ligand activity. Also included in the invention are isolated DNAs which hybridize to the DNA set forth in SEQ ID NO 1 or to the DNA set forth in SEQ ID NO 3 under stringent (e.g. 4xSSC at 65°C or 50% formamide and 4xSSC at 42°C), or relaxed (4xSSC at 50°C or 30-40% formamide at 42°C) conditions, and which have P-selectin ligand protein activity. Isolated DNA sequences which encode the P-selectin ligand protein but which differ from the DNA set forth in SEQ ID NO 1 or from the DNA set forth in SEQ ID NO 3 by virtue of the degeneracy of the genetic code and which have P-selectin ligand protein activity are also encompassed by the present invention. Variations in the DNA as set forth in SEQ ID NO 1 or in the DNA as set forth in SEQ ID NO 3 which are caused by point mutations or by induced modifications which enhance the P-selectin ligand activity, half-life or production level are also included in the invention. For the purposes of the present invention all references herein to the "DNA of SEQ ID NO 1" include, in addition to DNAs comprising the specific DNA sequence set forth in SEQ ID NO 1, DNAs encoding the mature P-selectin ligand protein of SEQ ID NO 2, DNAs encoding fragments of the P-selectin ligand protein of SEQ ID NO 2 which are capable of binding to P-selectin, DNAs encoding soluble forms of the P-selectin ligand protein of SEQ ID NO 2, allelic variations of the DNA sequence of SEQ ID NO 1, DNAs which hybridize to the DNA sequence of SEQ ID NO 1 and which encode proteins having P-selectin ligand protein activity, DNAs which differ from the DNA of SEQ ID NO 1 by virtue of degeneracy of the genetic code, and the variations of the DNA sequence of SEQ ID NO 1 set forth above. Similarly,

all reference to the DNA of SEQ ID NO 3 include in addition to the specific sequence set forth in SEQ ID NO 3, DNAs encoding the mature P-selectin ligand protein of SEQ ID NO 4, DNAs encoding fragments of the P-selectin ligand protein of SEQ ID NO 4 which are capable of binding to P-selectin, DNAs encoding soluble forms of the P-selectin ligand protein of SEQ ID NO 4, allelic variations of the DNA of SEQ ID NO 3, DNAs which hybridize to the DNA sequence of SEQ ID NO 3 and which encode proteins having P-selectin ligand protein activity, DNAs which differ from the DNA of SEQ ID NO 3 by virtue of degeneracy of the genetic code, and the variations of the DNA of SEQ ID NO 3 set forth above.

A DNA encoding a soluble form of the P-selectin ligand protein may be prepared by expression of a modified DNA in which the regions encoding the transmembrane and cytoplasmic domains of the P-selectin ligand protein are deleted and/or a stop codon is introduced to the codon for the amino acid at the carboxy terminus of the extracellular domain. For example, hydrophobicity analysis predicts that the P-selectin ligand protein set forth in SEQ ID NO 2 has a transmembrane domain comprised of amino acids 311 to 332 of SEQ ID NO 2 and a cytoplasmic domain comprised of amino acids 333 to 402 of SEQ ID NO 2. A modified DNA as described above may be made by standard molecular biology techniques, including site-directed mutagenesis methods which are known in the art or by the polymerase chain reaction using appropriate oligonucleotide primers. Methods for producing several DNAs encoding various soluble P-selectin ligand proteins are set forth in Example 5.

A DNA encoding other fragments and altered forms of P-selectin ligand protein may be prepared by expression of modified DNAs in which portions of the full-length sequence have been deleted or altered. Substantial deletions of the P-selectin ligand protein sequence can be made while retaining P-selectin ligand protein activity. For example, P-selectin ligand proteins comprising the sequence from amino acid 42 to amino acid 189 of SEQ ID NO 2, the sequence from amino acid 42 to amino acid 118 of SEQ ID NO 2, or the sequence from amino acid 42 to amino acid 89 of SEQ ID NO 2 each retain the P-selectin protein binding activity and the ability to bind to E-selectin. P-selectin ligand proteins in which one or more N-linked glycosylation sites (such as those at amino acids 65, 111, and 292 of SEQ ID NO 2) have been changed to other amino acids or deleted also retain P-selectin protein binding activity and the ability to bind E-selectin. P-selectin ligand proteins comprising from amino acid 42 to amino acid 60 of SEQ ID NO 2 (which includes a highly anionic region of the protein from amino acid 45 to amino acid 58 of SEQ ID NO 2) also retain P-selectin ligand protein activity; however, P-selectin ligand proteins limited to such sequence do not bind to E-selectin. Preferably, a P-selectin ligand protein retains at least one (more preferably at least two and most preferably all three) of the tyrosine

residues found at amino acids 41, 43 and 51 of SEQ ID NO. 1, mutation of which may contribute to P-selectin ligand protein activity. Construction of DNAs encoding these and other active fragments or altered forms of P-selectin ligand protein may be accomplished in accordance with methods known to those skilled in the art.

The isolated DNA of the invention may be operably linked to an expression control sequence such as the pMT2 or pED expression vectors disclosed in Kaufman *et al.*, *Nucleic Acids Res.* 19, 4485-4490 (1991), in order to produce the P-selectin ligand recombinantly. Many suitable expression control sequences are known in the art. General methods of expressing recombinant proteins are also known and are exemplified in R. Kaufman, *Methods in Enzymology* 185, 537-566 (1990). As defined herein "operably linked" means enzymatically or chemically ligated to form a covalent bond between the isolated DNA of the invention and the expression control sequence, in such a way that the P-selectin ligand protein is expressed by a host cell which has been transformed (transfected) with the ligated DNA-expression control sequence.

Several endoproteolytic enzymes are known which cleave precursor peptides at the carboxyl side of paired amino acid sequences (e.g., -Lys-Arg- and -Arg-Arg-) to yield mature proteins. Such enzymes are generally known as paired basic amino acid converting enzymes or PACE, and their use in recombinant production of mature peptides is extensively disclosed in WO 92/09698 and U.S. Application Serial No. 07/885,972, both of which are incorporated herein by reference. The PACE family of enzymes are known to increase the efficiency of proteolytic processing of precursor polypeptides in recombinant host cells. As mentioned above, the P-selectin ligand protein of the invention contains such a PACE cleavage site.

The soluble mature P-selectin ligand protein of the present invention may be made by a host cell which contains a DNA sequence encoding any soluble P-selectin ligand protein as described herein and a DNA sequence encoding PACE as described in WO 92/09698 and U.S. Application Serial No. 07/885,972, incorporated herein by reference, or using the DNA sequence of SEQ ID NO-5. Such a host cell may contain the DNAs as the result of co-transformation or sequential transformation of separate expression vectors containing the soluble P-selectin ligand protein DNA and the PACE DNA, respectively. A third DNA which encodes a 3-4FT may also be co-transformed with the DNAs encoding the P-selectin ligand protein and PACE. Alternatively, the host cell may contain the DNAs as the result of transformation of a single expression vector containing both soluble P-selectin ligand protein DNA and PACE DNA. Construction of such expression vectors is within the level of ordinary skill in molecular biology. Methods for co-transformation and transformation are also known.

Many DNA sequences encoding PACE are known. For example, a DNA encoding the form of PACE known as *fun* is disclosed in A.M.W. van den Brule and et al., *Nucleic Acids Res.* 17, 664 (1990), incorporated herein by reference. A cDNA encoding a soluble form of PACE, known as PACE-SOL, is set forth in SEQ ID NO 5. DNAs encoding other forms of PACE also exist, and any such PACE-encoding DNA may be used to produce the soluble mature P-selectin ligand protein of the invention, so long as the PACE is capable of cleaving the P-selectin ligand protein at amino acids 38-41. Preferably, a DNA encoding a soluble form of PACE is used to produce the soluble mature P-selectin ligand protein of the present invention.

The DNAs encoding a soluble form of the P-selectin ligand protein and PACE, separately or together, may be operably linked to an expression control sequence such as those contained in the pMT2 or pED expression vectors discussed above, in order to produce the PACE-cleaved soluble P-selectin ligand recombinantly. Additional suitable expression control sequences are known in the art. Examples 3(C) and 3(D) below set forth methods for producing the soluble mature P-selectin ligand protein of the invention.

A number of types of cells may act as suitable host cells for expression of the P-selectin ligand protein. Suitable host cells are capable of attaching carbohydrate side chains characteristic of functional P-selectin ligand protein. Such capability may arise by virtue of the presence of a suitable glycosylating enzyme within the host cell, whether naturally occurring, induced by chemical mutagenesis, or through transfection of the host cell with a suitable expression plasmid containing a DNA sequence encoding the glycosylating enzyme. Host cells include, for example, monkey COS cells, Chinese Hamster Ovary (CHO) cells, human kidney 293 cells, human epidermal A-431 cells, human Colo205 cells, 3T3 cells, CV-1 cells, other transformed primate cell lines, normal diploid cells, cell strains derived from *in vitro* culture of primary tissue, primary explants, HeLa cells, mouse L cells, BHK, HL-60, U937, or HaK cells.

The P-selectin ligand protein may also be produced by operably linking the isolated DNA of the invention and one or more DNAs encoding suitable glycosylating enzymes to suitable control sequences in one or more insect expression vectors, and employing an insect expression system. Materials and methods for baculovirus/insect cell expression systems are commercially available in kit form from, e.g., Invitrogen, San Diego, California, U.S.A. (the *MaxBac®* kit), and such methods are well known in the art, as described in Summers and Smith, Texas Agricultural Experiment Station Bulletin No. 1555 (1987), incorporated herein by reference. Soluble forms of the P-selectin ligand protein may also be produced in insect cells using appropriate isolated DNAs as described above. A DNA encoding a form of PACE may further be co-expressed in an insect host cell to produce a PACE-cleaved form of the P-selectin ligand protein.

5 Alternatively, it may be possible to produce the P-selectin ligand protein in either eukaryotes such as yeast or in prokaryotes such as bacteria. Potentially suitable yeast strains include *Saccharomyces cerevisiae*, *Schizosaccharomyces pombe*, *Kluyveromyces* strains, *Candida*, or any yeast strain capable of expressing heterologous proteins. Potentially suitable bacterial strains include *Escherichia coli*, *Bacillus subtilis*, *Salmonella typhimurium*, or any bacterial strain
10 capable of expressing heterologous proteins. If the P-selectin ligand protein is made in yeast or bacteria, it is necessary to attach the appropriate carbohydrates to the appropriate sites on the protein moiety covalently, in order to obtain the glycosylated P-selectin ligand protein. Such covalent attachments may be accomplished using known chemical or enzymatic methods.

The P-selectin ligand protein of the invention may also be expressed as a product of
15 transgenic animals, e.g., as a component of the milk of transgenic cows, goats, pigs, or sheep which are characterized by somatic or germ cells containing a DNA sequence encoding the P-selectin ligand protein.

The P-selectin binding activity of a P-selectin protein may be enhanced by co-
transformation of a host cell with a GlcNAc transferase, preferably UDP-GlcNAc:Gal β 1-
20 3GalNAc-R(GlcNAc to GalNAc) β 1-6 GlcNAc transferase (EC 2.4.1.102), also known as "core2 transferase."

O-linked glycans present on P-selectin ligand protein have been shown to be important
for binding to P-selectin (D. Sako *et al.*, Cell 75, 1179-1186 (1993)). It has been reported that sialyl Le^x on O-linked glycans of myeloid cells are presented on complex, branched structures
25 (Maemura, K. and Fukuda, M., J. Biol. Chem. 267, 24379-24386 (1992)). The enzyme responsible for generating such oligosacchande structures is "core2". The core2 enzyme activity is found at very low levels in COS cells and at trace levels in CHO cells. Host cells co-
transformed with DNAs encoding a P-selectin ligand protein, an α 1,3 α 1,4 fucosyltransferase and core2 can produce P-selectin ligand protein exhibiting 20-30 fold enhanced binding to P-
30 selectin.

In certain preferred embodiments, P-selectin ligand protein is produced by co-transfecting a host cell with DNAs encoding soluble P-selectin ligand protein, 3-4FT, core2 and PACE.

The P-selectin ligand protein of the invention may be prepared by culturing transformed host cells under culture conditions necessary to express a P-selectin binding glycoprotein. The
35 resulting expressed glycoprotein may then be purified from culture medium or cell extracts. Soluble forms of the P-selectin ligand protein of the invention can be purified by affinity chromatography over Lentil lectin-Sepharose® and subsequent elution with 0.5M α -methylmannoside. The eluted soluble P-selectin ligand protein can then be further purified and

concentrated by 10,000 ammonium sulfate precipitation step. The protein concentrate is resuspended, and further purified by size exclusion chromatography (e.g., TSK T4 6R SW). Alternatively, full length forms of the P-selectin ligand protein of the invention can be purified by preparing a total membrane fraction from the expressing cell and extracting the membranes with a non-ionic detergent such as Triton X-100. The detergent extract can then be passed over an affinity column comprised of immobilized P-selectin, and the P-selectin ligand protein can be eluted from the column with 10mM EDTA in a buffer containing 0.1% detergent. The material eluted from the affinity column can then be dialyzed to remove EDTA and further purified over a Lentil lectin-Sepharose 6 affinity column, again eluting with 0.5M α -methyl-L-mannoside.

Alternatively, the P-selectin ligand protein of the invention is concentrated using a commercially available protein concentration filter, for example, an Amicon or Millipore Pellicon ultrafiltration unit. Following the concentration step, the concentrate can be applied to a purification matrix such as a gel filtration medium. Alternatively, an ion exchange resin can be employed, for example, a matrix or substrate having pendant diethylaminoethyl (DEAE) groups. The matrices can be acrylamide, agarose, dextran, cellulose or other types commonly employed in protein purification. Alternatively, a cation exchange step can be employed. Suitable cation exchangers include various insoluble matrices comprising sulfopropyl or carboxymethyl groups. Sulfopropyl groups are preferred (e.g., S-Sepharose® columns). The purification of the P-selectin ligand protein from culture supernatant may also include one or more column steps over such affinity resins as concanavalin A-agarose, heparin-toyopearl® or Cibacrom blue 3GA Sepharose® or by hydrophobic interaction chromatography using such resins as phenyl ether, butyl ether, or propyl ether, or by immunaffinity chromatography.

Finally, one or more reverse-phase high performance liquid chromatography (RP-HPLC) steps employing hydrophobic RP-HPLC media, e.g., silica gel having pendant methyl or other aliphatic groups, can be employed to further purify the P-selectin ligand protein. Some or all of the foregoing purification steps, in various combinations, can also be employed to provide a substantially homogeneous isolated recombinant protein. The P-selectin ligand protein thus purified is substantially free of other mammalian proteins and is defined in accordance with the present invention as "isolated P-selectin ligand protein".

Isolated P-selectin ligand protein may be useful in treating conditions characterized by P-, E- or L-selectin mediated intercellular adhesion. Such conditions include, without limitation, myocardial infarction, bacterial or viral infection, metastatic conditions, inflammatory disorders such as arthritis, gout, uveitis, acute respiratory distress syndrome, asthma, emphysema, delayed type hypersensitivity reaction, systemic lupus erythematosus, thermal injury, such as burns or

4 translute, autoimmune thyroiditis, experimental allergic encephalomyelitis, multiple sclerosis, multiple organ injury syndrome secondary to trauma, diabetes, Raynaud's syndrome, neutrophilic dermatosis, Sweet's syndrome, inflammatory bowel disease, Graves' disease, glomerulonephritis, gingivitis, periodontitis, hemolytic-uremic syndrome, ulcerative colitis, Crohn's disease, necrotizing enterocolitis, granulocyte transfusion associated syndrome, cytokine-induced toxicity, and the like. Isolated P-selectin ligand protein may also be useful in organ transplantation, both
10 to prepare organs for transplantation and to quell organ transplant rejection. Accordingly, P-selectin ligand protein may be administered to a living or non-living organ donor, prior to organ removal. In addition, P-selectin ligand protein may be administered *ex-vivo* to the donor organ concomitantly with organ preservation solution, prior to, and/or subsequent to surgical
15 anastomosis with the recipient. Isolated P-selectin ligand protein may be used to treat hemodialysis and leukophoresis patients. Additionally, isolated P-selectin ligand protein may be used as an antimetastatic agent. Isolated P-selectin ligand protein may be used itself as an inhibitor of P-, E- or L-selectin-mediated intercellular adhesion or to design inhibitors of P-, E- or L-selectin-mediated intercellular adhesion. The present invention encompasses both
20 pharmaceutical compositions containing isolated P-selectin ligand protein and therapeutic methods of treatment or use which employ isolated P-selectin ligand protein.

Isolated P-selectin ligand protein, purified from cells or recombinantly produced, may be used as a pharmaceutical composition when combined with a pharmaceutically acceptable carrier. Such a composition may contain, in addition to P-selectin ligand protein and carrier, diluents,
25 fillers, salts, buffers, stabilizers, solubilizers, and other materials well known in the art. The term "pharmaceutically acceptable" means a non-toxic material that does not interfere with the effectiveness of the biological activity of the active ingredient(s). The characteristics of the carrier will depend on the route of administration. The pharmaceutical composition of the invention may also contain cytokines, lymphokines, or other hematopoietic factors such as M-CSF, GM-CSF,
30 IL-1, IL-2, IL-3, IL-4, IL-5, IL-6, IL-7, IL-8, IL-9, IL-10, IL-11, IL-12, G-CSF, Meg-CSF, stem cell factor, and erythropoietin. The pharmaceutical composition may contain thrombolytic or anti-thrombotic factors such as plasminogen activator and Factor VIII. The pharmaceutical composition may further contain other anti-inflammatory agents. Such additional factors and/or
35 agents may be included in the pharmaceutical composition to produce a synergistic effect with isolated P-selectin ligand protein, or to minimize side effects caused by the isolated P-selectin ligand protein. Conversely, isolated P-selectin ligand protein may be included in formulations of the particular cytokine, lymphokine, other hematopoietic factor, thrombolytic or anti-thrombotic factor, or anti-inflammatory agent to minimize side effects of the cytokine,

4 lymphokines, other hematopoietic factors, thrombolytic or anti-thrombotic factors, or anti-inflammatory agent.

The pharmaceutical composition of the invention may be in the form of a liposome in which isolated P-selectin ligand protein is combined, in addition to other pharmaceutically acceptable carriers, with amphipathic agents such as lipids which exist in aggregated form as
10 micelles, insoluble monolayers, liquid crystals, or lamellar layers which in aqueous solution. Suitable lipids for liposomal formulation include, without limitation, monoglycerides, diglycerides, sulfatides, lysolecithin, phospholipids, saponin, bile acids, and the like. Preparation of such liposomal formulations is within the level of skill in the art, as disclosed, for example, in U.S. Patent No. 4,235,371, U.S. Patent No. 4,501,728, U.S. Patent No. 4,837,028, and U.S.
15 Patent No. 4,737,323, all of which are incorporated herein by reference.

As used herein, the term "therapeutically effective amount" means the total amount of each active component of the pharmaceutical composition or method that is sufficient to show a meaningful patient benefit, i.e., healing of chronic conditions characterized by P-selectin- or E-selectin-mediated cellular adhesion or increase in rate of healing of such conditions. When
20 applied to an individual active ingredient, administered alone, the term refers to that ingredient alone. When applied to a combination, the term refers to combined amounts of the active ingredients that result in the therapeutic effect, whether administered in combination, serially or simultaneously.

In practicing the method of treatment or use of the present invention, a therapeutically
25 effective amount of isolated P-selectin ligand protein is administered to a mammal having a P-selectin-mediated disease state. Isolated P-selectin ligand protein may be administered in accordance with the method of the invention either alone or in combination with other therapies such as treatments employing receptor antagonists, ligand antagonists, cytokines, lymphokines, or other hematopoietic factors. When co-administered with one or more cytokines, lymphokines, or other hematopoietic factors, isolated P-selectin ligand protein may be administered either
30 simultaneously with the cytokine(s), lymphokine(s), other hematopoietic factor(s), thrombolytic or anti-thrombotic factors, or sequentially. If administered sequentially, the attending physician will decide on the appropriate sequence of administering isolated P-selectin ligand protein in combination with cytokine(s), lymphokine(s), other hematopoietic factor(s), thrombolytic or anti-thrombotic factors.
35

Administration of isolated P-selectin ligand protein used in the pharmaceutical composition or to practice the method of the present invention can be carried out in a variety of

5 conventional ways, such as, oral ingestion, inhalation, or cutaneous or subcutaneous or intravenous injection. Intravenous administration to the patient is preferred.

When a therapeutically effective amount of isolated P-selectin ligand protein is administered orally, isolated P-selectin ligand protein will be in the form of a tablet, capsule, powder, solution or elixir. When administered in tablet form, the pharmaceutical composition of
10 the invention may additionally contain a solid carrier such as a gelatin or an adjuvant. The tablet, capsule, and powder contain from about 5 to 95% isolated P-selectin ligand protein, and preferably from about 25 to 90% isolated P-selectin ligand protein. When administered in liquid form, a liquid carrier such as water, petroleum, oils of animal or plant origin such as peanut oil, mineral oil, soybean oil, or sesame oil, or synthetic oils may be added. The liquid form of the
15 pharmaceutical composition may further contain physiological saline solution, dextrose or other saccharide solution, or glycols such as ethylene glycol, propylene glycol or polyethylene glycol. When administered in liquid form, the pharmaceutical composition contains from about 0.5 to 90% by weight of isolated P-selectin ligand protein and preferably from about 1 to 50% isolated P-selectin ligand protein.

20 When a therapeutically effective amount of isolated P-selectin ligand protein is administered by intravenous, cutaneous or subcutaneous injection, isolated P-selectin ligand protein will be in the form of a pyrogen-free, parenterally acceptable aqueous solution. The preparation of such parenterally acceptable protein solutions, having due regard to pH, isotonicity, stability, and the like, is within the skill in the art. A preferred pharmaceutical composition for
25 intravenous, cutaneous, or subcutaneous injection should contain, in addition to isolated P-selectin ligand protein an isotonic vehicle such as Sodium Chloride Injection, Ringer's Injection, Dextrose Injection, Dextrose and Sodium Chloride Injection, Lactated Ringer's Injection, or other vehicle as known in the art. The pharmaceutical composition of the present invention may also contain stabilizers, preservatives, buffers, antioxidants, or other additive known to those of skill in the art.

30 The amount of isolated P-selectin ligand protein in the pharmaceutical composition of the present invention will depend upon the nature and severity of the condition being treated, and on the nature of prior treatments which the patient has undergone. Ultimately, the attending physician will decide the amount of isolated P-selectin ligand protein with which to treat each individual patient. Initially, the attending physician will administer low doses of isolated P-
35 selectin ligand protein and observe the patient's response. Larger doses of isolated P-selectin ligand protein may be administered until the optimal therapeutic effect is obtained for the patient, and at that point the dosage is not increased further. It is contemplated that the various

5 pharmaceutical compositions used to practice the method of the present invention should contain about 1 μ g to about 100 mg of isolated P-selectin ligand protein per kg body weight.

The duration of intravenous therapy using the pharmaceutical composition of the present invention will vary, depending on the severity of the disease being treated and the condition and potential idiosyncratic response of each individual patient. It is contemplated that the duration
10 of each application of the isolated P-selectin ligand protein will be in the range of 12 to 24 hours of continuous intravenous administration. Ultimately the attending physician will decide on the appropriate duration of intravenous therapy using the pharmaceutical composition of the present invention.

Isolated P-selectin ligand protein of the invention may also be used to immunize animals
15 to obtain polyclonal and monoclonal antibodies which specifically react with the P-selectin ligand protein and which may inhibit P-selectin-mediated cellular adhesion. Such antibodies may be obtained using the entire P-selectin ligand protein as an immunogen, or by using fragments of P-selectin ligand protein such as the soluble mature P-selectin ligand protein. Smaller fragments of the P-selectin ligand protein may also be used to immunize animals, such as the fragments set
20 forth below: amino acid 42 to amino acid 56 of SEQ ID NO:2 and amino acid 127 to amino acid 138 of SEQ ID NO:2. An additional peptide immunogen comprises amino acid 238 to amino acid 248 of SEQ ID NO:2, with an alanine residue added to the amino terminus of the peptide. Another peptide immunogen comprises amino acid 43 to amino acid 56 of SEQ ID NO:2 having a sulfated tyrosine in any or all of positions 46, 48 or 51. The peptide immunogens additionally
25 may contain a cysteine residue at the carboxyl terminus, and are conjugated to a hapten such as keyhole limpet hemocyanin (KLH). Additional peptide immunogens may be generated by replacing tyrosine residues with sulfated tyrosine residues. Methods for synthesizing such peptides are known in the art, for example, as in R.P. Merrifield, *J. Amer. Chem. Soc.* 85, 2149-2154 (1963); J.L. Krstenansky, *et al.*, *FEBS Lett.* 211, 10 (1987).

30 Monoclonal antibodies binding to P-selectin ligand glycoprotein or to complex carbohydrate moieties characteristic of the P-selectin ligand glycoprotein may be useful diagnostic agents for the immunodetection of inflammatory diseases and some forms of cancer. Some cancerous cells, such as small cell lung carcinomas, may express detectable levels of the P-selectin ligand protein. This abnormal expression of the P-selectin ligand protein by cancer cells may play
35 a role in the metastasis of these cells.

Neutralizing monoclonal antibodies binding to P-selectin ligand glycoprotein or to complex carbohydrates characteristic of P-selectin ligand glycoprotein may also be useful therapeutics for both inflammatory diseases and also in the treatment of some forms of cancer.

where abnormal expression of P-selectin ligand protein is involved. These neutralizing monoclonal antibodies are capable of blocking the selectin mediated intercellular adherence function of the P-selectin ligand protein. By blocking the binding of P-selectin ligand protein, the adherence of leukocytes to sites of inappropriate inflammation is either abolished or markedly reduced. In the case of cancerous cells or leukemic cells, neutralizing monoclonal antibodies against P-selectin ligand protein may be useful in detecting and preventing the metastatic spread of the cancerous cells which may be mediated by the P-selectin ligand protein. In addition, the monoclonal antibodies bound to these cells may target the cancerous cells for antibody-dependent cell mediated cytotoxicity (ADCC), thus helping to eliminate the cancerous cells. Human antibodies which react with the P-selectin ligand protein may be produced in transgenic animals which contain human immunoglobulin encoding genes in their germ lines. Example 7 below sets forth production of a rabbit polyclonal antibody specific P-selectin ligand protein fragments.

P-selectin ligand protein of the invention may also be used to screen for agents which are capable of binding to P-selectin ligand protein and thus may act as inhibitors of P-selectin- or E-selectin-mediated intercellular adhesion. Binding assays using a desired binding protein, immobilized or not, are well known in the art and may be used for this purpose using the P-selectin ligand protein of the invention. Appropriate screening assays may be cell-based, as in Examples 3 and 9 below. Alternatively, purified protein based screening assays may be used to identify such agents. For example, P-selectin ligand protein may be immobilized in purified form on a carrier and binding to purified P-selectin may be measured in the presence and in the absence of potential inhibiting agents. A suitable binding assay may alternatively employ purified P-selectin immobilized on a carrier, with a soluble form of P-selectin ligand protein of the invention.

Any P-selectin ligand protein may be used in the screening assays described above. For example, the full-length P-selectin ligand protein set forth in SEQ ID NO 2 from amino acid 1 to amino acid 402 may be used to screen for inhibitors; or the mature P-selectin ligand protein set forth in SEQ ID NO 2 from amino acid 42 to amino acid 402 may be used to screen for inhibitors, or the soluble mature P-selectin ligand protein set forth in SEQ ID NO 2 from amino acid 42 to amino acid 310 may be used to screen for inhibitors. Alternatively, the P-selectin ligand protein of SEQ ID NO 4 from amino acid 1 to amino acid 412, or a mature form of the P-selectin ligand protein as set forth in SEQ ID NO 4 from amino acid 42 to amino acid 412, or a soluble mature form of the P-selectin ligand protein set forth in SEQ ID NO 4 from amino acid 42 to amino acid 320 may be used to screen for inhibitors of intercellular adhesion in accordance with the present invention.

5 In such a screening assay, a first binding mixture is formed by combining P-selectin or E-selectin and P-selectin ligand protein, and the amount of binding in the first binding mixture (B_1) is measured. A second binding mixture is also formed by combining P-, E- or L-selectin, P-selectin ligand protein, and the compound or agent to be screened, and the amount of binding in the second binding mixture (B_2) is measured. The amounts of binding in the first and second
10 binding mixtures are compared, for example, by performing a B_2/B_1 calculation. A compound or agent is considered to be capable of inhibiting P-, E- or L-selectin mediated intercellular adhesion if a decrease in binding in the second binding mixture as compared to the first binding mixture is observed. The formulation and optimization of binding mixtures is within the level of skill in the art; such binding mixtures may also contain buffers and salts necessary to enhance or to
15 optimize binding, and additional control assays may be included in the screening assay of the invention.

Compounds found to reduce by at least about 10%, preferably greater than about 50% or more of the binding activity of P-selectin ligand protein to P-, E- or L-selectin may thus be identified and then secondarily screened in other selectin binding assays, including assays binding to L-selectin and *in vivo* assays. By these means compounds having inhibitory activity for selectin-mediated intercellular adhesion which may be suitable as anti-inflammatory agents may be identified.

EXAMPLE 1

25 CLONING OF THE P-SELECTIN LIGAND PROTEIN GENE

A. Construction of the HL60 cDNA library

An HL60 cDNA library was constructed for expression cloning the P-selectin ligand. PolyA⁺ RNA was isolated from total RNA from the human promyelocytic cell line HL60 (S. J. Collins, *et al.*, *supra*) using a Fast Track mRNA Isolation Kit (Invitrogen, San Diego, CA). Double stranded cDNA was synthesized from the polyA⁺ RNA fraction and blunt-end ligated with EcoRI adaptors (5'-AATTGGGTCGACTGTAGAG-3', SEQ ID NO 7, 5'-CTGTAGACTTGAAGG-3', SEQ ID NO 8). The cDNA was ligated into the expression vector pMT21 (R. Kaufman *et al.*, J. Mol. Cell. Biol. 9, 946-958 (1989) that had been incubated sequentially with EcoRI endonuclease and calf intestinal alkaline phosphatase and gel purified. The ligation product was
35 electroporated in 2 μ l aliquots into competent *E. coli* DH5 α cells and grown in 1 ml of SOB medium (J. Sambrook *et al.*, Molecular Cloning: A Laboratory Manual, New York, Cold Spring Harbor Laboratory Press, p190 (1989)) which has been supplemented with 10 mM MgCl₂, 10

5 mM MgSO₄ and 20% glycerol for one hour at 37°C. In order to divide the library into smaller subsets, an aliquot from each ml. of bacterial suspension was plated onto agar plates in the presence of ampicillin, and the number of colonies per ml was calculated. Assuming that each colony represented one cDNA clone, 600,000 clones were generated and divided into subsets of approximately 16,000 clones per pool. Each of the 38 pools were grown overnight in LB broth in the presence of ampicillin and the plasmids were purified over a CsCl gradient.

B. Screening for the P-selectin ligand protein gene

In the first stage, the LEC-γ1 binding assay of Example 4(A) was utilized to pan the HL60 cDNA library and thereby to enrich for the plasmid of interest. Six μg of each HL60 cDNA library pool was co-transfected with 2 μg of α 3/4FT gene (Example 2) into COS cells. Approximately 45 hours post-transfection, the COS cells were lifted from the plates by incubating the cells in 1 mM EGTA for 15 min. at 37°C, followed by scraping with cell lifters. The cells were washed twice in Hanks buffered saline solution containing 1 mM calcium (HBSS). The cells were resuspended in 4 ml of HBSS. The resuspended transfected COS cells were screened using the LEC-γ1 binding assay described in Example 4(A).

The plasmids from adherent COS cells were recovered from a Hirt's extract [B. Hirt, J. Mol. Biol., 26, 365-369 (1967)] and then electroporated into *E. coli* DH5α cells for amplification. The enriched population of plasmids was purified over a CsCl gradient and re-transfected along with the 3/4FT gene (Example 2) into COS cells. The transfection, screening, and plasmid amplification process was repeated for a total of three times before a pool that bound to the LEC-γ1-coated plates was visually detected. The positive plasmid pool was subsequently broken down into subsets. This involved electroporating the Hirt's extract from the positive pool into *E. coli* DH5α cells and quantitating colonies per ml as described above. Various pool sizes were produced by plating out a predetermined number of colonies on agar plates in the presence of ampicillin. Duplicate plates were prepared by performing nitrocellulose lifts and storing the filters on new agar plates. The duplicate plates served as reference plates for selecting individual or groups of colonies from any pool identified as being positive.

In the second stage of cloning, COS cells were co-transfected with the sublibrary pools and the 3/4FT gene by the same procedure used in the initial steps of screening. Forty-eight hours post-transfection, the transfected cells were screened using the fluorescent CHO.P-selectin assay of Example 4(B). Positive pools were further subdivided, as described above, until finally individual colonies were screened and positive clones identified. Using this method, a single positive clone, pMT21 PL35, was found to encode the P-selectin ligand protein. The DNA

5 sequence of the P-selectin ligand contained in pMT21-PLS5 is set forth in SEQ. ID NO. 8 and the binding characteristics of the P-selectin ligand protein encoded by pMT21-PLS5 are set forth in Example 4C below.

10 EXAMPLE 2

CLONING THE α 1,3/1,4 FUCOSYLTRANSFERASE GENE

The α 1,3/1,4 fucosyltransferase gene (3/4FT) was cloned from total human genomic DNA (Clontech Laboratories) by means of PCR. The sense oligonucleotide primer contained an XbaI site and the 5' terminus of the gene (5'-
15 TACCATACGGTCTAGAGCATGGATCCCGTGGGTGCAGCCCAAGC-3', SEQ. ID NO-9), and the antisense oligonucleotide primer contained an EcoRI site and the 3' terminus of the gene (5'-
CCGGAATTCTCAGGTGAACCAAGCCGC-3', SEQ. ID NO-10). The PCR product was sequentially digested with XbaI and EcoRI and purified by standard gel purification methods. This gene was then ligated with vector pMT3Sv2ADA (R. Kaufman, Methods in Enzymology, *supra*) that had
20 also been sequentially digested with XbaI and EcoRI and purified by standard gel purification methods. Competent HB101 cells (Biorad) were transformed with this ligation product and then plated on agar plates in the presence of ampicillin. Nitrocellulose filter lifts of ampicillin-resistant transformants were probed with a radiolabelled oligonucleotide (5'-
AAGTATCTGTCCAGGGCTTCAGGT-3', SEQ. ID NO-11) complementary to the nucleotide region
25 506-530 in the middle of the gene (J. Sambrook *et al.*, *supra*).

Plasmid DNA minipreps were prepared from twelve positive clones. The purified DNA was then digested with EcoRI and XbaI to identify the correct clone with the proper size insert. This clone (pEA.3/4FT) was then grown up large scale and the DNA isolated by CsCl density gradient banding (J. Sambrook *et al.*, *supra*). DNA sequencing confirmed the identity of the
30 3/4FT gene. The functionality of the gene was assessed in a cell-cell binding assay as follows. COS-1 monkey cells (clone M6; M. Horwitz *et al.*, Mol. Appl. Genet., 2:147-149, 1983) were transfected with 3/4FT using DEAE dextran followed by DMSO shock treatment and chloroquine incubation (L. Sompeyrac and K. Dana, Proc. Natl. Acad. Sci., 78:7575-7578 (1981); M. Lopata *et al.*, Nucleic Acids Res., 12:5707-5717, (1984); H. Luthman and G. Magnuson, Nucleic Acids
35 Res., 11:1295-1308, (1983)). The transfected COS cells were suspended and quantitated for binding to a CHO line expressing E-selectin (G. Larsen *et al.*, J. Biol. Chem. 267:11104-11110, (1992)). This assay confirmed that the COS cells transfected with 3/4FT can express the sialylated Lewis^x epitope on the cell surface.

EXAMPLE 3

EXPRESSION OF THE P-SELECTIN LIGAND PROTEIN

A. Expression of the P-selectin Ligand in LEC11 cells

10 Functional P-selectin ligand was expressed in the SLe^x-positive Chinese hamster ovary (CHO) cell line LEC11 (Campbell, C. and Stanley, P Cell 35:303-309, 1983) as follows: approximately 8 µg of plasmid containing the P-selectin ligand gene (pMT21-PL85, Example 1) was transfected into LEC11 cells. At 68 hours post-transfection, the cells were treated with 2.5 mM sodium butyrate for 4 hours. The cells were observed to induce P-selectin adhesion, as
15 determined using the 6-CFD labeled CHO P-selectin cell binding assay (described in Example 4, section B). In contrast, neither LEC11 cells alone nor LEC11 cells transfected with a control plasmid induced P-selectin adhesion.

B. Expression of Soluble P-Selectin Ligand in COS cells

20 COS cells were transfected with 8 µg pED.sPSL.T7 (see Example 5C) and 4 µg pEA.3/4 FT plasmid of Example 2, 8 µg pED.sPSL.T7 alone, or 8 µg plasmid vector (pMT21) and 4 µg pEA.3/4 FT gene. Forty-five hr post-transfection, the cells were rinsed twice in PBS and incubated overnight at 37°C in serum-free DMEM minus phenol red (JRH Biosciences) supplemented with 2 mM L-glutamine, 100 U/ml penicillin and 100 µg/ml streptomycin.
25 Phenylmethylsulfonyl fluoride, aprotinin and NaN₃ were added to final concentrations of 1mM, 2 µg/ml and 0.02%, respectively, and the conditioned medium was centrifuged to remove all debris.

For immunoprecipitation experiments, the labeled soluble P-selectin ligand protein was produced by co-transfecting COS cells with pED.sPSL.T7 and pEA.3/4 FT. At forty-five hr post-transfection, the COS cells were labeled with 250 µCi/ml ³⁵S methionine (NEN) for 5 hours and the medium was collected. Expression of sPSL.T7 protein was confirmed by immunoprecipitation with anti-T7 antibodies.

C. Expression of PACE-cleaved P-selectin ligand in COS Cells

35 COS cells were co-transfected with the pED.sPSL.T7 plasmid of Example 5C, the pEA.3/4FT cDNA of Example 2, and a plasmid containing the PACE cDNA as set forth in SEQ ID NO 5. A parallel control co-transfection was done using only the pED.sPSL.T7 plasmid and the pEA.3/4FT plasmid. After 45 hours, conditioned medium from these transfected COS cells

was coated onto plastic dishes and binding to CHO P-selectin cells (Example 4) was determined. An approximately two-fold increase in bound CHO P-selectin cells was observed for dishes coated with medium containing the P-selectin ligand co-expressed with PACE, as compared with medium containing P-selectin ligand which had not been co-expressed with PACE. Amino acid sequencing of the N-terminus of purified sPSL T7 protein from the PACE co-transfection showed that all of the ligand had been cleaved at the PACE consensus site (amino acids 38-41 of SEQ ID NO 1). Radiolabelling of co-transfected COS cells with ³⁵S-methionine and subsequent SDS-polyacrylamide gel electrophoresis and autoradiography showed that comparable quantities of the P-selectin ligand had been secreted in both co-transfections.

15 D. Expression of the P-selectin Ligand Protein in CHO Cells

A full-length form (amino acids 1-402) of the P-selectin ligand protein was expressed in the CHO(DUKX) cell line (Urlaub & Chasin, Proc. Natl. Acad. Sci. USA 77: 4216-4220 (1980)) as follows: approximately 25 µg of the pMT21-PL85 plasmid and approximately 5 µg of the pED.3/4FT (produced by restriction of pEA.3/4FT with EcoRI and XbaI and insertion of the resulting fragment into the pED plasmid) were co-transfected into CHO(DUKX) cells using the calcium phosphate method. Transfectants were selected for resistance to methotrexate. After two weeks, individual colonies were screened for SLe^x expression by using a conjugate of an anti SLe^x antibody (CSLEX-1, U.S. 4,752,569) and sheep red blood cells (sRBC) prepared by the chromic chloride method (Goding, J. W., J. Immunol. Methods 10: 61-66 (1976)) as follows: sRBC were washed with 0.15M NaCl until the wash became clear and then a 50% suspension of sRBC was prepared in 0.15M NaCl. One ml of 0.01% chromic chloride solution was added dropwise while vortexing to 0.2 ml of a sRBC suspension containing 50 µg of CSLEX-1. After incubating at 37°C for 30 minutes, 10 ml of phosphate buffered saline (PBS) solution was added to the reaction. The conjugate was washed once before resuspending into 10 ml of PBS. The plates containing transfectants were washed with PBS and then 3 ml of PBS and one ml of the sRBC/CSLEX-1 conjugate was added to each plate. Positive colonies were red on a transilluminator and were picked into alpha medium with 10% fetal bovine serum. After two weeks, colonies were subjected to stepwise amplification using methotrexate at concentrations of 2, 10, 25, 100, 250 nM. The stable cell line obtained was designated CD-PSGL-1 (R3-4). Expression of the P-selectin ligand protein was confirmed by immunoprecipitation studies using the polyclonal anti-P-selectin ligand protein antibody of Example 7(A). The functionality of the P-selectin ligand protein produced by the CD-PSGL-1 (R3-4) cell line was tested by assaying the transfectants for binding to LEC-7a as in Example 4(A).

5 The sPSL-TT protein was expressed in a stable CHO-PACE line which was already expressing the cDNA encoding PACE as set forth in SEQ ID NO 5 under adenovine deamivase selection (Kaufman, *et al.*, PNAS, USA, 83 3136-3140, 1986). The bsPSL-TT (25 µg) and pED 3/4FT (5 µg) plasmids were cotransfected into CHO-PACE cells using the calcium phosphate method. Transfectants were selected for resistance to methotrexate, and individual
10 colonies which bound to the sRBC/CSLEX-1 conjugate were picked. After two weeks in culture, the colonies were subjected to stepwise amplification as described above. The stable cell line obtained was designated CP/PSL-TT (R4.1). Expression of sPSL-TT protein was confirmed by standard immunoprecipitation methods using either a TT specific monoclonal antibody or the LEC-v1 chimera of Example 4(A). In a similar fashion, a stable cell line expressing the mature
15 full length form (amino acids 42-402) of the P-selectin ligand protein was obtained by cotransfection of pMT21-PL35 and pED 3/4FT into the CHO-PACE line.

Stable cell lines expressing the sPSL-Q protein of Example 5(B) and the sPSL-Fc protein of Example 5(D) were constructed as follows: plasmids pED-sPSL-Q (25 µg) or pED-sPSL-Fc (25 µg) were cotransfected with approximately 25 µg of the pED 3/4FT plasmid described above
20 and approximately 20 µg of a plasmid containing the PACE cDNA as set forth in SEQ ID NO 5, as well as the neomycin resistance gene into CHO(DUKX) cells using the calcium phosphate method. Transfectants were selected for resistance to methotrexate and the G418 antibiotic. Approximately two weeks later, individual colonies were screened for SLe^x expression using sRBC/CSLEX-1 conjugate binding. The positive colonies were picked in G418 medium at 1
25 mg/ml concentration. After 2-3 weeks in culture, cells were amplified with methotrexate in a stepwise selection. The stable cell lines obtained were designated CD-sPSL-Q (R3.2) and CD-sPSL-Fc (R8.1), respectively. The expression of sPSL-Q and sPSL-Fc protein was confirmed by standard immunoprecipitation method using the anti P-selectin ligand protein polyclonal antibody of Example 7(A).

30

EXAMPLE 4

ASSAYS OF P-SELECTIN-MEDIATED INTERCELLULAR ADHESION

35 A. LEC-v1 Binding Assay

A DNA encoding a chimeric form of P-selectin conjugated to the Fc portion of a human IgGyl (LEC-γ1) was constructed using known methods (Aruffo *et al.*, Cell 67 35-44, 1991), and

5 stably transfected into untr CHO cells (CHO 10. KK) for high level production of the chimera LEC-y1, which was purified for use in the binding assay set forth below.

10 Petri dishes were coated first with a polyclonal anti-human IgGy1 Fc antibody and then with LEC-y1. This method orients the LEC-y1 construct such that the P-selectin portion of the chimera molecule is presented on the surface of the plates. Adhesion of HL60 cells to the oriented LEC-y1 was quantitated in the presence and absence of calcium. HL60 adhesion was shown to be calcium dependent, confirming that the chimera molecule had retained functional binding of P-selectin to its ligand on HL60 cells. The binding of HL60 cells to oriented LEC-y1 was also shown to be blocked by a neutralizing monoclonal antibody to P-selectin, demonstrating the specificity of P-selectin binding.

15

B. Fluorescent CHO-P-selectin Binding Assay

The assay employed a fluorescently labeled CHO P-selectin cell line (Larsen *et al.*, J Biol. Chem. 267, 11104-11110 (1992)) that can bind to and form clusters on the surface of COS cells that are co-transfected with the P-selectin ligand gene and the 3/4 FT gene. The CHO P-selectin cells were suspended at 1.5×10^6 cells/ml in 1% fetal bovine serum in DME medium and labeled by adding 6-carboxyfluorescein diacetate (6-CFD) to a final concentration of 100 μ g/ml. After incubation at 37°C for 15 minutes, the cells were washed in medium and resuspended at 1×10^5 cells/ml. Five ml of the labeled cells were added to each washed COS transfectant-containing plate to be assayed and incubated at room temperature for 10 minutes. Nonadherent cells were removed by four washes with medium. The plates were then scanned by fluorescence microscopy for rosettes of adherent CHO-P-selectin cells.

25

C. Quantitative adhesion assay using radioactively labeled CHO:P-selectin cells

30 COS cells were co-transfected with the pMT21-PL35 plasmid of Example 1 and the pEA.3/4FT plasmid of Example 2 by the same procedure used in the initial stages of screening. As controls, COS cells were transfected with pMT21-PL35 alone, or with pEA.3/4FT alone, or with a similar plasmid containing no insert ("mock"). 24 hours post-transfection, the transfected cells were trypsinized and distributed into Costar 6-well tissue culture plates. CHO P-selectin cells were labeled for 16 hours with 3 H-thymidine using known methods and preincubated at 0.5×10^6 cells/ml for 30 minutes at 4°C in α medium containing 1% BSA (control), α medium containing 1% BSA, 5 mM EDTA and 5 mM EGTA, α medium containing 1% BSA and 10 μ g/ml of a neutralizing anti-P-selectin monoclonal antibody, and α medium containing 1% BSA and a non-neutralizing anti-P-selectin monoclonal antibody. The preincubated cells were then

35

added to the wells containing the transfected COS cells. After 4 h incubation in 10% and cells were removed by 4 changes of medium. The bound CHO-P-selectin cells were released by trypsinization and quantified by scintillation counting.

COS cells co-transfected with P-selectin ligand and the β -FT induced approximately 8-10 fold more binding of CHO-P-selectin cells relative to COS mock cells, assayed in the presence of EGTA and EDTA reduced binding to the level of the mock transfected COS cells. Likewise, incubation with neutralizing anti-P-selectin antibody also eliminated specific binding, whereas non-neutralizing antibody had no effect. In contrast, the binding of CHO-P-selectin to COS cells transfected with P-selectin ligand alone was not statistically different than binding to the mock-transfected COS in both the presence or absence of EDTA and EGTA, or anti-P-selectin antibodies. The binding of CHO-P-selectin cells to COS cells transfected with β -4 FT alone was approximately 2-fold greater than to the mock-transfected COS, but was unaffected by the presence or absence of EDTA and EGTA.

EXAMPLE 5

CONSTRUCTION OF SOLUBLE P-SELECTIN LIGANDS

The EcoRI adaptors used to generate the cDNA library from HL60 cells in Example 1 contain an XbaI restriction site (TCTAGA) just 5' of the beginning of SEQ ID NO 1 as it is located in the pMT21-PL35 plasmid. In order to generate soluble forms of the PSL, the pMT21-PL35 plasmid was restricted with XbaI and with HincII which cleaves after nucleotide 944 of SEQ ID NO 1. The approximately 950 bp fragment thus generated, containing all of the encoded extracellular segment of the ligand up to and including the codon for valine 295, was isolated and used to generate DNAs encoding soluble forms of the P-selectin ligand protein as set forth in sections A through D below.

A. Construction of psPSL.QC

The fragment was purified and ligated into mammalian expression vector pED between the XbaI and EcoRI sites, along with double stranded synthetic oligonucleotide DNA that recreated the codons from Asn 296 to Cys 310 and introduced a novel stop codon immediately following Cys 310. The sequence of the oligos is as follows.

5'-AACTAGCCAGTGGGAGCACCAGACCACATCTCTGTGAAGCACTGCTAG (SEQ ID NO 12)
5'-AATTCTAGCACTGCTTCACAGAGATGTGGTCTGGTGGTCCCACTGGCTAGTT (SEQ ID NO 13)

The resulting plasmid was designated mD-PSL- β QC and the protein expressed from the plasmid was designated sPSL- β QC.

B. Construction of psPSL-Q

The fragment was purified and ligated into the pED plasmid (Kautman et al., 1991) between the XbaI and EcoRI sites, along with the double stranded synthetic oligonucleotide DNA that recreated the codons from Asn 296 to Gln 309 and introduced a novel stop codon immediately following Gln 309. The sequence of the oligos is as follows:

5'-AACTACCCAGTGGGAGCACCAGACCATCTCTCTGTGAAGCACTAG SEQ ID NO 14

5'-AAATTCTACTGCTTCACAGAGATGTGGTGTGGTGCTCCCACTCGGTACTT SEQ ID NO 15

15 The resulting plasmid was designated pED sPSL Q, and the protein expressed from the plasmid was designated sPSL Q.

C. Construction of psPSL.T7

Oligonucleotides encoding 14 amino acids including an epitope derived from the phage T7 major capsid protein were synthesized, creating a C-terminal fusion of the epitope "tag" with an additional 32 amino acids derived from the vector sequence. Two oligonucleotides having the sequences

5'-CTAGACCCCGGGATGGCATCCATGACAGGAGGACAACAAATGGTAGGCCCTAG (SEQ ID NO 16) and

25 5'-AATTGTACGGGCTACCGATTCTTTCTCTCTCTCTCTGATGGATGGCATTCGCGGT-SEQ ID
NO 17)

were duplexed and ligated with the large XbaI-EcoRI fragment of mammalian expression plasmid pED. The resulting plasmid, pED T⁺ was restricted with XbaI and SmaI and ligated to the 950 bp XbaI-HincII fragment described above, resulting in plasmid pED sPSL T⁺.

30 The protein resulting from expression of pED.sPSL.T7 was designated sPSL.T7

D. Construction of Soluble P-selectin Ligand-IgGFc Chimera

The plasmid DNA encoding a soluble, extracellular form of the P-selectin ligand protein fused to the Fc portion of human immunoglobulin (IgG1) was constructed as follows: the mammalian expression vector pED-Fc contains sequences encoding the Fc region of a human IgG1 with a novel linker sequence enabling the fusion of coding sequences amino terminal to the hinge region via a unique XbaI restriction site. A three fragment ligation was performed: pED-Fc was restricted with XbaI and gel purified in linear form. The 950 bp fragment from pMT21-PL85

described above comprised the second treatment. The third treatment consisted of treated synthetic oligonucleotide (NA) having the following sequence:

5'-CTGCGCGCGCGCAAGT- (SEQ ID NO 18)

5'-CTAGACTGCGCGCGCGCAG- (SEQ ID NO 19)

The ligation products were grown as plasmid DNAs and individual clones having the correct configuration were identified by DNA sequencing. The plasmid was designated pED PSL-Fc. The DNA coding region of the resulting soluble P-selectin ligand-Fc fusion protein is shown in SEQ ID NO 6.

EXAMPLE 6

CHARACTERIZATION OF EXPRESSED P-SELECTIN LIGANDS

A. Binding Characterization of Full-Length P-selectin Ligand Protein Expressed on COS Cells

Co-transfection of COS cells with the pEA.3/4FT plasmid of Example 2 and the pMT21.PL85 plasmid of Example 1 yields COS cells which specifically bind to CHO P-selectin cells. This binding is observed only upon co-transfection of pEA.3/4FT and pMT21.PL85; use of either plasmid alone generates COS cells which do not bind to CHO P-selectin cells. No binding is observed between the parental CHO(DUKX) cell line which does not express P-selectin and COS cells co-transfected with pEA.3/4FT and pMT21.PL85. The binding between the co-transfected COS cells and CHO-P-selectin cells is sensitive to chelators of divalent ions such as EDTA and EGTA, consistent with the Ca^{++} dependency of P-selectin mediated cellular adhesion. A neutralizing anti-P-selectin monoclonal antibody blocked the binding between the CHO-P-selectin cells and the COS cells which had been co-transfected with pEA.3/4FT and pMT21.PL85, while a non-neutralizing anti-P-selectin monoclonal antibody had no effect on the binding. The antibody results indicate that the functional domains of P-selectin are required for binding to P-selectin ligand protein expressed on the surface of COS cells.

B. Electrophoretic Characterization of Full-Length P-selectin Ligand Expressed in COS Cells

Detergent extracts of co-transfected COS cells were prepared as follows: 45 hours post co-transfection, approximately 1.5×10^7 cells were suspended in 5 ml of lysis buffer (10 mM Piperazine-N,N'-bis[2-ethanesulfonic acid] (PIPES) pH 7.5, 100 mM KCl, 3 mM $MgCl_2$, 1 mM benzamide, 0.5 μ g/ml leupeptin, 0.75 μ g/ml pepstatin, 1 mM ethylenediamine, and 1 μ g/ml aprotinin) and lysed by sonication. Cellular debris was removed by low speed centrifugation (500 \times g, 10 minutes), and a membrane fraction collected by ultracentrifugation (100,000 \times g, 60 min.

5 The high speed membrane pellet was resuspended in an extraction buffer (10 mM 3-(N-Morpholino)propanesulfonic acid (MOPS), pH 7.5, 0.1 M NaCl, 0.02% Na₂S₂O₄, 1% Thesit[®] (Sigma), 1 mM benzamide, 0.5 µg/ml leupeptin, 0.75 µg/ml pepstatin, 1 mM ethylmaleimide, and 1 µg/ml aprotinin). Samples were then subjected to SDS polyacrylamide gel electrophoresis and transfer to nitrocellulose blots as follows: an aliquot of the detergent extract was suspended
10 in 1% SDS loading buffer and heated for 5 minutes at 100°C before loading onto an 8-16% polyacrylamide gel (reduced) or a 6% gel (non-reduced) and electrophoresed in the Laemmli buffer system. Blots were prepared using Immobilon-P[®] transfer membranes. The blots were immersed in 10 mM MOPS, pH 7.5, 0.1 M NaCl, 0.02% Na₂S₂O₄, 1 mM MgCl₂, 1 mM CaCl₂, and 10% non-fat milk overnight at 4°C. Blots were rinsed once in the above buffer, minus the milk,
15 and incubated in blotting buffer (10 mM MOPS, pH 7.5, 0.1 M NaCl, 1% bovine serum albumin, 0.05% Thesit[®], 1 mM MgCl₂, 1 mM CaCl₂) for 30 minutes at room temperature.

The blots were then probed for the P-selectin ligand as follows: 50 ng of a P-selectin/Fc chimera was pre-incubated with 3 µCi of ¹²⁵I-Protein A in blotting buffer for 30 minutes at room temperature. Additional excipients (e.g., EDTA, EGTA, monoclonal antibodies) could be added
20 to the pre-incubation mixture at this point to evaluate their effects on binding of the chimera to the P-selectin ligand. The pre-incubated mixture was then incubated with the blots (prepared as above) for 60 minutes at room temperature, and the blots were subsequently washed four times with the same blotting buffer (without bovine serum albumin), air dried, and autoradiographed at -70°C.

25 Under non-reducing conditions, two bands were observed with this technique for membrane extracts prepared from co-transfected COS cells. The major band migrated with an estimated molecular weight of approximately 220 kD, whereas the minor band migrated with a molecular weight of approximately 110 kD. Under reducing conditions, only a single band was observed with a molecular weight of approximately 110 kD, indicating that under non-reducing
30 conditions, the P-selectin ligand exists as a homodimer. The approximate molecular weight of the reduced monomer is greater than that predicted from the deduced amino acid sequence of the cDNA clone (45 kD), indicating that the expressed protein undergoes extensive post-translational modification (see Example 6(C)). The specificity of the P-selectin/Fc chimera was confirmed by the observation that a nonspecific IgG₁ probe yielded no bands on the blots. Additionally, the
35 binding of the P-selectin/Fc chimera to the blots was abolished by EDTA, EGTA, and a neutralizing anti-P-selectin monoclonal antibody. Specific bands on the blots were observed only from membrane extracts of COS cells co-transfected with the pEA3/4FT and pMT21/PL85

5 plasmids. Membrane extracts from control transfections (pEA.3/4FT or pMT21-PLS-5) were
run on SDS-PAGE and stained to yield comparable bands on blots.

C. Glycosylation of P-selectin Ligand Protein

10 The presence of covalently attached carbohydrate on recombinant P-selectin ligand and
its role in binding to P-selectin was determined as follows. COS cells were co-transfected with
pED-sPSL T7 of Example 5(C) and the pEA.3/4FT plasmid of Example 2. After 48 hours, the
cells were pulsed with 35 S-methionine. 200 μ l of 35 S-methionine-labeled sPSL T7 conditioned
medium was incubated with 5 μ g LEC- γ 1 in the presence of 2 mM CaCl_2 and 1 mg/ml bovine
serum albumin (BSA). After rotating for 2 hours at 4°C, Protein A-Sepharose beads (Pharmacia)
15 were added for 1 hour at 4°C, pelleted by centrifugation and washed twice in Tns buffered saline
(20 mM Tns-HCl, 150 mM NaCl pH 7.5, hereinafter TBS) containing 2 mM CaCl_2 and 1 mg/ml
BSA. The pellets were then resuspended and treated with neuraminidase (*Streptococcus*
pneumoniae), O-glycanase, and N-glycanase (all from Genzyme) as follows. All glycosidase
digestions were done at 37°C overnight. For neuraminidase digestion, the pellet was resuspended
20 in 50 μ l 2-(N-morpholino)-ethanesulfonic acid (MES) buffer, pH 6.5 (Calbiochem) and 0.1%
SDS, heated at 95°C for 5 minutes, then pelleted. The supernatant was modified to contain 1.4%
n-Octyl B-D-glucopyranoside (OGP), 10mM calcium acetate, 20 mM sodium cacodylate and 2.5
mM PMSE, final pH 7.0. Eight μ l neuraminidase was added for a final concentration of 1 unit/ml.
For neuraminidase/O-glycanase digestion, the sample was prepared as above and along with the
25 neuraminidase, the O-glycanase was also added to a final concentration of 0.1 unit/ml. For N-
glycanase digestion, the pellet was resuspended in 54 μ l MES buffer and 1% SDS, heated at 95°C
for 5 minutes, then pelleted. The supernatant was modified to contain 0.2 M sodium phosphate,
3.5% OGP, and 2.5 mM PMSE, final pH 8.5. N-glycanase was added for a final concentration
of 12 units/ml and incubated as above.

30 The effect of glycosidase treatment on sPSL T7 was assessed in two ways. For this, each
digested protein sample was divided into two equal fractions. One fraction was precipitated with
the P-selectin polyclonal antibody of Example 7(A), to show the effect of digestion on the
electrophoretic mobility. The other fraction was precipitated with the LEC- γ 1 chimera of
Example 4(A), to assess the remaining P-selectin ligand binding activity after digestion. The
35 immunoprecipitated samples were analyzed by SDS-polyacrylamide gel electrophoresis under
reducing conditions and autoradiography.

In the absence of glycosidase treatment, autoradiography revealed comparable bands
with molecular weights of 110 kDa for each precipitation. When the P-selectin ligand protein

5 was treated with neuraminidase, anti-P-selectin ligand polyclonal antibody precipitation revealed
 a slight decrease in mobility, consistent with removal of sialic acid residues. The amount of P-
 selectin ligand protein precipitated by LEC-y1 was significantly reduced after neuraminidase
 treatment, consistent with the role of sialic acid residues in the P-selectin/P-selectin ligand
 interaction. When the P-selectin ligand protein was treated with both neuraminidase and O-
 10 glycanase, a substantial increase in electrophoretic mobility was observed after precipitation with
 the anti-P-selectin ligand polyclonal antibody, indicating that a number of O-linked
 oligosaccharide chains had been removed. However, removal of O-linked oligosaccharides from
 the P-selectin ligand protein may not have been complete, since the electrophoretic mobility did
 not correspond to a protein with a molecular weight of 38 kD, as would be predicted from the
 15 amino acid sequence set forth in SEQ ID NO.1. The neuraminidase/O-glycanase digested P-
 selectin ligand protein bound to LEC-y1 very poorly, further indicating the role of
 oligosaccharides in the P-selectin/P-selectin ligand interaction. Treatment of the purified P-
 selectin ligand with N-glycanase resulted in a slight increase in electrophoretic mobility,
 demonstrating that some of the consensus sites for N-linked glycosylation are occupied. The
 20 amount of P-selectin ligand protein precipitated by LEC-y1 was slightly reduced, indicating that
 N-linked glycosylation also contributes to the P-selectin/P-selectin ligand interaction, though not
 as dramatically as sialylation and O-linked glycosylation.

EXAMPLE 7

25 POLYCLONAL ANTIBODIES SPECIFIC FOR P-SELECTIN LIGANDS

A. Polyclonal Rabbit anti-P-selectin Ligand Protein/Maltose Binding Protein Fusion Protein

30 The anti-P-selectin ligand polyclonal antibody was generated by immunizing rabbits with
 a fusion protein generated in *E. coli*. The fusion protein consisted of the amino terminal one-third
 of the P-selectin ligand (amino acids 1 to 110 of SEQ ID NO.1) fused in frame to the maltose
 binding protein (Maina, C. V. *et al.*, Gene 74, 365-373, 1988), Riggs, P., in Current Protocols in
Molecular Biology, F. M. Ausubel *et al.*, Eds., Greene Associates/Wiley-Interscience (New York,
 1990) chapter 16.6). Under conditions employed herein, the fusion protein antibody recognizes
 35 the P-selectin ligand protein.

B. Polyclonal Rabbit Anti-sPSL.T⁷ Protein

A soluble form of the invention (sPSL.T⁷, see example 5)C1) was purified to apparent
 homogeneity according to the following scheme. COS cells were transfected with three plasmids.

the encoding each of the following: sPSL.T7 (Example 5.C), 3.4FT (Example 2), and a single form of PACE (as set forth in SEQ ID NO.5). After 72 hours, the conditioned medium was collected and recombinant sPSL.T7 was purified as follows:

Conditioned medium was diluted two fold with 50 mM MOPS, 150 mM NaCl, 1.5 mM CaCl₂, and 0.5 mM MnCl₂, pH 7.2, and applied to a column of lentil lectin-Sepharose 4B equilibrated in the same buffer. After loading, the column was washed with the same buffer until the optical absorbance at 280 nm dropped to a stable baseline. The column was then eluted with the same buffer which had been adjusted to 0.5 M α -methyl-mannoside and 0.3 M NaCl. Recombinant sPSL.T7 was collected over 5-15 column volumes of this elution buffer. The lentil lectin eluate was then subjected to a 0-70% ammonium sulfate precipitation by adding 472g of ammonium sulfate per liter of column eluate at 4°C. After stirring for 30 minutes, the precipitate was resuspended in a minimal volume of TBS (20 mM Tris-HCl, 150 mM NaCl, pH 7.5) and applied to a TSK G4000SW_{XL} gel filtration column equilibrated in TBS. The flow rate on the column was 0.5 ml/min and a guard column was employed. In aliquots of < 250 μ l, the resuspended ammonium sulfate pellet was injected on the column and fractions analyzed by SDS-PAGE with Western analysis. Fractions containing sPSL.T7 were pooled and then used for immunizing rabbits.

Antibodies to sPSL.T7 were generated in the standard fashion by antigen priming and subsequent boosting over a 3 month period. Specifically, primary immunization was performed by mixing 50 μ g of sPSL.T7 (denatured by mixing in 0.1% SDS and heating for 10 minutes at 100°C) with complete Freund's adjuvant and injected at five sites subcutaneously. The second (and all subsequent) boosts were performed by mixing 25 μ g of sPSL.T7 (denatured by mixing in 0.1% SDS and heating for 10 minutes at 100°C) (12.5 μ g for the third and subsequent boosts) with incomplete Freund's adjuvant and injecting at two sites subcutaneously (or later, intramuscularly) every two weeks. Test bleeds were performed every two weeks to monitor antibody titer. When the antibody titer reached a suitable level, a larger scale bleed was performed and a total serum fraction prepared. This polyclonal antibody preparation was used to inhibit the specific binding of HL60 cells to CHO P-selectin cells in a manner similar to that described in Example 4.

This assay employed fluorescently-labeled HL60 cells (labelled with BCECFAM, 2',7'-bis-(2-carboxymethyl)-5-(and-6)-carboxyfluorescein, acetoxymethyl ester) binding to CHO cells plated on the bottom of microtiter plates. The labelled HL60 cells were pre-incubated with either sera containing polyclonal antibody or with pre-immune sera for 30 minutes at 4°C. The cells were then washed and incubated with the CHO P-selectin cells for 10 minutes. The plates were

then washed and the fluorescence read with a fluorescence microtiter plate reader. Using this assay, a 1:15 dilution of the anti-PSL T7 polyclonal serum resulted in essentially complete inhibition of HL60 cell binding to CHO P-selectin. Demonstrable inhibition of HL60 binding to CHO P-selectin was still observed at antiserum dilutions of 1:150. Preimmune serum had no effect on HL60 cell binding to CHO P-selectin.

EXAMPLE 8

COTRANSFORMATION WITH CORE2

A. Isolation of the cDNA encoding Core2 GlcNAc Transferase

The cDNA encoding core2 GlcNAc transferase was isolated by standard molecular biology techniques. Two oligos were designed at the 5' and 3' end (including translational initiation and termination codon, respectively) based on the published human core2 sequence (Bierhuizen, M.F.A., Fukuda, M., Proc. Natl. Acad. Sci. 89, 9326-9330 (1992)). The pools of an HL60 cDNA library (Sako, D., Cell 75, 1179-1186 (1993)) were used as template to amplify the core2 coding sequence by a standard PCR protocol. The PCR amplified fragment was purified and subcloned into pED vector. To isolate cDNA, the pools which gave a positive signal in the PCR reaction were transformed into *E. coli* and plated. Transformants were transferred onto nitrocellulose filters and hybridized with a ³²P radiolabelled PCR fragment according to standard protocols. Positive clones were picked and purified by replating. The sequence of the cDNA and PCR clone was confirmed by dideoxy sequencing.

B. Generation of Stable PSGL-1 Chinese Hamster Ovary Cell Lines Expressing Core2 Enzyme

A cell line made in accordance with the methods of Example 3 expressing full-length P-selectin ligand protein and 3/4 fucosyltransferase was co-transfected with core2 cDNA and a neomycin resistance gene (pMT4Neo) by standard calcium phosphate methods. After about two weeks, stable G418-resistant transfectants were picked either as single isolates or in a pool. These transfectants were grown in 1 mg/ml G418 complete DMEM media and analyzed for core2 enzyme activity (Higgins, E.A., *et al.*, J. Biol. Chem. 266, 6280-6290 (1991)). Positive clones or pools found positive for core2 activity were analyzed for P-selectin ligand binding to P-selectin by various methods. In a similar fashion, cell lines expressing either P-selectin ligand protein or soluble P-selectin ligand protein with both the 3/4 fucosyltransferase and PACE enzymes (see Example 3) were used to isolate stable cotransfectants of core2 as described above.

C. Effects of Core2 on P-selectin Binding Activity

The effects of core2 on P-selectin binding activity was evaluated by three different methods:

Binding of PSC1 transcripts to mRNAs and single Polyoma + P
selected IgG columns

48-well plates were coated with 1 μ g/ml anti-human Fc antibody in 50 mM Tris pH 9.5 at 4 $^{\circ}$ C for five to six hours. After washing twice with HBSS buffer, P-selectin-IgG chimera (0.1 μ g/ml conc.) Example 5 was plated in HBSS buffer overnight at 4 $^{\circ}$ C. The plates were blocked with BSA (3 mg/ml) at 4 $^{\circ}$ C for three to four hours. In the case of soluble P-selectin ligand protein, the protein was coated directly onto plates in the same buffer. The 3 H labelled CHO cells were lifted with 2 mM EGTA, washed three times with PBS, and resuspended to a final density of 10^6 cells/ml. A 300 μ l aliquot of this suspension was added to each well (300,000 cells/well). After incubating for 12 minutes at room temperature, wells were washed four times with serum free DMEM to remove unbound cells. Bound cells were lifted with 5 mM EGTA and counted in scintillation counter. U937 cells, used as a positive control for native P-selectin ligand protein binding, were pretreated with gamma globulin (5 mg/ml) to block endogenous Fc receptor before binding to P-selectin IgG chimera. Comparative binding data are shown in Fig. 1.

2. Immunoprecipitation of PSGL-1 with P-selectin/IgG Chimera

20 Recombinant full-length or soluble P-selectin ligand protein prepared from transformants, with and without additional core2, was labelled with 35 S-methionine and subsequently immunoprecipitated with either the anti P-selectin ligand protein polyclonal antibody or P-selectin ligand/IgG chimera as described previously in Examples 7 and 5: Sako, D., Cell 175, 1179-1186 (1993). Data are depicted in Fig. 2.

3 Flow Cytometry

Stable murine P-selectin ligand protein transfectants (with and without core2) were analyzed by standard FACS techniques using either P-selectin:IgG chimera (LebY1 - Example 5) or anti P-selectin ligand protein monoclonal antibody (MAB 275, raised against a peptide having the sequence from amino acid 42 to amino acid 56 of SEQ ID NO 2). Both reagents were pre-conjugated to FITC labelled Protein A. Cells were analyzed by FACS after incubating with this conjugate for 30 minutes at 4°C in the presence of 2 mM CuCl₂. Data are depicted in Fig. 3.

EXAMPLE 9

E-SELECTIN BINDING OF P-SELECTIN BINDING PROTEIN

E-Selectin/IgG chimera was made as described in Example 5 for the P-selectin IgG chimera using an E-selectin encoding DNA including amino acids -21 to 536 of the sequence reported in Bevilacqua *et al.*, Science, 243:1160 (1989).

U937 cells (approximately 6.5×10^6) were recovered from tissue culture plates and divided equally into two 50 mL cultures (final concentration of 1.3×10^6 cells/mL) containing fresh complete RPMI medium and 50 μ Ci/mL of 3 H-glucosamine hydrochloride (labels the protein-linked carbohydrate of glycoproteins [Varki, FASEB 5:226-235 (1991)]). After 48 hours incubation, the cells from both cultures were recovered by centrifugation and washed three times with PBS. The pelleted cells were suspended in 2.5 mL each of a lysis buffer containing 1% Triton X-100 and disrupted by probe sonication for two minutes. The detergent lysates were placed on ice for three hours and then resonicated for an additional two minutes. The lysates were centrifuged at 16,000 rpm for five minutes, the supernatants were recovered and each adjusted to 12 mL with lysis buffer containing no detergent. To one of the two diluted cell lysates was added 100 μ L of protein A sepharose precoupled with P-selectin/IgG chimera (see Example 5) and to the other was added 100 μ L of protein A sepharose precoupled with E-selectin/IgG chimera. Both chimeric proteins were present at a density of approximately 2 mg protein/mL of resin. Binding reactions were allowed to proceed overnight at 4 degrees C with end-over-end mixing. On occasion, purified membranes from U937 cells served as the starting material for the detergent extraction of labeled proteins. In these cases, the detergent extraction and affinity precipitation steps were essentially identical to the above.

Following incubation, the two parallel reaction mixtures were each centrifuged at 2,000 rpm and supernatants were discarded. The resin pellets were washed four times with buffer (10 mM MOPS, 100 mM NaCl, 1 mM CaCl_2 , 1 mM MgCl_2 , 0.02% NaN_3 , pH 7.5 with Triton X-100 [0.25% for the first and second washes, 0.1% for the third wash and 0.01% for the fourth wash]). A final 1 mL pre-elution wash of each resin pellet using buffer containing 0.01% Triton X-100 was conducted and these were retained for quantitation of radioactive counts by liquid scintillation counting (LSC). The resins were then eluted overnight at 4 degrees C with end-over-end mixing in 1 mL each of buffer containing 0.01% Triton X-100 and 10 mM EDTA. The supernatants were recovered by centrifugation and then quantitated by LSC.

Autoradiography of the materials released from the resins by EDTA was performed by electrophoresis of samples (approximately 10,000 cpm samples concentrated by Centricon-10 units where needed) on 10% cross-linked SDS-PAGE gels, subsequent treatment of the gels with ENHANCE (Dupont) as per the manufacturer's instructions followed by drying for two hours on a commercially available gel dryer (Bio-Rad). Exposure of the dried gels to X-ray film was conducted for a minimum of three days at -80 degrees C.

Elution of immobilized E- or P-selectin, previously exposed to detergent extracts of U937 cells and exhaustively washed with EDTA yielded liberated, 3 H-glucosamine labeled

proteins. The amount of radioactivity recovered from the EDTA eluates was at least 10 fold higher than the counts observed in the final, pre-EDTA washes. This observation suggests that both P- and E-selectin chimeras affinity captured ligands from U937 whole cell lysates in an EDTA-dependent manner and that captured ligands were subsequently released upon treatment of the resins with EDTA.

The evaluation of the proteins released by EDTA from the two chimeras was performed by SDS-PAGE and autoradiography under reducing and non-reducing conditions. commercially available ¹²⁵I-labeled molecular weight standards were employed. As shown by the autoradiograph depicted in Fig. 4, the released counts from the whole cell lysates treated with the P-selectin chimera (lanes 2 and 10) and the E-selectin chimera (lanes 4 and 8) correlated to a major species of 200 kD molecular weight, non-reduced (lanes 2 and 4), and 100 kD reduced (lanes 8 and 10). In different experiments depicted in Fig. 4, where purified membrane extracts were used as the starting material in place of whole cells, both the E-selectin chimera (lane 3, non-reduced and lane 9, reduced) and the P-selectin chimera (not shown) gave similar results. Other experiments have demonstrated that the major U937 glycoprotein which binds to P-selectin is immunoreactive with Rb3026, a polyclonal antibody raised against recombinant sPSGL1 77. Therefore, P- and E-selectin specifically recognize a single major glycoprotein species with identical properties in each case.

Example 10

Production and Analysis of Deleted or Altered Forms of Soluble P-Selectin Ligand Protein

A. Generation of DNA Constructs

Truncated forms of the P-selectin ligand protein-IgG chimeras were generated as follows. Plasmid pED.PSL.Fc was restricted with PstI and NotI and the 6kb fragment comprising the Fc portion and vector, pEDFc6kb, was gel purified. Plasmid constructs pED 149 Fc, pED 47 Fc and pED 19 Fc were created by standard PCR technique, using the following pairs of oligonucleotide primers:

"Upstream" primer for all constructs:

5'-CCAGGTCCAACTGCAGGTCCACTCTAGAGGGCACTTCTTCTGGGGCCAGG-3' SEQ ID NO 20.

"Downstream" primer for 148Fc:

5'-TATTATCTGTGCGGGCGCGCTCCAGAACCCATGGCTGCTGTTGCACTGG-3' SEQ ID NO 21.

"Downstream" primer for 47Fc:

5'-TATTATCTGTGCGGGCGCGCAGGCTCCACACTGCTAG-3' SEQ ID NO 22.

"Downstream" primer for 19Fc:

5 5'-TATTATCTGTGCGCGCGCGCGAGGGCTCGCTTCTGCGAG-3' (SEQ ID NO 23)

The template DNA for PCR reaction was pED PSL Fc. The PCR conditions were 94°C, 1 min., 42°C, 1 min., 72°C, 3 min., 25 cycles, using a Perkin-Elmer Thermocycler. After completion of the last cycle, the reaction was treated with Klenow enzyme at 25°C for 30 min., extracted with phenol chloroform, sodium acetate added to 0.3M, and the PCR product DNA was precipitated
10 with 2.5 volumes of ethanol. The DNA pellet was rinsed with 70% ethanol and residual ethanol was evaporated. The resuspended DNA was digested with PstI and NotI, gel purified and ligated with the pEDFc6kb fragment described above. Correct constructs were identified by restriction analysis and confirmed by DNA sequencing.

Plasmid pED ΔY148 Fc, pED H24 Q70.148.Fc were created by site directed mutagenesis
15 (Maniatis et al., 1989, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratories) using pED 148Fc as template and the following mutagenesis oligonucleotides for ΔY148: 5'-CGGAGACAGGCCACCGAATTGCTGCCAGAAACG-3' (SEQ ID NO 24)

for H24: 5'-CCTCCAGAAATGCTGAGGCACAGCACTGACACCACTCCTG-3' (SEQ ID NO 25)

for Q70: 5'-GAGCTGGCCAAACATGGGGCAACTGTCCACGGATTTCAGCAG-3' (SEQ ID NO 26)

20 Positive clones were identified by colony hybridization (Maniatis et al. supra).

pED FFFE.148.Fc was constructed by restricting pED.ΔY148 Fc with EcoRI and ligating the following duplexed oligonucleotides:

5'-AATTCGAGTTCTCTAGATTTTG-3' (SEQ ID NO 27) and

5'-AATTCAAAATCTAGGAACCTG-3' (SEQ ID NO 28).

25 Constructs of the series pED FYYD.19.Fc, pED FFYD.19.Fc and pED FFFD.19.Fc, were made by restricting pED.ΔY148.Fc with EcoRI and NotI and ligating the following duplexed oligonucleotides:

for pED.FYYD.19.Fc:

5'-AATTCGAGTACCTAGATTATGATTTCTCTGCCAGAACTGAGCCTCCGC-3' (SEQ ID NO 29)

30 and

5'-GGCCGCGGAGGCTCAGTTTCTGGCAGGAAATCATAATCTAGGTAACCTG-3' (SEQ ID NO 30).

for pED FFYD.19.Fc,

5'-AATTCGAGTTCTCTAGATTATGATTTCTCTGCCAGAACTGAGCCTCCGC-3' (SEQ ID NO 31)

and

35 5'-GGCCGCGGAGGCTCAGTTTCTGGCAGGAAATCATAATCTAGGAACTG-3' (SEQ ID NO 32).

for pED FFFD.19.Fc:

5'-AATTCGAGTTCTCTAGATTTCGATTTCTCTGCCAGAACTGAGCCTCCGC-3' (SEQ ID NO 33)

and

5 5'-GGGCGCTGGAGGGTCACTTTCTTCCAGGAAATCGAAATCTTCCAACTGCA-3' SEQ ID NO 14

B. Plate Binding Assay for Analysis of Deleted or Altered Forms of Soluble P-Selectin Ligand Protein

The individual plasmid DNAs encoding the various mutated forms of soluble PSGL-1/Fc chimeras were co-transfected with pEA 3/4FT and PACE cDNA in COS cells as described in Example 3(c). 50mls of serum free medium, collected 40-64 hours post transfection from approximately 10^7 COS cells, was purified on a column of 0.25ml of protein A sepharose (Pharmacia) equilibrated with TBS supplemented with 2mM CaCl_2 . After washing with 20mls of TBS/ CaCl_2 , the bound material was eluted with 0.5mls of 0.1M acetic acid, 0.15M NaCl, 2mM CaCl_2 . The eluted material was neutralized with 1/20th volume 3M Tris pH 9.0. The material was quantitated by measuring absorbance at 280nm and by comassie blue staining of PAGE/SDS/Laemmli gels.

In order to produce non-sulfated forms of soluble PSGL-1, COS cell transfections of the relevant Fc chimeras were performed as described above except that following transfection the cells were cultured in the presence of 50mM Chlorate (Sigma).

Quantitative adhesion of CHO-P-selectin, CHO E-selectin and CHO L-selectin expressing cells was performed as described in Example 4(c), with the following modifications: COS cell and antibodies were omitted. Instead, 48-well microtiter plates (Costar) were coated for 16 hours at 4°C with varying quantities of protein A-purified soluble PSGL-1/Fc chimeras. The unbound material was removed and the coated wells were treated with Hanks buffered saline (HBS) with 1mg/ml BSA and 2mM CaCl_2 for 1 hour at 4°C. Tritium labeled CHO selectin expressing cells were added and binding quantitated as described in Example 4(c).

C. Effects of Alteration of N-Linked Glycosylation Sites

Constructs expressing three P-selectin ligand-IgG chimeras were constructed to examine the effects of N-linked glycosylation sites on selectin binding. These constructs had the following characteristics:

- 148.Fc amino acids 42-189 of SEQ ID NO 2
- Q70 148.Fc amino acids 42-189 of SEQ ID NO 2, with the asparagine residue at position 111 of SEQ ID NO 2 replaced with a glutamine residue
- H24 Q70 148.Fc amino acids 42-189 of SEQ ID NO 2, with the asparagine residue at position 65 of SEQ ID NO 2 with a histidine residue and the asparagine residue at position 111 of SEQ ID NO 2 replaced with glutamine residue

These constructs are schematically represented in Fig. 6

The binding of these constructs to protein A and P-selectin-IgG chimera (LEC-γ1) was compared. The results of these experiments are shown in Fig. 7. Comparison of lanes 4, 5 and 6 in the autoradiograph demonstrates that removal of one or both of the first two N-linked glycosylation sites in soluble P-selectin ligand protein does not significantly effect its binding to P-selectin.

10 D. Effects of Tyrosines

Constructs were made to examine the role of tyrosine in P-selectin ligand protein binding to selectins by alteration of the anionic region of the soluble protein. The following constructs were made:

ΔY148.Fc amino acids 42-189 of SEQ ID NO 2, with amino acids 46-52
15 deleted

FFFE148.Fc amino acids 42-189 of SEQ ID NO:2, with the tyrosine residues at
positions 46, 48 and 51 replaced with phenylalanine residues and the
aspartic acid residue at position 52 replaced with a glutamic acid residue

These constructs are schematically represented in Fig. 8.

20 The degree and sites of sulfation of P-selectin ligand protein were examined by expressing relevant constructs in the presence of radioactively labelled sulfate. The degree of sulfation of 148.Fc and ΔY148.Fc were compared to that of a P-selectin-IgG chimera, which was not sulfated. Results are depicted in Fig. 9. These data demonstrate that the majority of sulfate incorporation is into the anionic region of the P-selectin ligand protein.

25 Additional constructs were made to determine whether the sulfation of the anionic region occurred at the tyrosine residues. The following additional constructs were made:

FYYD19.Fc amino acids 42-60 of SEQ ID NO 2, with the tyrosine residue
at position 46 of SEQ ID NO:2 replaced with a phenylalanine
residue

30 FFYD19.Fc amino acids 42-60 of SEQ ID NO 2, with the tyrosine residues
at positions 46 and 48 of SEQ ID NO 2 replaced with a
phenylalanine residues

FFFD19.Fc amino acids 42-60 of SEQ ID NO 2, with the tyrosine residues
at positions 46, 48 and 51 of SEQ ID NO 2 replaced with
35 phenylalanine residues

These constructs are schematically represented in Fig. 9

The degree of sulfation of these constructs was compared to 19.Fc + YYYD19.Fc.
Results are shown in Fig. 10. FYYD19.Fc showed significant sulfation while FFFD19.Fc was

5 substantially less sulfated. Thus, the tyrosine residues in the anionic region of P-selectin ligand proteins are the major site of sulfation.

Removal of sulfate from P-selectin ligand protein substantially reduces its binding to P-selectin. The binding of 148.Fc treated with chlorate to P-selectin was examined. As shown in Fig. 11, inhibition of sulfation by chlorate treatment substantially reduced the amount of P-selectin ligand protein binding to P-selectin.

10 E. Effects of C-terminal Deletions

Several additional C-terminal deleted constructs were made as follows:

254.Fc amino acids 42-295 of SEQ ID NO.2

47.Fc amino acids 42-88 of SEQ ID NO.2

15 19.Fc amino acids 42-60 of SEQ ID NO.2

These constructs are schematically represented in Fig. 12.

The binding of 254.Fc, 148.Fc, 47.Fc and 19.Fc to P-selectin, E-selectin and L-selectin was tested. Figs. 23 and 24 compare the binding of these deletion chimeras to selectins and controls. Results are also summarized in Fig. 12.

20 F. Binding to P-selectin and E-selectin Expressing Cells

Binding of various constructs described above to cells expressing P-selectin and E-selectin was compared using a quantitative plate binding assay of Example 4(c) (which is schematically described in Fig. 13).

Fig. 14 compares the binding of 148.Fc, Δ Y.148.Fc, FFFE.148.Fc and human IgG1 to P-selectin expressing CHO cells. Deletion of all of the tyrosine residues in the anionic region in Δ Y.148.Fc eliminated binding. Changing the tyrosine residues to phenylalanine residues in FFFE.148.Fc substantially reduced binding as compared to 148.Fc. Thus, it was demonstrated that the presence of the full length anionic region is essential to P-selectin binding and that P-selectin binding is enhanced by sulfation in this region. Fig. 14 also reports control experiments demonstrating that 148.Fc, Δ Y.148.Fc and FFFE.148.Fc do not bind to CHO cells which do not express selectin.

Fig. 15 compares the binding of 148.Fc, Δ Y.148.Fc, FFFE.148.Fc and human IgG1 to E-selectin expressing CHO cells. E-selectin binding was unaffected by the deletions or alterations of the native sequence. Thus, it was demonstrated that the anionic region is not required for E-selectin binding.

Fig. 16 summarizes the results of Figs. 14 and 15.

Fig. 17 compares the binding of 47 Fc to P- and E-selectin expressing CHO cells. 47 Fc demonstrated substantial binding to both selectins despite deletion of the N-linked glycosylation sites at positions 111 and 292 of SEQ ID NO:2.

Fig. 19 compares the binding of FYYD.19 Fc, FFFD.19 Fc, H24.Q70.148 Fc, 148 Fc, and human IgG1 to P-selectin expressing CHO cells. Replacement of all of the tyrosine residues in the anionic region in FFFD.19.Fc eliminated binding. Changing the tyrosine residue at position 46 to a phenylalanine residue in FYYD.19.Fc substantially reduced binding as compared to 148 Fc. Alteration of the N-linked glycosylation sites in H24.Q70.148.Fc did not affect binding. Thus, it was demonstrated that P-selectin binding is enhanced by sulfation in the anionic region and that N-linked glycosylation is not required for P-selectin binding. Fig. 19 also reports control experiments demonstrating that FYYD.19.Fc, FFFD.19.Fc, H24.Q70.148.Fc and 148.Fc do not bind to CHO cells which do not express selectin more than human IgG1 alone.

Fig. 20 compares the binding of FYYD.19 Fc, FFFD.19.Fc, H24.Q70.148 Fc, 148 Fc, and human IgG1 to E-selectin expressing CHO cells. Truncation of the ligand protein to the degree of FYYD.19.Fc and FFFD.19.Fc substantially reduced E-selectin binding. Alteration of the N-linked glycosylation sites in H24.Q70.148.Fc did not significantly affect E-selectin binding. Thus, it was demonstrated that P-selectin ligand proteins comprising amino acids 42 to 60 of SEQ ID NO:2 can selectively bind P-selectin, and, to a substantially less, extent E-selectin.

Fig. 21 summarizes the results of Figs. 19 and 20.

G. Conclusions Regarding P- and E-Selectin Binding

Although applicants do not wish to be bound by any theory, these data allow several conclusions regarding the relationship between P-selectin binding and E-selectin binding by P-selectin ligand proteins. N-linked carbohydrates are not required for binding of a P-selectin ligand protein to either P- or E-selectin. P-selectin ligand proteins as small comprising as little as amino acids 42-60 of SEQ ID NO:2 are capable of binding to P-selectin, and, to a substantially less, extent E-selectin.

Fig. 22 depicts a proposed schematic model for binding of P-selectin ligand proteins to P- and E-selectin. O-linked sLe^x carbohydrate has been demonstrated to be required for both P- and E-selectin binding. Data presented herein demonstrate that sulfated tyrosine residues are implicated in P-selectin binding, but not E-selectin binding. Applicants' data also suggests that no N-linked glycosylation binding site is required.

Example 11

Examination of Aggregation Phenomena and Dimer Formation in Forms of PSGL-1

A panel of PSGL-1 mutants were constructed by site-directed mutagenesis and/or PCR amplification with primers that introduced a stopcodon. The template for all mutagenesis
10 experiments was pPL35.R16 (ATCC 75577, deposited by applicants).

The first group of mutants (C310S and C327S) encode full-length PSGL-1 R16 with only one amino acid change compared to wild-type PSGL-1 R16 (Cys to Ser at position 310 or 327, respectively). COS cells, co-transfected with pEA.3/4FT and the mutants C310S or C327S, were labeled with ³⁵S-methionine. Cell lysates were prepared and the mutant proteins were
15 immunoprecipitated with the P-selectin ligand polyclonal antibody of Example 7(A) and analyzed by SDS-polyacrylamide gel electrophoresis under non-reducing and reducing conditions.

The mutant C327S as well as wild-type PSGL-1 R16 migrated as a homodimer under non-reducing conditions and as a monomer under reducing conditions. In contrast, the mutant C310S migrated as a monomer both under non-reducing and reducing conditions, indicating that
20 the cysteine at position 310 is required for dimer formation of PSGL-1.

Both mutants were also analyzed for their ability to bind to P-selectin. Detergent extracts of co-transfected COS cells were precipitated with the LEC-γ1 chimera of Example 4(A). The precipitates were analyzed by SDS-PAGE under non-reducing and reducing conditions and by autoradiography. Both PSGL-1 R16 and C327S were efficiently precipitated by LEC-γ1, whereas
25 C310S binding to LEC-γ1 was greatly reduced, indicating that the dimeric form of PSGL-1 binds P-selectin more tightly than the monomeric form.

The second set of mutants encode soluble forms of PSGL-1 R16 and are listed in Table 1. The mutant ΔTM was generated by site-directed mutagenesis and has a deletion of the transmembrane domain (amino acids 313-333) followed by RLSRKA. The mutants L311, L312,
30 A313, I314, L315, A318 and T322 were generated by site-directed mutagenesis or PCR amplification with PCR primers that introduced a stop codon in the desired position. The name of the mutant refers to the C-terminal amino acid of each truncated soluble form of PSGL-1 R16. The mutants were analyzed according to the following criteria:

1. Expression and secretion from transfected COS cells
- 35 2. Dimer versus monomer formation
3. Lack of aggregate formation
4. P-selectin binding (LEC-γ1 chimera)

5 The mutants Δ TM and I316 fulfilled all four criteria. The shorter soluble forms of PSGL-1, such as sPSL.QC of Example 5(A), L311, L312, A313, I314 and L315 did not form dimers as well and the longer soluble forms of PSGL-1, such as sPSL.T7 of Example 5(C), A318 and T322 formed high molecular weight aggregates which were less desirable.

10 CHO cells, already expressing 3/4 fucosyltransferase and Core2 transferase, were transfected with psPSL.T7, Δ TM, I316 or psPSL.QC and amplified using methotrexate. Stable clones were isolated and labeled with 35 S-methionine. Conditioned media was either analyzed directly or first precipitated with LEC-y1 and then analyzed by SDS-PAGE under non-reducing and reducing conditions (Figure 25). The results indicated that Δ TM and I316 were most efficient in dimer formation and P-selectin binding.

Table I

20	Mutant	Dimer Formation	High MW Aggregates	P-selectin binding
	PSL.QC	+	-	+
	L311	+	-	+
25	L312	-	-	-
	A313	-	-	-
30	I314	-	-	-
	L315	-	-	-
	I316	++	-	++
35	A318	--	-	--
	T322	++	+	--
40	Δ TM	++	-	--

Example 12

Specificity of PSGL-1 Binding to P- and E-Selectins

45 Materials A chimeric protein comprising the extracellular domain of human E-selectin and the Fc portion of human IgG₁ was constructed analogously to the P-selectin chimera, LEC-y1, described earlier. The soluble E-selectin chimera was expressed in baculovirus-infected

- 8 French monoclonal hybridoma cells (Invitrogen) and purified to homogeneity by Protein A Sepharose chromatography. Plasmid vectors pEA3-4FT, pPL35, pFCD43, and pEA3-PACE, for CCS expression of α 1,3-1,4-fucosyltransferase (Fuc-TIII), PSGL-1, CD43 (leukostatin), and soluble paired basic amino acid converting enzyme (PACE), respectively, have been described herein and in the literature (Sako et al. (1993) *Cell* **75**, 1179-1186; Rehemtulla, A. & Kaufman, R. J. (1992) *Curr. Opin. Biotechnol.* **3**, 560-565; Wasley et al. (1993) *J. Biol. Chem.* **268**, 8453-8465).
- 10 Fuc-TVII cDNA (plasmid pMT-FT7) was cloned from an HL60 cDNA expression library using oligonucleotide probes derived from the published sequence (Natsuka et al. (1994) *Journal of Biological Chemistry*, **269**, 16789-16794; Sasaki et al. (1994) *J. Biol. Chem.* **269**, 14730-14737). A polyclonal neutralizing rabbit antibody, Rb3443, was raised against a peptide comprising the first 15 amino acids of the mature (PACE-cleaved) N-terminus of PSGL-1. Monoclonal anti-CD43 antibodies from either Becton Dickinson or Biodesign International and isotype control antibodies were coupled to a solid support consisting of Sepharose TM-4B with a covalently attached goat affinity-purified antibody to mouse IgG (Cappel, Organon Teknika Corporation). Affinity coupling of selectin chimeras and murine antibodies to Protein A Sepharose 4 Fast Flow (Pharmacia) and to the anti-mouse IgG resin, respectively, was carried out at a ratio of 2 mg protein/ml of resin. Antiserum Rb3443 was coupled to Protein A Sepharose at 1 ml/ml resin. Coupling efficiencies, indicated by micro-BCA assay (Pierce) of the post-reacted supernatants, were at least 95%. Aprotinin and pepstatin were from Boehringer Mannheim and benzamidine, leupeptin, and phenylmethylsulfonyl fluoride (PMSF) were from Sigma.
- 25 Labeling and Membrane Extraction of Myeloid Cells. U937 or HL60 cells grown in suspension to a density of $\sim 1.3 \times 10^7$ cells/ml were labeled in 50 ml of RPMI1640 medium supplemented with 10% fetal bovine serum and 2.5 mCi of ^3H -glucosamineHCl (Dupont/NEN) for 48 hr. Activities of greater than 1 cpm/cell were routinely obtained by this technique. The labeled cells were washed with PBS, resuspended in cell lysis buffer (10 mM MOPS, 150 mM NaCl, 4 mM CaCl_2 , and 4 mM MgCl_2 , pH 7.5 containing protease inhibitors 20 mg/ml aprotinin, 10 mM benzamidine, 20 mg/ml leupeptin, 5 mg/ml pepstatin, and 10 mM PMSF) and subjected to several cycles of probe sonication on ice. Nuclei and cell debris were removed by low speed centrifugation and the cell membranes recovered from the supernatant by centrifugation at 100,000g RCF for 1 hr, washed by resuspension and high speed centrifugation in cell lysis buffer containing 1 M NaCl, and finally resuspended in 3 ml membrane solubilization buffer (cell lysis buffer containing 1% Triton X-100). Several cycles of sonication and incubation on ice were employed to solubilize the membrane fraction. Finally, a low speed centrifugation step was employed to remove insoluble membrane residue.
- 35

4 Labeling and Membrane Extraction of Transfected U937 Cells. U937 Myeloid cells were transfected using DEAE dextran and chloroquine (25), employing 3 μ g of plasmids pPL45, pFCD43 and 4 μ g of pEA-SPACE, as well as 4 μ g of pEA-3-4FT or pMT-FT7. After 4-48 hr recovery the transfected cells were starved in serum- and methionine-free DME medium for 30 min and then fed [35 S]-methionine in serum-free DME for 5 hr. The labeled cells were washed, incubated with EGTA to loosen them from the dish surface, scraped from the dish, pelleted, and suspended in cold 10mM PIPES buffer, pH 7.5, containing 100 mM KCl, 3 mM NaCl, 3.5 mM MgCl₂, and protease inhibitors (see above). Membrane extraction then was carried out by sonication, low speed centrifugation, high speed centrifugation, and solubilization in membrane lysis buffer as above, for labeled myeloid cells.

15 Affinity Precipitations. Membrane extracts were diluted 1:4 or 1:5 with cell lysis buffer or with TBSC buffer (20 mM TrisHCl, 150 mM NaCl, 2 mM CaCl₂, pH 7.5) supplemented with 5 mg/ml bovine serum albumin (approximately 99%, Sigma). Extracts thus diluted to 0.2-0.25% Triton X-100 were incubated with human IgG -Protein A Sepharose with end-over-end mixing at 4°C overnight. The precleared supernatants then were reacted for 6-12 hrs at 4°C with Protein A Sepharose precoupled with E- or P-selectin chimeras, control human IgG₁, Rb3443 or with rabbit pre-immune serum or with anti-CD43 antibody or isotype control precoupled to goat anti-mouse IgG Sepharose. The resins were washed 5 or more times in buffer containing 0.1-0.5% Triton X-100 until the radioactivity of the wash supernatants was reduced to background level. Elution of proteins bound specifically to P- or E-selectin resins was accomplished with 10 mM EDTA or 5 mM EDTA/5 mM EGTA at room temperature or by boiling in SDS-PAGE sample buffer (Laemmli, U. K. (1970) *Nature* **227**, 680-685), whereas elution of proteins bound to antibody resins was achieved exclusively by the latter means. For resolution under reducing conditions, dithiothreitol was added to the sample buffer to a final concentration of 100 mM. Samples thus prepared were resolved by SDS-PAGE on 7.5% gels, treated with Enhance (Dupont), dried, and exposed to autoradiography film.

For sequential affinity capture experiments, membrane extracts were precleared, affinity precipitated with P- or E-selectin or human IgG₁, and washed as above. Samples then were eluted twice from the resins with 5 mM EDTA in 10 mM MOPS, 150 mM NaCl, pH 7.5 for 1 hr at 4°C with tumbling. The first and second eluates were combined and then immunoprecipitated with immobilized Rb3443 according to the protocols outlined above.

RESULTS

Soluble E- and P-selectin chimeras were used, in parallel with control human IgG, to probe detergent-solubilized membrane extracts of 3 H-glucosamine-labeled U937 cells as described

under Methods. Examination of eluates from the immobilized selectins by SDS-PAGE autoradiography (Fig. 26) revealed the presence in both P- and E-selectin eluates of a major protein species with identical electrophoretic properties: Mr 200-kDa non-reduced with conversion to a species of Mr 120-kDa following reduction (Fig. 26, lanes 2 and 3, respectively). Occasionally, additional bands were observed in both E- and P-selectin eluates presumably corresponding to this major band and reflecting the presence of naturally reduced material (the 120-kDa species in non-reduced samples) and incomplete reduction (the 200-kDa species in reduced samples). Additionally, a trace band of Mr 150-kDa in the E-selectin eluate which was unaffected by reduction with DTT was occasionally observed. No bands were observed in control experiments using immobilized human IgG₁ (Fig. 26, lane 1) or where elution of selectin resins was performed in the absence of EDTA or SDS (data not shown). Essentially identical results were obtained using HL-60 cells (data not shown). Hence, the nature of these recognition events is interpreted to be specific metal-dependant interactions of these proteins with the respective selectins, presumably via the lectin domains (Lasky, L. A. (1992) *Science* **258**, 964-969; Drickamer, K. (1988) *J. Biol. Chem.* **263**, 9557).

The metal-dependant recognition and electrophoretic behavior of the major band precipitated with E-selectin was consistent with the properties of the previously identified P-selectin counterreceptor, P-selectin glycoprotein ligand or PSGL-1 (Moore et al. (1994) *J. Biol. Chem.* **269**, 23318-23327; Moore et al. (1992) *J. Cell Biol.* **118**, 445-456; Sako et al.).

To assess whether this species was indeed PSGL-1, EDTA eluates of the both E- and P-selectin precipitates were subsequently reacted with the PSGL-1 specific polyclonal antiserum Rb3443. As shown in Fig. 27, the major band isolated by affinity capture with either selectin was immunoprecipitated using this antiserum (lanes 3 and 4, respectively). No species were detected after immunoprecipitation of the control IgG₁ EDTA eluate (Fig. 27, lane 5). Direct immunoprecipitations using fresh ¹²⁵I-labeled U937 membrane extracts confirmed the specificity of Rb3443: precipitation with Rb3443 results in the recovery of a single band with the electrophoretic properties of PSGL-1 (Mr 200-kDa non-reduced, Mr 120-kDa reduced, Fig. 27, lane 2) whereas precipitation with pre-immune antiserum fails to capture any material (Fig. 27, lane 1). These results indicate that the major protein species specifically captured from myeloid cells by both E- and P-selectins is PSGL-1.

To further assess the specificity of E-selectin for PSGL-1, U937 membrane lysates were probed directly for the presence of CD43 (or leukosialin), an abundant cell surface sialoglycoprotein known to bear the major portion of myeloid cell SLe^x residues (Maemura, K. & Fukuda, M. (1992) *J. Biol. Chem.* **267**, 24379-24386). Thus, membrane extracts of

8 H-glycosamine-labeled U937 cells were probed with an anti-CD43 antibody in parallel with the
PSGL-I specific antiserum Rb3443 and control antibodies as described under "Methods". From
identical quantities of membrane lysate, the CD43 antibody precipitated in excess of 30-fold
greater radioactive counts than did the PSGL-I antiserum. Evaluation of the immunoprecipitates
by SDS-PAGE/autoradiography (Fig. 28) revealed a single specific band for each antibody.
10 Rb3443 captured a single species with the electrophoretic characteristics of PSGL-I (Fig. 28,
Lane 2). In contrast, the CD43 antibody precipitated a species with an Mr 120-kDa which was
insensitive to reduction (Fig. 28, Lane 4), consistent with the absence of cysteine in CD43.
Immunoprecipitations with control antibodies (Fig. 28, lanes 1 and 3) proved negative as
expected. There appears to be considerably greater quantities of CD43 than PSGL-I in U937
15 cells, consistent with the quantitation of these proteins in HL-60 cells (Ushiyama et al. (1993) /
Biol. Chem. **268**, 15229-15237). Thus, the inability of E-selectin to precipitate CD43 from
myeloid cells does not appear to be due to its absence in these cell lines. While we cannot exclude
the possibility that E-selectin captures trace quantities of CD43 (i.e., the low-intensity Mr
120-kDa band in Fig. 26, non-reduced lane 3 which is also consistent with monomeric PSGL-I),
20 PSGL-I appears to be the major protein precipitated from myeloid membrane extracts.

Recombinant PSGL-I expressed in COS cells is best achieved with cotransfection of the
PSGL-I cDNA with a cDNA encoding an $\alpha(1,3/1,4)$ fucosyltransferase (Fuc-TIII) for P-selectin
binding (Sako et al.). Interestingly, initial efforts to demonstrate E-selectin recognition of
recombinant PSGL-I failed: E-selectin was unable to capture the counterreceptor from
25 cotransfected COS cell membrane lysates under conditions where P-selectin capture was
successful. One interpretation of this result is that Fuc-TIII was able to modify recombinant
PSGL-I for recognition by P-selectin but was unable to replicate the appropriate modifications
found in myeloid PSGL-I necessary for E-selectin recognition. The recent cloning of a myeloid
fucosyltransferase, Fuc-TVII (Natsuka et al. (1994) *Journal of Biological Chemistry*, **269**, 16789-
30 16794; Sasaki et al. (1994) *J. Biol. Chem.*, **269**, 14730-14737), that is also capable of generating
SLex carbohydrate structures, allowed evaluation of this interpretation.

COS cells were cotransfected with cDNAs encoding either PSGL-I or CD43 and either
Fuc-TIII or Fuc-TVII. Membrane lysates were prepared from the transfected COS cells and these
were precipitated with either immobilized E- or P-selectin chimeras or with antibodies to either
35 PSGL-I or to CD43. The precipitated products were evaluated by SDS-PAGE/autoradiography
following their release by EDTA/EGTA (for selectin mediated binding) or by boiling in SDS (for
immunoprecipitations). The results are shown in Fig. 29.

5 As observed in Fig. 29A, E-selectin capture of COS-expressed PSGL-1 was dependent upon the nature of the fucosyltransferase used in the transfection. In three separate experiments, Fuc-TVII, but not Fuc-TIII, supported PSGL-1 precipitation by E-selectin. The inability of Fuc-TIII to confer E-selectin reactivity to PSGL-1 cannot be attributed to a lack of PSGL-1 expression as the specific antiserum Rb3443 immunoprecipitated significant and comparable
10 quantities of PSGL-1 from both Fuc-TIII and Fuc-TVII transfections (Fig. 29C). Furthermore, P-selectin was capable of precipitating equivalent quantities of PSGL-1 with either fucosyltransferase (Fig. 29B), demonstrating that Fuc-TIII and Fuc-TVII are expressed and active in these cotransfections.

Within the COS recombinant expression system as in myeloid cells, high-affinity
15 E-selectin recognition was also dependent upon the presence of an appropriate polypeptide. Although the polypeptide length, apparent molecular weight, and high frequency and specific types of posttranslational modifications are similar in CD43 and PSGL-1 (Maemura, K. & Fukuda, M. (1992) *J. Biol. Chem.* **267**, 24379-24386), neither fucosyltransferase was able to confer high-affinity E-selectin (or P-selectin) recognition to recombinant leukosialin in
20 cotransfected COS cells (Fig. 29A). Immunoprecipitations with the anti-CD43 antibody indicate that comparable quantities of leukosialin were expressed in both Fuc-TIII and Fuc-TVII cotransfections (Fig. 29C). The failure of E-selectin to capture CD43 was not due to lack of fucosyltransferase activity within the cotransfected COS cells. FACS analysis of COS cells transfected with either PSGL-1 or CD43 and either Fuc-TIII or Fuc-TVII all show high levels of
25 reactivity with the SLeX specific antibody CSLEX-1. Therefore, these results suggest that high-affinity E-selectin recognition requires the presence of a specific polypeptide(s) that is appropriately modified by a specific fucosyltransferase.

Example 13

Inhibition of P-Selectin/PSGL-1 Binding by PSGL-1 Derived Peptides

30 A number of peptides derived from the sequence of PSGL-1 (SEQ ID NO 2) were tested for their ability to inhibit P-selectin/PSGL-1 binding. The tested peptides are listed in Fig. 30.

Inhibition was tested according to the following protocol. The wells of a 96 well plate
35 were coated overnight at 4°C with PSGL-1 in 50 µl of 10 mM MOPS, 150 mM NaCl, 1 mM CaCl₂, 1 mM MgCl₂ at pH 7.5. After removal of the liquid from the wells, 150 µl of 10 mM MOPS, 150 mM NaCl, 1 mM CaCl₂, 1 mM MgCl₂, 0.05% tween-20, 0.05% gelatin at pH 7.5 was added per well to block the unoccupied sites. After 1-2 hr to 2 hr, the block buffer was removed

from the wells and 100 μ l of a complex of 200 μ l P-selectin-human IgG Fc dimer (2 μ g/ml), biotinylated goat anti-human antibody, and streptavidin-conjugated alkaline phosphatase, which had been allowed to tumble at room temperature for 30 minutes to 1 hour plus any potential inhibitors were added per well. Each plate was shaken and tapped to remove the block from plate. The incubation proceeded for 1 hr at room temperature rotating in the dark. The unbound complex was washed off the plate with two 150 μ l portions of 10 mM MOPS, 150 mM NaCl, 1 mM CaCl_2 , 1 mM MgCl_2 , 0.05% tween-20 followed by 150 μ l of 1M diethanolamine, 0.5 mM MgCl_2 . The chromogenic substrate for alkaline phosphatase, PNPP, in 10 mM DEA, 0.5 mM MgCl_2 was added and the plate is then read at 405 nm.

The results of these assays are reported in Fig. 30. The peptides comprising amino acids 48-51 (in which the tyrosine residues have been phosphorylated) and amino acids 42-56 of SEQ ID NO. 2 provided particularly desirable results.

Example 14

Purification of a Soluble Form of PSGL-1

Substantial purification of a soluble form of P-selectin ligand protein has been achieved according to the protocol described below.

A soluble P-selectin ligand protein, I316 (amino acid 42 to amino acid 316 of SEQ ID NO. 2) was expressed in CHO cells as described herein. CHO cell conditioned media was concentrated with a Pellicon ultrafiltration membrane unit (Millipore) with either 10,000 molecular weight cutoff (MWCO) or 30,000 MWCO to about 10 times the original concentration. The buffer was then exchanged into 25 mM Tris, 1mM CaCl_2 , pH 7.4.

The buffer-exchanged concentrate was loaded onto a Toyopearl QAE 550C (TosoHaas) column. Alternatively, the buffer exchange step can be omitted and the concentrate can be diluted one part concentrate to three parts 25 mM Tris, 1mM CaCl_2 , pH 7.4, and then loaded onto the column. The column was washed with 5-10 column volumes (CV) of 25 mM Tris, 1mM CaCl_2 , pH 7.4 at 4°C.

The P-selectin ligand protein eluted from the column with a linear NaCl gradient (0 M NaCl to 1.0 M NaCl) in the 25 mM Tris, 1mM CaCl_2 , pH 7.4 buffer in approximately five column volumes. Two peaks were eluted from the column. The second peak contained the P-selectin ligand protein and was collected in bulk.

The peak from the QAE column was concentrated with a tangential flow ultrafiltration membrane (Millipore) with a 30,000 MWCO and was then buffer exchanged into 25 mM Tris, 150 mM NaCl, 1mM CaCl_2 , pH 7.4 at 4°C.

5 The buffer-exchanged concentrate was loaded onto a Jacalin-Agarose column (10 mg/ml) at 4°C. The column was washed with the dialfiltration buffer and the P-selectin ligand protein was eluted with a gradient of methyl α -D-galactopyranoside (0-100 mM) in 450 mM methyl α -D-galactopyranoside at 20°C. Fractions from the Jacalin column were analyzed by SDS-PAGE and the purest fractions were pooled.

10

Example 15

P-Selectin Ligand Protein Fusions

Four fusions of a P-selectin ligand protein with a different amino acid sequence were constructed: 47 Fc, 47 AGP, 47 BMP and 47 IL11.

15

47 Fc: A cDNA was constructed encoding the signal peptide, PACE cleavage site and first 47 amino acids of the mature P-selectin ligand sequence fused to a mutated Fc region of human IgG1 at His224 of the native Fc sequence. The sequence of the cDNA construct is reported as SEQ ID NO:35. The fusion point is a novel NotI site at nucleotide 261. The amino acid sequence encoded by the cDNA construct is reported as SEQ ID NO:36. The mature amino acid sequence of the encoded fusion protein begins at amino acid 42 of SEQ ID NO:36. The mutations in the Fc portion were a change of Leu 234 and Gly237 of the native Fc sequence to Ala.

20

47 AGP: A cDNA was constructed encoding the signal peptide, PACE cleavage site and first 47 amino acids of the mature P-selectin ligand sequence fused to the first leucine residue of mature human AGP. The sequence of the cDNA construct is reported as SEQ ID NO:37. The fusion point is a novel NotI site at nucleotide 261. The amino acid sequence encoded by the cDNA construct is reported as SEQ ID NO:38. The mature amino acid sequence of the encoded fusion protein begins at amino acid 42 of SEQ ID NO:38.

25

47 BMP: A cDNA was constructed encoding the signal peptide, PACE cleavage site and first 47 amino acids of the mature P-selectin ligand sequence fused to the sequence of mature human BMP-2 (with its first 3 amino acids deleted). The sequence of the cDNA construct is reported as SEQ ID NO:39. The fusion point is a novel NotI site at nucleotide 261. The amino acid sequence encoded by the cDNA construct is reported as SEQ ID NO:40. The mature amino acid sequence of the encoded fusion protein begins at amino acid 42 of SEQ ID NO:40.

30

47 IL11: A cDNA was constructed encoding the signal peptide, PACE cleavage site and first 47 amino acids of the mature P-selectin ligand sequence fused to mature human IL-11. The sequence of the cDNA construct is reported as SEQ ID NO:41. The fusion point is a novel NotI site at nucleotide 261. The amino acid sequence encoded by the cDNA construct is reported as

35

- 5 SEQ ID NO 42 The mature amino acid sequence of the encoded protein begins at amino acid 42 of SEQ ID NO 42

Patent and literature references cited herein are incorporated as if fully set forth

SEQUENCE LISTING

I. GENERAL INFORMATION

1. APPLICANT: Larsen, Glenn
Sako, Blanne
Chang, Xiao Jia
Weidman, Georgette M.
Dumming, Dale
Kumar, Ravindra
Shaw, Gray

2. TITLE OF INVENTION: NOVEL P-SELECTIN LIGAND PROTEIN

3. NUMBER OF SEQUENCES: 40

IV. CORRESPONDENCE ADDRESS:

A. ADDRESSEE: LEGAL AFFAIRS
B. STREET: 87 CAMBRIDGE PARK DRIVE
C. CITY: CAMBRIDGE
D. STATE: MA
E. COUNTRY: USA
F. ZIP: 02140

V. COMPUTER READABLE FORM:

A. MEDIUM TYPE: Floppy disk
B. COMPUTER: IBM PC compatible
C. OPERATING SYSTEM: PC-DOS/MS-DOS
D. SOFTWARE: PatentIn Release #1.0, Version #1.05

VI. CURRENT APPLICATION DATA:

A. APPLICATION NUMBER:
B. FILING DATE:
C. CLASSIFICATION:

VII. PRIOR APPLICATION DATA:

A. APPLICATION NUMBER: US 97 948,660
B. FILING DATE: 28-OCT-1997

VIII. PRIOR APPLICATION DATA:

A. APPLICATION NUMBER: US 98 112,608
B. FILING DATE: 16-AUG-1998

IX. PRIOR APPLICATION DATA:

A. APPLICATION NUMBER: PCT US98 10168
B. FILING DATE: 28-OCT-1998

X. PRIOR APPLICATION DATA:

A. APPLICATION NUMBER: US 98 038,398
B. FILING DATE: 18-APR-1999

XI. PRIOR APPLICATION DATA:

A. APPLICATION NUMBER: US 98 028,305
B. FILING DATE: 10-SEP-1999

VII. PRIOR APPLICATION DATA

A. APPLICATION NUMBER: WO 97/41870
 B. FILING DATE: 18-APR-1998

VIII. ATTORNEY AGENT INFORMATION

A. NAME: BROWN, SCOTT A.
 B. REGISTRATION NUMBER: 30,724
 C. REFERENCE DOCKET NUMBER: GI 5113F

(IX) TELECOMMUNICATION INFORMATION:

(A) TELEPHONE: (617) 498-6004
 B) TELEFAX: (617) 976-6661

I. INFORMATION FOR SEQ ID NO:1:

II. SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1649 base pairs
 B) TYPE: nucleic acid
 C) STRANDEDNESS: single
 D) TOPOLOGY: linear

III. MOLECULE TYPE: cDNA

(VI) ORIGINAL SOURCE:

(A) ORGANISM: Homo sapiens
 (G) CELL TYPE: Promyelocyte
 (H) CELL LINE: HL60

VII. IMMEDIATE SOURCE:

(B) CLONE: PMT21:PL35

(IX) FEATURE:

(A) NAME KEY: 5'UTR
 B) LOCATION: 1..59

IX. FEATURE:

A. NAME KEY: CDS
 B) LOCATION: 61..1648

IX. FEATURE:

(A) NAME KEY: 3'UTR
 (B) LOCATION: 1669..1649

XI. SEQUENCE DESCRIPTION: SEQ ID NO:1

```

GGCACTTCTT CTGGGCCCCAG GAGGCAGCTG TCCCATGCTT TCGTGGCCAG CCGGGTCCC      59
ATG CCG CTG CAA CTC CTC CTG TTG CTC ATC CTA CTC GGC CCG GGC AAC      107
Met Pro Leu Gln Leu Leu Leu Leu Ile Leu Leu Gly Pro Gly Asn
      1           5           10           15
AGC TTG CAG CTC TGG GAG ACC TGG GCA GAT GAA GCG CAG AAA GCG TTG      155
Ser Leu Gln Leu Trp Asp Thr Trp Ala Asp Gln Ala Gln Lys Ala Leu
      20           25           30

```

CTT CTT CTT CTT CTT CTT CAA CTT CAA CAA CTT CTT CAA CTT CAA CAA	1
Leu Leu Leu Leu Ala Asp Arg Arg Arg Gln Ala Thr Gln Tyr Gln Tyr	
15 45 45	
CTA CAT CAT CAT CTT CTT CTT CCA CAA ACC CAG CTT CCA CAA ATG CTT ACC	151
Leu Asp Thr Asp Phe Leu Pro Gln Thr Gln Pro Pro Gln Met Leu Arg	
51 55 60	
AAC ACC ACT GAC ACC ACT CTT CTT ACT GGG CTT CCA ACC CTT GAG TCT	199
Asn Ser Thr Asp Thr Thr Pro Leu Thr Gly Pro Gly Thr Pro Gln Ser	
65 70 75 80	
ACC ACT CTT CAG CTT CTT CCA ACC CTT TCT ACT GGG CTT GAT CCA CCA	147
Thr Thr Val Gln Pro Ala Ala Arg Arg Ser Thr Gly Leu Asp Ala Gly	
85 90 95	
GGG CCA CTC ACA GAG CTC ACC ACC GAG CTC GCG AAC ATG GGG AAC CTC	195
Gly Ala Val Thr Gln Leu Thr Thr Gln Leu Ala Asn Met Gly Asn Leu	
100 105 110	
TCC ACC GAT TCA CCA CTT ATG GAG ATA CAG ACC ACT CAA CCA CCA GCG	443
Ser Thr Asp Ser Ala Ala Met Gln Ile Gln Thr Thr Gln Pro Ala Ala	
115 120 125	
ACC GAG CCA CAG ACC ACT CCA CTC CCA GCC ACA GAG CCA CAG ACA ACT	431
Thr Gln Ala Gln Thr Thr Pro Leu Ala Ala Thr Gln Ala Gln Thr Thr	
130 135 140	
CCA CTC ACC GCG ACC GAG CCA CAG ACC ACT CCA CTC CCA GCC ACA GAG	539
Arg Leu Thr Ala Thr Gln Ala Gln Thr Thr Pro Leu Ala Ala Thr Gln	
145 150 155 160	
CCA CAG ACC ACT CCA CCA CCA GCG ACC GAA CCA CAG ACC ACT CAA CCG	587
Ala Gln Thr Thr Pro Pro Ala Ala Thr Gln Ala Gln Thr Thr Gln Pro	
165 170 175	
ACA CCG CTC GAG CCA CAG ACC ACT CCA CCA CCA GCG ATG GAG CCA CAG	615
Thr Gln Leu Gln Ala Gln Thr Thr Ala Pro Ala Ala Met Gln Ala Gln	
180 185 190	
ACC ACT CCA CCA CCA GCG ATG GAA CCA CAG ACC ACT CCA CCA CCA GCG	693
Thr Thr Ala Pro Ala Ala Met Gln Ala Gln Thr Thr Pro Pro Ala Ala	
195 200 205	
ATG CAG CCA CAG ACC ACT CAA ACC ACA GCG ATG CAG CCA CAG ACT ACT	701
Met Gln Ala Gln Thr Thr Gln Thr Thr Ala Met Gln Ala Gln Thr Thr	
210 215 220	
CCA CCA GAA GCG ACC GAG CCA CAG ACC ACT CAA CCG ACA GCG ACC GAG	779
Ala Pro Gln Ala Thr Gln Ala Gln Thr Thr Gln Pro Thr Ala Thr Gln	
225 230 235 240	
CCA CAG ACC ACT CCA CTC CCA GCG ATG GAG CCG CTC TTC ACA GAA CCG	807
Ala Gln Thr Thr Pro Leu Ala Ala Met Gln Ala Leu Ser Thr Gln Pro	
245 250 255	

AGT TTT ATA GAG GGT GTG TTT ATG GAA GTT ATT AAT GAA AAT GT GTT Arg Ala Thr Gly Ala Leu Ser Met Glu Asn Thr Thr Leu His Gly Leu 361 366 371	1010
TTT ATA GGT TTT TTT GTG TTT TTT GTT ACT GAG AAG GGC ATT GTG ATG Phe Ile Pro Phe Ser Val Ser Ser Val Thr His Lys Gly Ile Pro Met 375 380 385	1011
GCA GGC AGC AAT TTT TCC GTG AAC TAC CCA GTG GGC GGC CCA GAC GAC Ala Ala Ser Asn Leu Ser Val Asn Tyr Pro Val Gly Ala Pro Asp His 390 395 400	1012
ATC TTT GTG AAG GAG TCC GTG GTG GGC ATC GTA ATT TTT GGC GTG GTG Ile Ser Val Lys Glu Cys Leu Leu Ala Ile Leu Ile Leu Ala Leu Val 405 410 415 420	1013
GCC ACT ATC TTT GTG TTT GTG TTT ACT GTG GTG GTG GGC GTG GGC GTG TCC Ala Thr Ile Phe Phe Val Cys Thr Val Val Leu Ala Val Arg Leu Ser 425 430 435	1014
GCC AAG GGC GAC ATG TAC GGC GTG GGT AAT TAC TCC GGC ACC GAG ATG Arg Lys Gly His Met Tyr Pro Val Arg Asn Tyr Ser Pro Thr Glu Met 440 445 450	1015
GTG TCC ATC TCA TCC GTG TTT GGT GAT GGC GGT GAG GGC GGC TCT GGC Val Cys Ile Ser Ser Leu Leu Pro Asp Gly Gly Glu Gly Pro Ser Ala 455 460 465	1016
ACA GGC AAT GGC GGC GTG TCC AAG GGC AAG AGC GGC GGC GTG ACC CCA Thr Ala Asn Gly Gly Leu Ser Lys Ala Lys Ser Pro Gly Leu Thr Pro 470 475 480	1017
GAG GGC AGG GAG GAC GGT GAG GGC GAT GAC GTG ACC GTG GAG AGT TTT Glu Pro Arg Glu Asp Arg Glu Gly Asp Asp Leu Thr Leu His Ser Phe 485 490 495 500	1018
TTG GGT TAGGTGACTG TGGCATCTGT TTTGGCAAGA TGGGACCTGG AGGGGCTGTG Leu Phe	1019
GTGGGGCACC GTTGAGTGCC CAGACCCCAA TGGACAGCTG TGGGTTTCTT GGGAGAGGCT	1020
TGGGCATGGG CATCTTCAGG GAAGGAAGCTG TGGCAGGCA AACAGGACAA GAGGAGCTG	1021
GGGTCAGCA GAGGGCAAG TGGAGGACAG TTTTCTCTCT TTTTCTCTCT GAAGGGCAGG	1022
CACATTTTCA GGGAGGTCCA AGGAGGAGG CCATTTACTT GAGACAGATT GTCTCTCTCT	1023
TCTCTCTCT CATCTCTCTT GGGTCTCTCT AACATCTCTT ATGGCTCTCT CCGCTTCTCT	1024
TGGTCACTGG AGTCTCTCTT CCATCTACCT AAGG	1025

2 INFORMATION FOR SEQ ID NO.1

1. SEQUENCE CHARACTERISTICS

- A LENGTH: 413 amino acids
- B TYPE: coding area
- C TOPOLGY: linear

11. MOLECULE TYPE: protein

X1. SEQUENCE DESCRIPTION: SEQ ID NO:1

```

Met Pro Leu Gln Leu Leu Leu Leu Ile Leu Leu Gly Pro Glu Asn
 1           5           10           15
Ser Leu Gln Leu Trp Asp Thr Trp Ala Asp Glu Ala Glu Lys Ala Leu
 20           25           30
Gly Pro Leu Leu Ala Arg Asp Arg Arg Gln Ala Thr Glu Tyr Glu Tyr
 35           40           45
Leu Asp Tyr Asp Phe Leu Pro Glu Thr Glu Pro Pro Glu Met Leu Arg
 50           55           60
Asn Ser Thr Asp Thr Thr Pro Leu Thr Gly Pro Gly Thr Pro Glu Ser
 65           70           75           80
Thr Thr Val Glu Pro Ala Ala Arg Arg Ser Thr Gly Leu Asp Ala Gly
 85           90           95
Gly Ala Val Thr Glu Leu Thr Thr Glu Leu Ala Asn Met Gly Asn Leu
100          105          110
Ser Thr Asp Ser Ala Ala Met Glu Ile Gln Thr Thr Glu Pro Ala Ala
115          120          125
Thr Glu Ala Gln Thr Thr Pro Leu Ala Ala Thr Glu Ala Gln Thr Thr
130          135          140
Arg Leu Thr Ala Thr Glu Ala Gln Thr Thr Pro Leu Ala Ala Thr Glu
145          150          155          160
Ala Gln Thr Thr Pro Pro Ala Ala Thr Glu Ala Gln Thr Thr Glu Pro
165          170          175
Thr Gly Leu Glu Ala Gln Thr Thr Ala Pro Ala Ala Met Glu Ala Gln
180          185          190
Thr Thr Ala Pro Ala Ala Met Glu Ala Gln Thr Thr Pro Pro Ala Ala
195          200          205
Met Glu Ala Gln Thr Thr Gln Thr Thr Ala Met Glu Ala Gln Thr Thr
210          215          220
Ala Pro Glu Ala Thr Glu Ala Gln Thr Thr Glu Pro Thr Ala Thr Glu
225          230          235          240
Ala Gln Thr Thr Pro Leu Ala Ala Met Glu Ala Leu Ser Thr Glu Pro

```

```

145      15      165
Ser Ala Thr Glu Ala Leu Ser Met Glu Asp Thr Thr Lys Arg Gly Leu
160      165      170
Phe Ile Pro Phe Ser Val Ser Ser Val Thr His Lys Glu Ile Pro Met
175      180      185
Ala Ala Ser Asn Leu Ser Val Asn Tyr Pro Val Gly Ala Pro Asp His
190      195      200
Ile Ser Val Lys Glu Cys Leu Leu Ala Ile Leu Ile Leu Ala Leu Val
205      210      215      220
Ala Thr Ile Phe Phe Val Cys Thr Val Val Leu Ala Val Arg Leu Ser
225      230      235
Arg Lys Gly His Met Tyr Pro Val Arg Asn Tyr Ser Pro Thr Glu Met
240      245      250
Val Cys Ile Ser Ser Leu Leu Pro Asp Gly Gly Glu Gly Pro Ser Ala
255      260      265
Thr Ala Asn Gly Gly Leu Ser Lys Ala Lys Ser Pro Gly Leu Thr Pro
270      275      280
Glu Pro Arg Glu Asp Arg Glu Gly Asp Asp Leu Thr Leu His Ser Phe
285      290      295      300
Leu Pro

```

2. INFORMATION FOR SEQ ID NO:3:

1. SEQUENCE CHARACTERISTICS:

- A) LENGTH: 1039 base pairs
- B) TYPE: nucleic acid
- C) STRANDEDNESS: single
- D) TOPOLOGY: linear

11. MOLECULE TYPE: cDNA, synthetic

12. ORIGINAL SOURCE:

- A) ORGANISM: Homo sapiens
- B) CELL TYPE: placenta

13. FEATURE:

- A) NAME KEY: CDS
- B) LOCATION: 1..1039

14. SEQUENCE DESCRIPTION: SEQ ID NO:3

ATG GGT GTG GAA GTG GTG GTG TTG GTG ATG GGA GTG GGG GTG GTG AAC

GAA AAT ACA GAT ATG GAG GCA GAG AGG AAT GCA GCA AAA GTT AGG GAG
 Ser Thr Thr Ala Met Ile Ala Glu Thr Thr Ala Ser Ile Ala Thr Glu
 233 237 241

GCA GAG AGG AGT CAA GCG ACA GCG AGG GAG GCA GAG AGG AGT GCA GTG
 Ala Ser Thr Thr Gln Pro Thr Ala Thr Glu Ala Glu Thr Thr Pro Leu
 245 251 255

GCA GCG ATG GAG GCG CTG TCC ACA GAA GCG AGT GCG ACA GAG GCG CTG
 Ala Ala Met Glu Ala Leu Ser Thr Glu Pro Ser Ala Thr Glu Ala Leu
 260 265 270

TCC ATG GAA GGT ACT AGC AAA AGA GGT CTG TTC ATA GCG TTT TCT CTG
 Ser Met Glu Pro Thr Thr Lys Arg Gly Leu Phe Ile Pro Phe Ser Val
 275 280 285

TCC TCT GTT ACT GAC AAG GCG ATT GCG ATG GCA GCG AGC AAT TTG TCC
 Ser Ser Val Thr His Lys Gly Ile Pro Met Ala Ala Ser Asn Leu Ser
 290 295 300

GTG AAG TAC GCA GTG GCG GCG GCA GAG GAG ATG TCT GTG AAG CAG TCG
 Val Asn Tyr Pro Val Gly Ala Pro Asp His Ile Ser Val Lys Gln Cys
 305 310 315 320

CTG CTG GCG ATC GTA ATC TTG GCG CTG GTG GCG ACT ATC TTC TTC GTG
 Leu Leu Ala Ile Leu Ile Leu Ala Leu Val Ala Thr Ile Phe Phe Val
 325 330 335

TGC ACT GTG GTG CTG GCG GTC GCG CTG TCC GCG AAG GCG GAC ATG TAC
 Cys Thr Val Val Leu Ala Val Arg Leu Ser Arg Lys Gly His Met Tyr
 340 345 350

GCG GTG GGT AAT TAC TCC GCG AGC GAG ATG GTG TCC AGC TCA TTC CTG
 Pro Val Arg Asn Tyr Ser Pro Thr Glu Met Val Cys Ile Ser Ser Leu
 355 360 365

TTC GCG GAT GCG GGT GAG GCG GCG TCT GCG ACA GCG AAT GCG GCG CTG
 Leu Pro Asp Gly Gly Glu Gly Pro Ser Ala Thr Ala Asn Gly Gly Leu
 370 375 380

TCC AAG GCG AAG AGC GCG GCG CTG AGC GCA GAG GCG AGG GAG GAG GGT
 Ser Lys Ala Lys Ser Pro Gly Leu Thr Pro Glu Pro Arg Glu Asp Arg
 385 390 395 400

GAG GTG GAT GAG GTG AGC GTG GAG AGC TTC GTG TCT TAG
 Glu Gly Asp Asp Leu Thr Leu His Ser Phe Leu Pro
 405 410

(2) INFORMATION FOR SEQ ID NO:4:

A. SEQUENCE CHARACTERISTICS:

- A. LENGTH: 410 amino acids
- B. TYPE: amino acid
- C. TOPOLOGY: linear

4. NUCLEOTIDE SEQUENCE (nt)

4.1. SEQUENCE DESCRIPTION: SEQ ID NO: 4

Met Pro Leu Gln Leu Leu Leu Leu Ile Leu Leu Gly Pro Gly Asn
1 5 10 15

Ser Leu Gln Leu Trp Asp Thr Trp Ala Asp Gln Ala Gln Lys Ala Leu
20 25 30

Gly Pro Leu Leu Ala Arg Asp Arg Arg Gln Ala Thr Gln Tyr Gln Tyr
35 40 45

Leu Asp Tyr Asp Phe Leu Pro Gln Thr Gln Pro Pro Gln Met Leu Arg
50 55 60

Asn Ser Thr Asp Thr Thr Pro Leu Thr Gly Pro Gly Thr Pro Gln Ser
65 70 75 80

Thr Thr Val Gln Pro Ala Ala Arg Arg Ser Thr Gly Leu Asp Ala Gly
85 90 95

Gly Ala Val Thr Gln Leu Thr Thr Gln Leu Ala Asn Met Gly Asn Leu
100 105 110

Ser Thr Asp Ser Ala Ala Met Gln Ile Gln Thr Thr Gln Pro Ala Ala
115 120 125

Thr Gln Ala Gln Thr Thr Gln Pro Val Pro Thr Gln Ala Gln Thr Thr
130 135 140

Pro Leu Ala Ala Thr Gln Ala Gln Thr Thr Arg Leu Thr Ala Thr Gln
145 150 155 160

Ala Gln Thr Thr Pro Leu Ala Ala Thr Gln Ala Gln Thr Thr Pro Pro
165 170 175

Ala Ala Thr Gln Ala Gln Thr Thr Gln Pro Thr Gly Leu Gln Ala Gln
180 185 190

Thr Thr Ala Pro Ala Ala Met Gln Ala Gln Thr Thr Ala Pro Ala Ala
195 200 205

Met Gln Ala Gln Thr Thr Pro Pro Ala Ala Met Gln Ala Gln Thr Thr
210 215 220

Gln Thr Thr Ala Met Gln Ala Gln Thr Thr Ala Pro Gln Ala Thr Gln
225 230 235 240

Ala Gln Thr Thr Gln Pro Thr Ala Thr Gln Ala Gln Thr Thr Pro Leu
245 250 255

Ala Ala Met Gln Ala Leu Ser Thr Gln Pro Ser Ala Thr Gln Ala Leu
260 265 270

Ser Met Glu Ala Thr Thr Lys Arg Gly Leu Phe Ile Val Thr Val
 275 28 285
 Ser Ser Val Thr His Lys Gly Ile Pro Met Ala Ala Ser Val Leu Ser
 290 295 300
 Val Asn Tyr Pro Val Gly Ala Pro Asp His Ile Ser Val Lys Glu Lys
 305 310 315 320
 Leu Leu Ala Ile Leu Ile Leu Ala Leu Val Ala Thr Ile Phe Phe Val
 325 330 335
 Lys Thr Val Val Leu Ala Val Arg Leu Ser Arg Lys Gly His Met Tyr
 340 345 350
 Pro Val Arg Asn Tyr Ser Pro Thr Glu Met Val Lys Ile Ser Ser Leu
 355 360 365
 Leu Pro Asp Gly Gly Glu Gly Pro Ser Ala Thr Ala Asn Gly Gly Leu
 370 375 380
 Ser Lys Ala Lys Ser Pro Gly Leu Thr Pro Glu Pro Arg Glu Asp Arg
 385 390 395 400
 Glu Gly Asp Asp Leu Thr Leu His Ser Phe Leu Pro
 405 410

(C) INFORMATION FOR SEQ ID NO:5:

1. SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1151 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

11. MOLECULE TYPE: cDNA

11. IMMEDIATE SOURCE:
 (B) CLONE: padescl

11. SEQUENCE DESCRIPTION: SEQ ID NO:5

ATGGAGCTGA GGGGCTGCTT GGTATGGGTC GTAGGAGCAA GAGGAAGCTT TGTCTCTGTA 41
 GCACTGTATG CTCAGGGCCA GAAGGTCTTC ACCAAGACCT GGGGTGTGCG CATCTCTGGA 101
 GGGGAGCGG TGGCAACAG TGTGGCAGCG AAGCATGGGT TCTCAACCT GGGCCAGATC 161
 TTGGGGGACT ATTACCACTT CTGGCATGCA GGAAGTACCA AGGGTCTGCT CTGGCTTCAG 221
 GGGGGGGGGG AGAGGGGGCT GAGAGGGGAG GTCAAGTAC AGTGGCTGCA AGAGGAGGTC 281

—

AATTGCTGTA AATGCTGCTGTT GTTGGCTGATC GTACTGGGCTT GTGGCAACAG GTTGCAGCTG 4
 AGGACTGAG AGTGGCTGCTT AGGACAGGCTT GCACTGCTT GTTGGCTGTA GTTGCAGCTG 11
 GGGCAAGCTT TGGGGGAGG GTTGGCTGCTT TCACTGCTT GTTGCAGCTG A 111

2. INFORMATION FOR SEQ ID NO:6.

- A. SEQUENCE CHARACTERISTICS:
 A. LENGTH: 1887 base pairs
 B. TYPE: nucleic acid
 C. STRANDEDNESS: single
 D. TOPOLOGY: linear

II. MOLECULE TYPE: cDNA

VI. ORIGINAL SOURCE:
 A. ORGANISM: Homo sapiens

VII. IMMEDIATE SOURCE:
 B. CLONE: pPSL.F2

XI. SEQUENCE DESCRIPTION: SEQ ID NO:6.

ATGGCTGTTC AACTGCTGCT GTTGGCTGATC GTACTGGGCTT GTGGCAACAG GTTGCAGCTG 40
 TGGGACAGCTT GGGCAGATGA AGCCGAGAAA GCGTTGGGTC GCGTGGCTTC GCGGGACCGG 100
 AGACAGGCTCA CGGAATATGA GTACCTAGAT TATGATTTTC TCCGAGAAAC GGAGCGCTCA 160
 GAAATGCTCA GGAACAGCAC TGACACCACT GGTGTGACTG GCGGTGGAAC GCGTGAATCT 220
 ACGCTGTTCG AGCGTGGCTG AAGGCGTTCT ACTGGGCTTC ATGAGGAGG GCGACTGACA 280
 GAGCTGAGCA GCGAGCTGGC CAACATGGGG AAGCTGTGCA GCGATTGAG ACGTATGAG 340
 ATACAGAGCA CTCACCAAGC AGCCACGGAG GCACAGAGCA CTCGACTGCT AGCCACAGAG 400
 GCACAGAGAA CTCGACTGAC GCGCAGGGAG GCACAGAGCA CTCGACTGCT AGCCACAGAG 460
 GCACAGAGCA CTCGACAGC AGCCACGGAA GCACAGAGCA CTCGACTGCT AGCCACAGAG 520
 GCACAGAGCA CTCGACAGC AGCCATGGAG GCACAGAGCA CTCGACAGC AGCCATGGAG 580
 GCACAGAGCA CTCGACAGC AGCCATGGAG GCACAGAGCA CTCGACAGC AGCCATGGAG 640
 GCACAGAGCA CTCGACAGC AGCCACGGAG GCACAGAGCA CTCGACAGC AGCCACGGAG 700
 GCACAGAGCA CTCGACTGGC AGCCATGGAG GCGCTGTGCA GAGAAAGGAG TGGCAGAGAG 760
 GCGCTGTGCA TGGAAAGCTAC TACCAAAAGA GTGTGTTTCA TACGCTTTTC TGTGTCTCTT 820
 GTTACTGACA AGGCGATTTC CATGGCAGCC AGCAATTTCT GCGTGGCTTC GCGGCACTCT 880

AGAGAGAGAA GTTACACAGT TTAAGTCTCT TTAAGACCTG AATCTCTCTT TTAAGTCTCT
 CTCTCTCTCT TCTCCGAGAA ATCCAGAGAG AGGCTCATGA TCTCCGAGAG CTCTACCTT
 AGATCTCTTG TCTCGACCT GAGCCAGCAA GAGCTCGAGG TTAAGTCTAA CTCTACCTG
 GAGCTCTCTG AGCTGCATAA TCCCAAGACA AAGCCGCGCG AGGAGCAGTA CAACAGCAGG
 TACCTCTCTG TCAGCTCTCT GAGCTCTCTG CAGCAGGACT GGCTGAATGG CAAGCAGTAC
 AACTGCAAGG TCTCAACAA AGGCTCTCTG CTCCGATCG AGAAAACCAT CTCCAAAGCG
 AAAGCGCAGG CCGGAGAACT ACAGCTCTAC AGCTCTCTCT CATCCCGGGA GGAGATGAGT
 AAGAGCAGG TCAGCTCTG CTCTCTCTCT AAAGCTCTCT ATCCAGCGGA CATCTCTCTG
 GAGTGGGAGA GCAATGGGA CCGGAGAGAG AACTACAAGA CCAGCTCTCT CTCTCTCTG
 TCGAGCTCT CTCTCTCTCT CTATAGCAAG CTCTCTCTG ACAAGAGCAG CTCTCTCTG
 CGGAGCTCT TCTCTCTCT CTCTCTCTCT GAGCTCTCT ACAAGCAGTA CAGCCAGAGG
 AGCTCTCTCT TCTCTCTCT TAAATAG

1. INFORMATION FOR SEQ ID NO:7:

- (A) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 19 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(X1) SEQUENCE DESCRIPTION: SEQ ID NO:7

AATCTCTCTG ACTCTAGAG

2. INFORMATION FOR SEQ ID NO:8:

- (A) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 15 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(X1) SEQUENCE DESCRIPTION: SEQ ID NO:8:

TTCTAGAGTC GACCG

1. INFORMATION FOR SEQ ID NO:9:

1. SEQUENCE CHARACTERISTICS:
A. LENGTH: 40 base pairs
B. TYPE: nucleic acid
C. STRANDEDNESS: single
D. TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

*1. SEQUENCE DESCRIPTION: SEQ ID NO:9:

TAGCATACGC TCTAGAGCAT GATCCCCCTG GGTCCAGCCA AGC

40

2. INFORMATION FOR SEQ ID NO:10:

1. SEQUENCE CHARACTERISTICS:
A. LENGTH: 37 base pairs
B. TYPE: nucleic acid
C. STRANDEDNESS: single
D. TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

*1. SEQUENCE DESCRIPTION: SEQ ID NO:10:

CCGGAATTCT CAGGTGAACC AAGCCGC

37

3. INFORMATION FOR SEQ ID NO:11:

1. SEQUENCE CHARACTERISTICS:
A. LENGTH: 35 base pairs
B. TYPE: nucleic acid
C. STRANDEDNESS: single
D. TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

*1. SEQUENCE DESCRIPTION: SEQ ID NO:11:

AAGTATCTGT CAGGGGCTTC CAGGT

35

4. INFORMATION FOR SEQ ID NO:12:

1. SEQUENCE CHARACTERISTICS:
A. LENGTH: 48 base pairs
B. TYPE: nucleic acid
C. STRANDEDNESS: single
D. TOPOLOGY: linear

11. MOLECULE TYPE: cDNA

11.1 SEQUENCE DESCRIPTION: SEQ ID NO:12:

AAGTACCCAG TGGGAGCACC AGACCACATC TCTGTGAAGC ACTGCTAG
48

12. INFORMATION FOR SEQ ID NO:12:

1. SEQUENCE CHARACTERISTICS:
A. LENGTH: 50 base pairs
B. TYPE: nucleic acid
C. STRANDEDNESS: single
D. TOPOLOGY: linear

11.2 MOLECULE TYPE: cDNA

11.1 SEQUENCE DESCRIPTION: SEQ ID NO:13:

AATTGTACCA CTGCTTCACA GAGATGTGCT CTGGTCTCTC CACTGGGTAG TT
80

13. INFORMATION FOR SEQ ID NO:13:

1. SEQUENCE CHARACTERISTICS:
A. LENGTH: 45 base pairs
B. TYPE: nucleic acid
C. STRANDEDNESS: single
D. TOPOLOGY: linear

11.2 MOLECULE TYPE: cDNA

11.1 SEQUENCE DESCRIPTION: SEQ ID NO:14:

AAGTACCCAG TGGGAGCACC AGACCACATC TCTGTGAAGC ACTAG
40

14. INFORMATION FOR SEQ ID NO:14:

1. SEQUENCE CHARACTERISTICS:
A. LENGTH: 45 base pairs
B. TYPE: nucleic acid
C. STRANDEDNESS: single
D. TOPOLOGY: linear

11.2 MOLECULE TYPE: cDNA

11.1 SEQUENCE DESCRIPTION: SEQ ID NO:15:

SEQUENCE CHARACTERISTICS: SEQUENCE ID NO. 16

1. INFORMATION FOR SEQ ID NO. 16:

A. SEQUENCE CHARACTERISTICS:

- A. LENGTH: 50 base pairs
- B. TYPE: nucleic acid
- C. STRANDEDNESS: single
- D. TOPOLOGY: linear

II. MOLECULE TYPE: cDNA

III. SEQUENCE DESCRIPTION: SEQ ID NO. 16:

GTAGACCCCGG CATGGCATCC ATGACAGGAG GACACAAAT CTTAGGCTCT AG

1. INFORMATION FOR SEQ ID NO. 17:

A. SEQUENCE CHARACTERISTICS:

- A. LENGTH: 50 base pairs
- B. TYPE: nucleic acid
- C. STRANDEDNESS: single
- D. TOPOLOGY: linear

II. MOLECULE TYPE: cDNA

III. SEQUENCE DESCRIPTION: SEQ ID NO. 17:

AATTGTACCG CTTAGCCATT TGTGTGCTCT CTCTCATGGA TCCATCCCG GGT

1. INFORMATION FOR SEQ ID NO. 18:

A. SEQUENCE CHARACTERISTICS:

- A. LENGTH: 13 base pairs
- B. TYPE: nucleic acid
- C. STRANDEDNESS: single
- D. TOPOLOGY: linear

II. MOLECULE TYPE: cDNA

III. SEQUENCE DESCRIPTION: SEQ ID NO. 18:

CTGGGGGCTCC AGT

1. INFORMATION FOR SEQ ID NO. 19:

A. SEQUENCE CHARACTERISTICS:

- A. LENGTH: 17 base pairs
- B. TYPE: nucleic acid

1 STRANDEDNESS: single
2 TOPOLOGY: linear

11 MOLECULE TYPE: cDNA

X1 SEQUENCE DESCRIPTION: SEQ ID NO:19:

GTAGACTGCG GCGGCGAG

17

1 INFORMATION FOR SEQ ID NO:20:

1 SEQUENCE CHARACTERISTICS:
A LENGTH: 50 base pairs
B TYPE: nucleic acid
C STRANDEDNESS: single
D TOPOLOGY: linear

11 MOLECULE TYPE: cDNA

X1 SEQUENCE DESCRIPTION: SEQ ID NO:20:

GCAGGTCCAA CTCACGGTCC ACTCTAGAGG GCATTCTTTC TGGGCCCCAGG

50

1 INFORMATION FOR SEQ ID NO:21:

1 SEQUENCE CHARACTERISTICS:
A LENGTH: 50 base pairs
B TYPE: nucleic acid
C STRANDEDNESS: single
D TOPOLOGY: linear

11 MOLECULE TYPE: cDNA

X1 SEQUENCE DESCRIPTION: SEQ ID NO:21:

TATTATCTGT GCGGCGCGCGG TCCAGAACCG ATGGGTCTTG GTTCCACTCG

50

1 INFORMATION FOR SEQ ID NO:22:

1 SEQUENCE CHARACTERISTICS:
A LENGTH: 40 base pairs
B TYPE: nucleic acid
C STRANDEDNESS: single
D TOPOLOGY: linear

11 MOLECULE TYPE: cDNA

X1. SEQUENCE DESCRIPTION: SEQ ID NO:23:
TATTATCTGT CCCCCCCCCG AGGATGCTCC AGAGTGGTAG

1. INFORMATION FOR SEQ ID NO:23:

1. SEQUENCE CHARACTERISTICS:
A. LENGTH: 36 base pairs
B. TYPE: nucleic acid
C. STRANDEDNESS: single
D. TOPOLOGY: linear

11. MOLECULE TYPE: cDNA

X1. SEQUENCE DESCRIPTION: SEQ ID NO:23:
TATTATCTGT CCCCCCCCCG AGGATGCTCC TGTGGCAG

1. INFORMATION FOR SEQ ID NO:24:

1. SEQUENCE CHARACTERISTICS:
A. LENGTH: 33 base pairs
B. TYPE: nucleic acid
C. STRANDEDNESS: single
D. TOPOLOGY: linear

11. MOLECULE TYPE: cDNA

X1. SEQUENCE DESCRIPTION: SEQ ID NO:24:
CCGAGACAGG CCACCGAATT CCGCCAGAA ACC

1. INFORMATION FOR SEQ ID NO:25:

1. SEQUENCE CHARACTERISTICS:
A. LENGTH: 40 base pairs
B. TYPE: nucleic acid
C. STRANDEDNESS: single
D. TOPOLOGY: linear

11. MOLECULE TYPE: cDNA

X1. SEQUENCE DESCRIPTION: SEQ ID NO:25:
CCCGAGAAA TCGTGGGCA CAGCACTGAC ACCACTCTC

1. INFORMATION FOR SEQ ID NO:26:

1. SEQUENCE CHARACTERISTICS:
A. LENGTH: 40 base pairs
B. TYPE: nucleic acid
C. STRANDEDNESS: single
D. TOPOLOGY: linear

11. MOLECULE TYPE: cDNA

X1. SEQUENCE DESCRIPTION: SEQ ID NO:16:

GAGCTGGGCA ACATGGGGGA ACTGTCCAGC GATTCAGCAG

2. INFORMATION FOR SEQ ID NO:17:

1. SEQUENCE CHARACTERISTICS:
A. LENGTH: 21 base pairs
B. TYPE: nucleic acid
C. STRANDEDNESS: single
D. TOPOLOGY: linear

11. MOLECULE TYPE: cDNA

X1. SEQUENCE DESCRIPTION: SEQ ID NO:17:

AATTCGAGTT CCTAGATTTT G

2. INFORMATION FOR SEQ ID NO:18:

1. SEQUENCE CHARACTERISTICS:
A. LENGTH: 21 base pairs
B. TYPE: nucleic acid
C. STRANDEDNESS: single
D. TOPOLOGY: linear

11. MOLECULE TYPE: cDNA

X1. SEQUENCE DESCRIPTION: SEQ ID NO:18:

AATTCAAAAT CTAGGAATC G

2. INFORMATION FOR SEQ ID NO:19:

1. SEQUENCE CHARACTERISTICS:
A. LENGTH: 48 base pairs
B. TYPE: nucleic acid
C. STRANDEDNESS: single
D. TOPOLOGY: linear

11. MOLECULE TYPE: cDNA

K1 SEQUENCE DESCRIPTION: SEQ ID NO:29

AATTGAGTA CTGAGATTAT GATTTCCTTC CAGAAACTGA GCTTCCTCT

L INFORMATION FOR SEQ ID NO:30:

1. SEQUENCE CHARACTERISTICS:
 - A. LENGTH: 48 base pairs
 - B. TYPE: nucleic acid
 - C. STRANDEDNESS: single
 - D. TOPOLOGY: linear

11 MOLECULE TYPE: cDNA

K1 SEQUENCE DESCRIPTION: SEQ ID NO:30

GGCCCCGGAG GTCAGTTTC TGGCAGGAAA TCATAATCTA GGTACTCG

L INFORMATION FOR SEQ ID NO:31:

1. SEQUENCE CHARACTERISTICS:
 - A. LENGTH: 48 base pairs
 - B. TYPE: nucleic acid
 - C. STRANDEDNESS: single
 - D. TOPOLOGY: linear

11 MOLECULE TYPE: cDNA

K1 SEQUENCE DESCRIPTION: SEQ ID NO:31

AATTGAGTT CTGAGATTAT GATTTCCTTC CAGAAACTGA GCTTCCTCT

L INFORMATION FOR SEQ ID NO:32

1. SEQUENCE CHARACTERISTICS:
 - A. LENGTH: 48 base pairs
 - B. TYPE: nucleic acid
 - C. STRANDEDNESS: single
 - D. TOPOLOGY: linear

11 MOLECULE TYPE: cDNA

K1 SEQUENCE DESCRIPTION: SEQ ID NO:32

GGCCCCGGAG GTCAGTTTC TGGCAGGAAA TCATAATCTA GGTACTCG

1. SEQUENCE INFORMATION: SEQ ID NO:13

1. SEQUENCE CHARACTERISTICS
- A. LENGTH: 49 base pairs
 - B. TYPE: nucleic acid
 - C. STRANDEDNESS: single
 - D. TOPOLOGY: linear

11. MOLECULE TYPE: cDNA

12. SEQUENCE DESCRIPTION: SEQ ID NO:13

AACTTGAGATT GGTAGATTTC GATTTCCTGC CAGAAATCA GCTCTGCG

43

1. INFORMATION FOR SEQ ID NO:14

1. SEQUENCE CHARACTERISTICS
- A. LENGTH: 49 base pairs
 - B. TYPE: nucleic acid
 - C. STRANDEDNESS: single
 - D. TOPOLOGY: linear

11. MOLECULE TYPE: cDNA

12. SEQUENCE DESCRIPTION: SEQ ID NO:14

GGGCGGGAG GTCAGTTTC TGGCAGGAAA TCGAAATCA GCAACTTC

44

1. INFORMATION FOR SEQ ID NO:15

1. SEQUENCE CHARACTERISTICS
- A. LENGTH: 340 base pairs
 - B. TYPE: nucleic acid
 - C. STRANDEDNESS: double
 - D. TOPOLOGY: linear

11. MOLECULE TYPE: cDNA

12. FEATURE

- A. NAME KEY: CDS
- B. LOCATION: 1..339

12. SEQUENCE DESCRIPTION: SEQ ID NO:15

ATGCTCTGCG AACTGCTGCT GTTCTGATC CTACTGCGCG CTGCGAAGAG GTTGCAGCTC

50

TGGCAGACTT GGGCAGATCA AGGCGAGAAA GCTTTGGGTC GCTGCTTTGC GCGCGAAGCG

100

```

ATGAGAGGCGA TGAATATGCA TGAATTAAT TATGATTTTC TGTGAGAAAT TGAATTTGTA 10
TAAATTTGTA TGAAGAGGAG TGAGAGCACT GTGTGAGTG GGGGTGGAAG TGTGAGTTCT 20
AGGACTGTG AGGCTGCTGC GGGGGGGGAG AGATGGGAG GTTGGGAGG AGTTGAAAGT 30
TTGGGGGGAG GTTCACTGTT GTTCTTTGCG CCAAAACGCA AGGACAGGCT CATGATCTTC 40
TGGAGCGCTG AGGTGACATG GTTGGTGGTG GAGGTGAGCC AGGAAGAGCC TGAGGTCAAG 50
TTCAACTGCT ATGTGAGCGG GTTGGAGGTG CATAATGCCA AGACAAAGCC GCGGGAGGAG 60
TACTACAACA GGAGGTACCG TGTGGTGAGC GTCTGACCG TGTGACCA GAAGTGGCTG 70
AATGGCAAGG ATACAACTG CAAGGTCTTC AAGAAAGCCC TGGAGTCCC CATGAGAAAA 80
AGCATTTGCA AAGGCAAGG GGAGCGGCGA GAAGGACAGG TGTACAGCT GTTGGGATCC 90
TGGGAGGAGA TGACCAAGAA GGAGGTGAG GTGAGCTGCG TGGTCAAGG GTTGTATCCC 100
AGGACATCG GTTGGAGTG GGAGAGCAAT GGGGAGCGCG AGAACAACCA CAAGACCAAG 110
GTTCGGCTGC TGGACTCGCA GGGCTGCTTC TTCTCTATA GCAAGGTGAG GTTGGAGAA 120
AGCAGGTGGC AGCAGGGGAA GTTCTTTGCA TGTCTGCTGA TGCATGAGGC TGTGACAAAG 130
CACTACAGCG AGAAGAGGCT GTCTCTGTC GCGGGTAAAT GA 140

```

2. INFORMATION FOR SEQ ID NO:36:

1. SEQUENCE CHARACTERISTICS:

- A) LENGTH: 313 amino acids
- B) TYPE: amino acid
- C) STRANDEDNESS:
- D) TOPOLOGY: linear

2. MOLECULE TYPE: protein

3. SEQUENCE DESCRIPTION: SEQ ID NO:36

```

Met Pro Leu Gln Leu Leu Leu Leu Ile Leu Leu Gly Pro Gly Asp
1          5          10          15

Ser Leu Gln Leu Trp Asp Thr Trp Ala Asp Gln Ala Gln Cys Ala Leu
20          25          30

Gly Pro Leu Leu Ala Arg Asp Arg Arg Gln Ala Thr Gln Tyr Gln Tyr
35          40          45

Leu Asp Tyr Asp Phe Leu Pro Gln Thr Gln Pro Pro Gln Met Leu Arg
50          55          60

```

184
 185
 186
 187
 188
 189
 190
 191
 192
 193
 194
 195
 196
 197
 198
 199
 200
 201
 202
 203
 204
 205
 206
 207
 208
 209
 210
 211
 212
 213
 214
 215
 216
 217
 218
 219
 220
 221
 222
 223
 224
 225
 226
 227
 228
 229
 230
 231
 232
 233
 234
 235
 236
 237
 238
 239
 240
 241
 242
 243
 244
 245
 246
 247
 248
 249
 250
 251
 252
 253
 254
 255
 256
 257
 258
 259
 260
 261
 262
 263
 264
 265
 266
 267
 268
 269
 270
 271
 272
 273
 274
 275
 276
 277
 278
 279
 280
 281
 282
 283
 284
 285
 286
 287
 288
 289
 290
 291
 292
 293
 294
 295
 296
 297
 298
 299
 300
 301
 302
 303
 304
 305
 306
 307
 308
 309
 310
 311
 312
 313
 314
 315
 316
 317
 318
 319
 320
 321
 322
 323
 324
 325
 326
 327
 328
 329
 330
 331
 332
 333
 334
 335
 336
 337
 338
 339
 340
 341
 342
 343
 344
 345
 346
 347
 348
 349
 350
 351
 352
 353
 354
 355
 356
 357
 358
 359
 360
 361
 362
 363
 364
 365
 366
 367
 368
 369
 370
 371
 372
 373
 374
 375
 376
 377
 378
 379
 380
 381
 382
 383
 384
 385
 386
 387
 388
 389
 390
 391
 392
 393
 394
 395
 396
 397
 398
 399
 400
 401
 402
 403
 404
 405
 406
 407
 408
 409
 410
 411
 412
 413
 414
 415
 416
 417
 418
 419
 420
 421
 422
 423
 424
 425
 426
 427
 428
 429
 430
 431
 432
 433
 434
 435
 436
 437
 438
 439
 440
 441
 442
 443
 444
 445
 446
 447
 448
 449
 450
 451
 452
 453
 454
 455
 456
 457
 458
 459
 460
 461
 462
 463
 464
 465
 466
 467
 468
 469
 470
 471
 472
 473
 474
 475
 476
 477
 478
 479
 480
 481
 482
 483
 484
 485
 486
 487
 488
 489
 490
 491
 492
 493
 494
 495
 496
 497
 498
 499
 500
 501
 502
 503
 504
 505
 506
 507
 508
 509
 510
 511
 512
 513
 514
 515
 516
 517
 518
 519
 520
 521
 522
 523
 524
 525
 526
 527
 528
 529
 530
 531
 532
 533
 534
 535
 536
 537
 538
 539
 540
 541
 542
 543
 544
 545
 546
 547
 548
 549
 550
 551
 552
 553
 554
 555
 556
 557
 558
 559
 560
 561
 562
 563
 564
 565
 566
 567
 568
 569
 570
 571
 572
 573
 574
 575
 576
 577
 578
 579
 580
 581
 582
 583
 584
 585
 586
 587
 588
 589
 590
 591
 592
 593
 594
 595
 596
 597
 598
 599
 600
 601
 602
 603
 604
 605
 606
 607
 608
 609
 610
 611
 612
 613
 614
 615
 616
 617
 618
 619
 620
 621
 622
 623
 624
 625
 626
 627
 628
 629
 630
 631
 632
 633
 634
 635
 636
 637
 638
 639
 640
 641
 642
 643
 644
 645
 646
 647
 648
 649
 650
 651
 652
 653
 654
 655
 656
 657
 658
 659
 660
 661
 662
 663
 664
 665
 666
 667
 668
 669
 670
 671
 672
 673
 674
 675
 676
 677
 678
 679
 680
 681
 682
 683
 684
 685
 686
 687
 688
 689
 690
 691
 692
 693
 694
 695

INFORMATION FOR SEQ ID NO: 37

SEQUENCE CHARACTERISTICS

- [illegible]

11. MOLECULE TYPE: protein

12. FEATURE:

A. NAME KEY: CDS

B. LOCATION: 1..617

13. SEQUENCE DESCRIPTION: SEQ ID NO:17:

```

ATGCGTCTGCG AACTGCTGCT GTTCTGTGATG CTACTGGGGG CTGGCAACAG GTTGCACGTG      10
TGGGACACCT GGGCAGATGA AGCCGAGAAA GCGTTGGGTC GCGTCTTTCG CCGGGACGCG      20
AGACAGGGCA CCGAATATGA CTACCTAGAT TATGATTTTC TGGCAGAAAC CGAGCTGTCA      30
GAAATGCTGA GGAACAGGAC TGACACCACT GTTCTGACTG GCGCTGAAAC GCGTCACTCT      40
ACCACTGTGG AGCTGCTGTC GCGGCGGCTG TGTGCGCAAC TAGTACCGCT GCGCATCACC      50
AAGCGCACCG TCGACAGAT CACTGGCAAG TCGTTTTATA TCGCATCGCG GTTTCGAAAC      60
GAGGAGTACA ATAAGTCGCT TCAGGAGATC CAAGCAACCT TCTTTTACTT CACCGCGAAC      70
AAGACAGAGG ACACGATCTT TCTCAGAGAG TACCAGACCG GACAGGACCA GTGCATCTAT      80
AACACCACTT ACCTGAATGT CCGCGGGGAA AATGGGACCA TCTCCAGATA GCTGGGAGGC      90
CAAGAGCATT TCGCTCACTT GCTGATCTTC AGGGACACCA AGACCTACAT GCTTCTTTTT      100
GACGTGAACG ATGAGAAGAA CTGGGGGCTG TCTGTCTATG CTGACAAGCG AGAGAGGACC      110
AAGGAGGCAAC TGGGAGAGTT CTACGAAGCT CTGACTGCT TCGGCATTTC CAACTCAGAT      120
GTCTGTACA CCGATTGGAA AAAGGATAAG TGTGAGCGAC TCGAGAAGCA CTACGAGAAC      130
GAGAGGAAC AGGAGGAGGG CGAATCTTAG

```

1. INFORMATION FOR SEQ ID NO:16:

1. SEQUENCE CHARACTERISTICS:

(A) LENGTH: 169 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS:

(D) TOPOLOGY: linear

11. MOLECULE TYPE: protein

13. SEQUENCE DESCRIPTION: SEQ ID NO:16:

```

Met Pro Leu Gln Leu Leu Leu Leu Ile Leu Leu Gly Pro Gly Asn
 1         5         10         15

```

10 INFORMATION FOR SEQ ID NO: 29:

LENGTH: 1114 base pairs
TYPE: double strand
ORGANISM: double
TOPOLOGY: linear

-- MOLECULE TYPE : cDNA

1X FEATURE:

A) NAME KEY : IDS

B) LOCATION: 1..1311

(X1) SEQUENCE DESCRIPTION: SEQ ID NO:39:

```

ATGGTGGCCG GGACCCGCTG TCTTCTAGCG TTGCTGCTTC GCGAGGTGCT CCTGGGCGCC 60
GGGGCTGGCC TGCTTCCGGA GCTGGGCGCC AGGAAGTTCT GGGCGGCGTC GTGGGCGCGC 120
CCTCATCCCG AGCCCTCTGA CGAGGTGCTG AGCGAGTTCT AGTTGCGGCT GCTCAGCATG 180
TTGGGCGCTGA AACAGAGACC CACCCCGAGC AGGGACGCGG TGGTGCCCCC CTACATGCTA 240
GACCTGTATC GCAGGCACTC AGGTGAGCGG GGCTCAGCCG CCCCAGACCA CCGGTTGGAG 300
AGGGCAGCCA GCCGAGCCAA CACTGTGCGC AGCTTCCACC ATGAAGAATC TTTGGAAGAA 360
CTACGAGAAA CGAGTGGGAA AACAACCCGG AGATTCTTCT TTAATTTAAG TCTATCCGCC 420
ACGGAGGAGT TTATCACCTC AGCAGAGCTT CAGGTTTTTC GAGAACAGAT GCAAGATGCT 480
TTAGGAAACA ATAGCAGTTT CCATCACCAG ATTAATATTT ATGAAATCAT AAAACCTGCC 540
ACAGCCAACCT CGAAATTCCC CGTGACCAGA CTTTGAGACA CGAGGTGGGT GAATCAGAAT 600
GCAAGCAGGT GGGAAAGTTT TGATGTCACC CCGGCTGTGA TGGGTTGGAC TGCACAGGGA 660
CACGCCAACC ATGGATTCTG GGTGGAAGTG CCCCAGTTGG AGGAGAAACA AGGTCTCTCC 720
AAGAGACATG TTAGGATAAG CAGGTCTTTG CACCAAGATG AACACAGCTG CTCACAGATA 780
AGGCCATTGC TAGTAACTTT TGGCCATGAT GGAAAAGGGC ATCTCTCTCA CAAAACAGAA 840
AAACGTCAGG CCACCGAATA TGAGTACCTA GATTATGATT TCCTGCCAGA AACGGAGGCT 900
CCAGAAATGC TGAGGAACAG CACTGACACC ACTCCTCTGA CTGGGCGCTG AACCTCTGAG 960
TCTACCACTG TGGAGCCTGC TGCAAGGCGG AAACGCGCTA AGTCCAGCTG TAAGAGACAC 1020
CTTTTGTAGG TGGACTTCAG TGAGTGGGGT TGGAAATGACT CGATTGTGGC TTTTGTGGGG 1080
TATCAGCGCT TTTACTGCCA CGGAGAATGC CTTTTCTCTC TGGCTGATCA TGTGAAGTCC 1140
ACTAATCATG CCATTGTTCA GACGTTGGTC AACTCTGTGA ACTCTAAGAT TCTTAAGGCA 1200
TGCTGTGTCC CGACAGAACT CAGTGCTATC TCGATGCTGT ACCTTGACGA GAATGAAAAG 1260
GTGTATTAA AGAACTATCA GGACATGGTT GTGGAGGGTT GTGGGTCTCT CTAG 1311

```

(X2) INFORMATION FOR SEQ ID NO:40:

1 SEQUENCE CHARACTERISTICS
 A LENGTH: 216 amino acids
 B TYPE: single chain
 C STRANDEDNESS:
 D TOPOLOGY: linear

11 MOLECULE TYPE: protein

X1 SEQUENCE DESCRIPTION: SEQ ID NO:401

Met Val Ala Gly Thr Arg Lys Leu Leu Ala Leu Leu Leu Pro Gln Val
 1 5 10 15
 Leu Leu Gly Gly Ala Ala Gly Leu Val Pro Gln Leu Gly Arg Arg Lys
 20 25 30
 Phe Ala Ala Ala Ser Ser Gly Arg Pro Ser Ser Gln Pro Ser Asp Gln
 35 40 45
 Val Leu Ser Gln Phe Gln Leu Arg Leu Leu Ser Met Phe Gly Leu Lys
 50 55 60
 Gln Arg Pro Thr Pro Ser Arg Asp Ala Val Val Pro Pro Tyr Met Leu
 65 70 75 80
 Asp Leu Tyr Arg Arg His Ser Gly Gln Pro Gly Ser Pro Ala Pro Asp
 85 90 95
 His Arg Leu Gln Arg Ala Ala Ser Arg Ala Asn Thr Val Arg Ser Phe
 100 105 110
 His His Gln Gln Ser Leu Gln Gln Leu Pro Gln Thr Ser Gly Lys Thr
 115 120 125
 Thr Arg Arg Phe Phe Phe Asn Leu Ser Ser Ile Pro Thr Gln Gln Phe
 130 135 140
 Ile Thr Ser Ala Gln Leu Gln Val Phe Arg Gln Gln Met Gln Asp Ala
 145 150 155 160
 Leu Gly Asn Asn Ser Ser Phe His His Arg Ile Asn Ile Tyr Gln Ile
 165 170 175
 Ile Lys Pro Ala Thr Ala Asn Ser Lys Phe Pro Val Thr Arg Leu Leu
 180 185 190
 Asp Thr Arg Leu Val Asn Gln Asn Ala Ser Arg Trp Gln Ser Phe Asp
 195 200 205
 Val Thr Pro Ala Val Met Arg Trp Thr Ala Gln Gly His Ala Asn His
 210 215 220

```

Tyr Phe Val Val Glu Val Ala His Leu Glu Glu Ser Ala Glu Glu Leu
235          237          239          241
Cys Arg His Val Arg Ile Ser Arg Ser Leu His Glu Asp Glu His Ser
245          250          255
Tyr Ser Glu Ile Arg Pro Leu Leu Val Thr Phe Gly His Asp Glu Lys
260          265          270
Gly His Pro Leu His Lys Arg Glu Lys Arg Glu Ala Thr Glu Tyr Glu
275          280          285
Tyr Leu Asp Tyr Asp Phe Leu Pro Glu Thr Glu Pro Pro Glu Met Leu
291          295          300
Arg Asn Ser Thr Asp Thr Thr Pro Leu Thr Gly Pro Gly Thr Pro Glu
305          310          315          320
Ser Thr Thr Val Glu Pro Ala Ala Arg Arg Lys Arg Leu Lys Ser Ser
325          330          335
Cys Lys Arg His Pro Leu Tyr Val Asp Phe Ser Asp Val Gly Trp Asn
340          345          350
Asp Trp Ile Val Ala Pro Pro Gly Tyr His Ala Phe Tyr Cys His Gly
355          360          365
Glu Cys Pro Phe Pro Leu Ala Asp His Leu Asn Ser Thr Asn His Ala
370          375          380
Ile Val Glu Thr Leu Val Asn Ser Val Asn Ser Lys Ile Pro Lys Ala
385          390          395          400
Cys Cys Val Pro Thr Glu Leu Ser Ala Ile Ser Met Leu Tyr Leu Asp
405          410          415
Glu Asn Glu Lys Val Val Leu Lys Asn Tyr Glu Asp Met Val Val Glu
420          425          430
Gly Cys Gly Cys Arg
435

```

1. INFORMATION FOR SEQ ID NO:41:

```

SEQUENCE CHARACTERISTICS
A. LENGTH: 795 base pairs
B. TYPE: nucleic acid
C. STRANDEDNESS: double
D. TOPOLOGY: linear

```

11. MOLECULE TYPE: cDNA

12. FEATURE:

A. NAME KEY: CDS

B. SEQUENCE INFORMATION

X1 SEQUENCE DESCRIPTION: SEQ ID NO: 41

```

ATGCGCTGTGCG AAGTGGCTGCT GTTGGTGGATG CTACTGGGGCG CTGGCAACAG GTTGGAGGTG      10
TGGGACACCT GGGCAGATGA AGCCGAGAAA GGCTTGGGGTG CCTTGGTTGC GCGGGACGGG      20
AGACAGGGCA CGGAATATGA CTACCTAGAT TATGATTTGC TGGCAGAAAG CGAGTGTGCA      30
GAAATGCTGA GGAACAGCAC TGACACCACT CCTGTGACTG GGCTTGGAAAC CCTTGAGTGT      40
ACCACTGTGG AGCTGCTGCG GCGGGCGGCA GTTGGGGCGCG CTGGAGTTTC CCGAGACCTT      50
GGGGCGGAGG TGGACAGCAC GTTGGTGGTG AGCCGCTCTG TCGTGGCGGA CAGGGCGGAG      60
CTGGCTGGAC AGCTGAGGGA GAAATTTGCA GGTGACGGGG ACCACAACCT GGATTGCTTG      70
CGCACCTTGG CGATGAGTGC GCGGGGACTG GGAGCTCTAC AGCTCGGAGG TGTGGTGACA      80
AGGCTGGGAG GGGACCTACT GTCTTACCTG CGGCACGTGC AGTGGGTGGG CCGGGCAGGT      90
GGCTCTTTCC TGAAGACCTT GGAGCGCGAG CTGGGCACCC TGCAGGGCGCG ACTGGACGGG      100
CTGCTGGCGC GGCTGCAGCT CTTGATGTGC CGCCTGGGCG TGCCCCAGCC ACCCGCGGAC      110
CGCGCGGGCG CCGCGGTGGC GCGCGCTGC TGAGCCTGGG GGGGGATCAG GCGCGCGGAC      120
GCCATCTTGG GGGGGCTGCA CTTGACACTT GACTGGGCGG TGAGGGGACT GTTGGTGGTG      130
AAGACTCGGC TGTGA                                     135

```

C. INFORMATION FOR SEQ ID NO: 41:

- 1. SEQUENCE CHARACTERISTICS
 - A) LENGTH: 134 amino acids
 - B) TYPE: amino acid
 - C) STRANDEDNESS:
 - D) TOPOLOGY: linear

11. MOLECULE TYPE: protein

X2 SEQUENCE DESCRIPTION: SEQ ID NO: 42:

```

Met Pro Leu Gln Leu Leu Leu Leu Ile Leu Leu Gly Pro Gly Asn
1           5           10           15

Ser Leu Gln Leu Trp Asp Thr Trp Ala Asp Glu Ala Glu Lys Ala Leu
20           25           30           35

```

Gly Pro Leu Leu Ala Arg Asp Arg Arg Ala Ala Thr Val Thr Leu Thr
 35 40 45
 Leu Asp Tyr Asp Phe Leu Pro Gln Thr Gln Pro Val Gln Met Leu Arg
 50 55 60
 Asn Ser Thr Asp Thr Thr Pro Leu Thr Gly Pro Gly Thr Pro Gln Ser
 65 70 75 80
 Thr Thr Val Gln Pro Ala Ala Arg Pro Pro Pro Gly Pro Pro Arg Val
 85 90 95
 Ser Pro Asp Pro Arg Ala Gln Leu Asp Ser Thr Val Leu Leu Thr Arg
 100 105 110
 Ser Leu Leu Ala Asp Thr Arg Gln Leu Ala Ala Gln Leu Arg Asp Lys
 115 120 125
 Phe Pro Ala Asp Gly Asp His Asn Leu Asp Ser Leu Pro Thr Leu Ala
 130 135 140
 Met Ser Ala Gly Ala Leu Gly Ala Leu Gln Leu Pro Gly Val Leu Thr
 145 150 155 160
 Arg Leu Arg Ala Asp Leu Leu Ser Tyr Leu Arg His Val Gln Trp Leu
 165 170 175
 Arg Arg Ala Gly Gly Ser Ser Leu Lys Thr Leu Gln Pro Gln Leu Gly
 180 185 190
 Thr Leu Gln Ala Arg Leu Asp Arg Leu Leu Arg Arg Leu Gln Leu Leu
 195 200 205
 Met Ser Arg Leu Ala Leu Pro Gln Pro Pro Pro Asp Pro Pro Ala Pro
 210 215 220
 Pro Leu Ala Pro Pro Ser Ser Ala Trp Gly Gly Ile Arg Ala Ala His
 225 230 235 240
 Ala Ile Leu Gly Gly Leu His Leu Thr Leu Asp Trp Ala Val Arg Gly
 245 250 255
 Leu Leu Leu Leu Lys Thr Arg Leu
 260

CLAIMS

1. An isolated DNA encoding a fusion protein comprising: (a) a first amino acid sequence comprising amino acid 42 to amino acid 60 of SEQ ID NO 2; and (b) a second amino acid sequence derived from the sequence of a protein other than P-selectin ligand.
2. The DNA of claim 1 which further comprises an expression control sequence operably linked to said nucleotide sequence.
3. A host cell transformed with the DNA of claim 2.
4. A process for producing a fusion protein, which comprises:
 - (a) culturing the host cell of claim 3 under condition suitable for expression of the fusion protein; and
 - (b) purifying the fusion protein from the culture medium.
5. The DNA of claim 1 wherein said first amino acid sequence comprises amino acid 42 to amino acid 402 of SEQ ID NO:2.
6. The DNA of claim 1 wherein said first amino acid sequence comprises amino acid 42 to amino acid 310 of SEQ ID NO 2.
7. The DNA of claim 1 wherein said first amino acid sequence comprises amino acid 42 to amino acid 88 of SEQ ID NO 2.
8. The DNA of claim 1 wherein said first amino acid sequence comprises amino acid 42 to amino acid 118 of SEQ ID NO:2.
9. The DNA of claim 1 wherein said first amino acid sequence comprises amino acid 42 to amino acid 159 of SEQ ID NO 2.

10. The DNA of claim 1 wherein said second amino acid sequence is linked to the C-terminus of said first amino acid sequence.
11. The DNA of claim 10 wherein said sequences are linked by a linking sequence.
12. The DNA of claim 1 wherein said second amino acid sequence is joined to the N-terminus of said first amino acid sequence.
13. The DNA of claim 12 wherein said sequences are linked by a linking sequence.
14. The DNA of claim 1 wherein said second amino acid sequence is derived from a protein selected from the group consisting of an antibody, a cytokine, a growth factor, a differentiation factor, a hormone, an enzyme, a receptor or fragment thereof and a ligand.
15. The DNA of claim 14 wherein said second amino acid sequence is derived from the sequence of an antibody.
16. The DNA of claim 15 wherein said second amino acid sequence is derived from the Fc portion of an antibody.
17. The DNA of claim 15 wherein said second amino acid sequence is a mutation of a sequence derived from an antibody.
18. The DNA of claim 14 wherein said second amino acid sequence is derived from the sequence of a cytokine.
19. The DNA of claim 14 wherein said second amino acid sequence is derived from the sequence of a growth factor.
20. The DNA of claim 19 wherein said growth factor is a BMP.
21. The DNA of claim 7 comprising the nucleotide sequence of SEQ ID NO 15 from nucleotide 123 to nucleotide 939.

Fig. 1

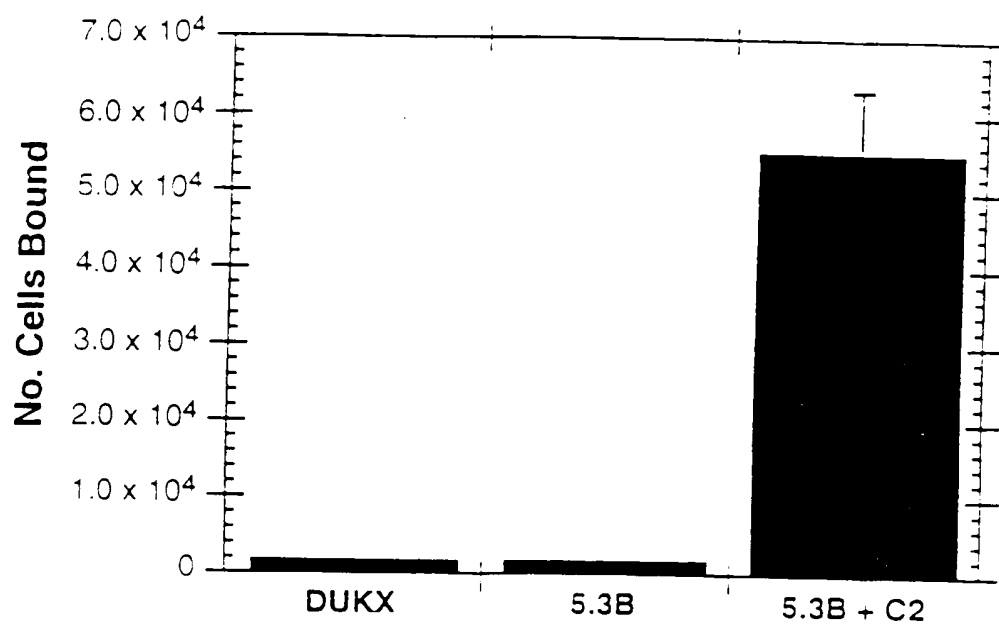


Fig. 2

poly PSL Ab		Lec γ 1	
-	+	-	+



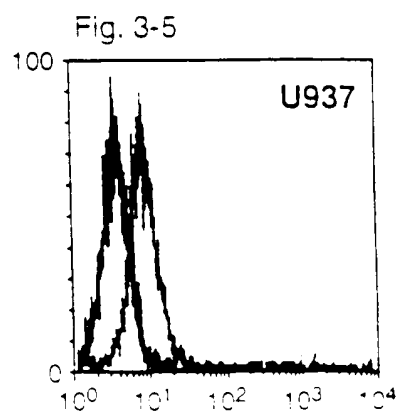
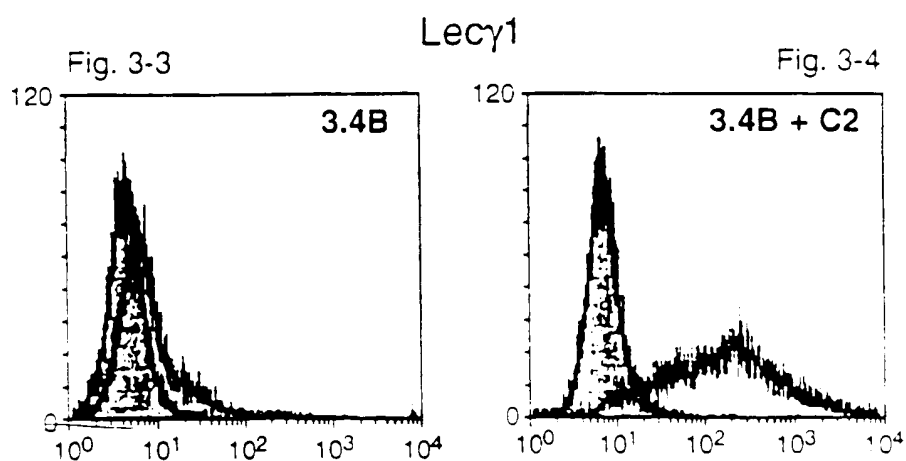
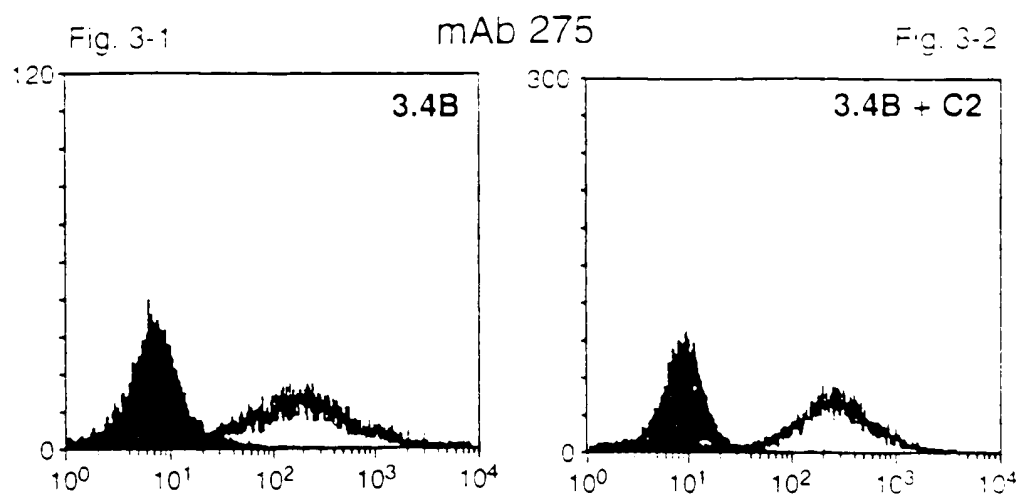


Fig. 4

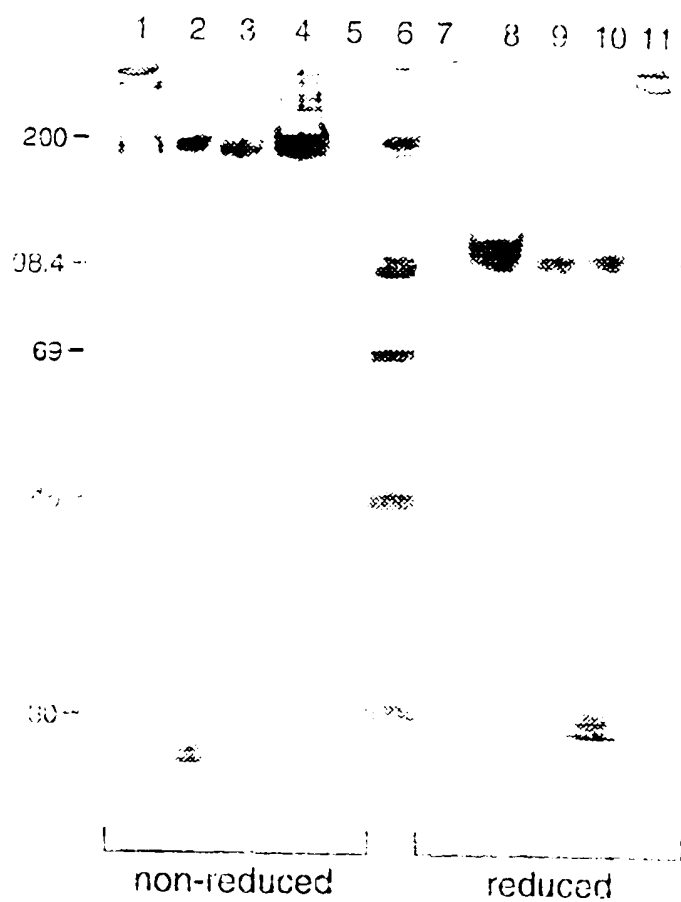


Fig. 5

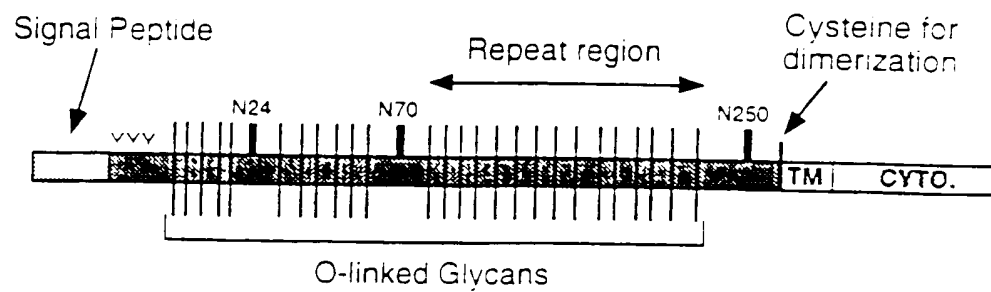
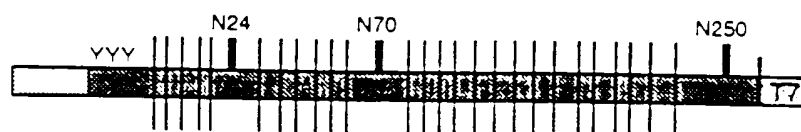
**Soluble "T7" Truncated Form:**

Fig. 6

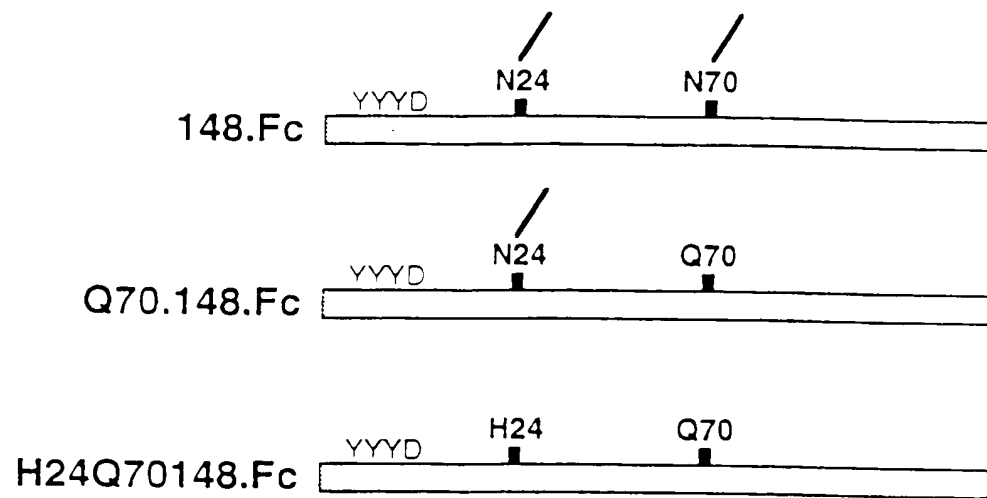


Fig. 7

1 2 3 4 5 6



³⁵S-Met Labeled

Captured with Protein A

1. 148.Fc
2. 148.Q70.Fc
3. 148.H24.Q70.Fc

Captured with P-Sel.Fc

4. 148.Fc
5. 18.Q70.Fc
6. 148.H24.Q70.Fc

Fig. 8

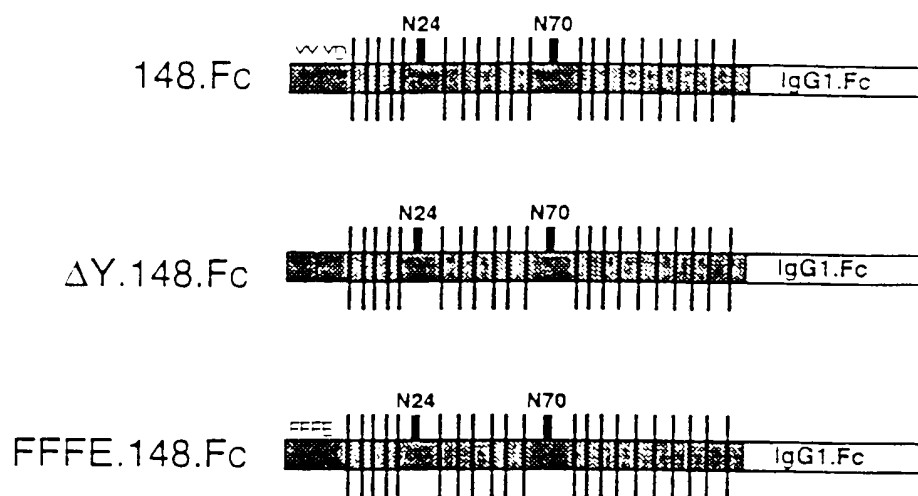
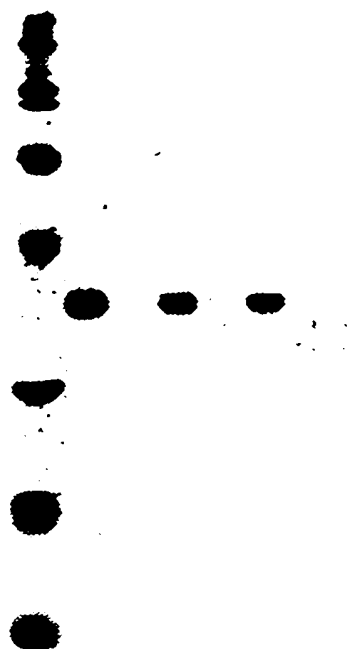


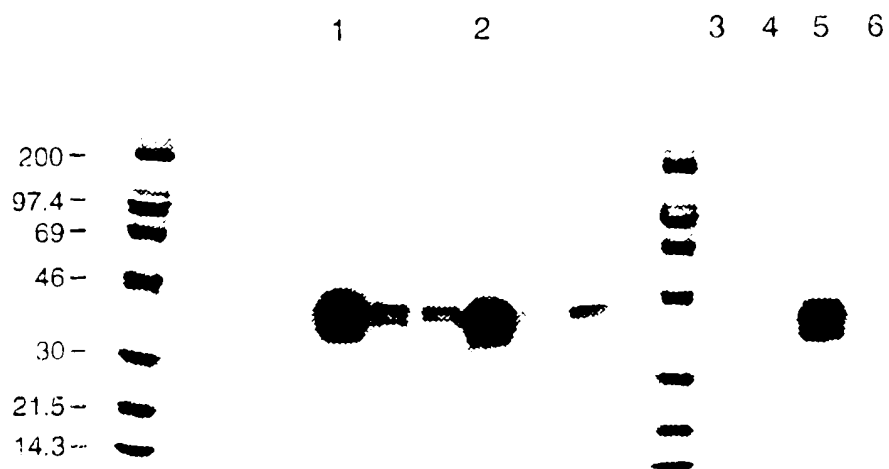
Fig. 9

M 1 2 3 4 5 6 7 8



	band	2.5 kb
M.	mw marker	
1.	YYY.19Fc	+
2.	YYY.19Fc	+
3.	FYY.19Fc	-
4.	FYY.19Fc	+
5.	FFY.19Fc	-
6.	FFY.19Fc	+
7.	FFF.19Fc	-
8.	FFF.19Fc	+

Fig. 10



35S-labeled protein

12.5% TBS

³⁵SO₄ Labeled

- 3. Mock
- 4. P-Sel.LE.Fc
- 5. FYY.19.Fc
- 6. FFF.19.Fc

Fig. 11

	³⁵ S-Met Labeled				³⁵ S-SO ₄ Labeled			
	[[
Chlorate	-	-	+	+	-	-	+	+
P-Sel.Fc	-	+	-	+	-	+	-	+
Protein A	+	-	+	-	+	-	+	-



Fig. 12

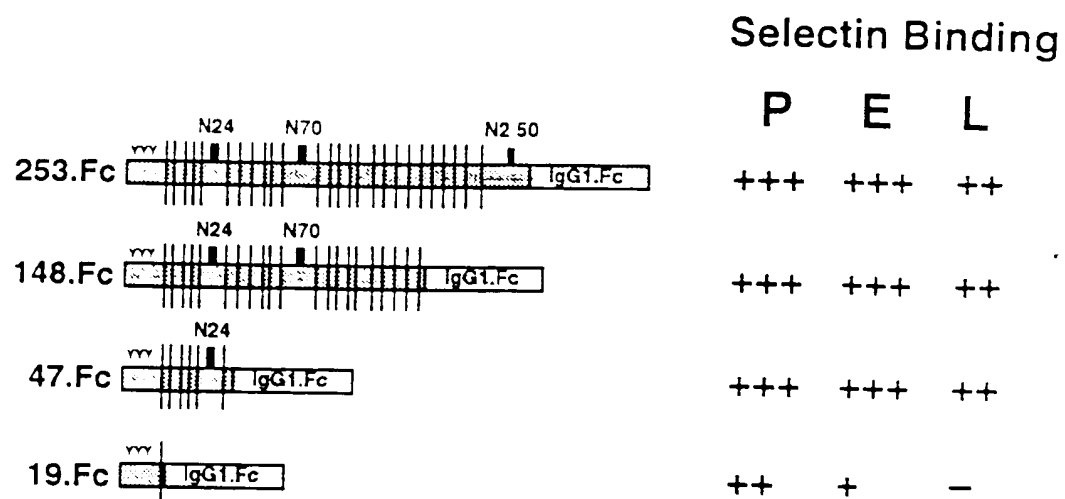


Fig. 14-1

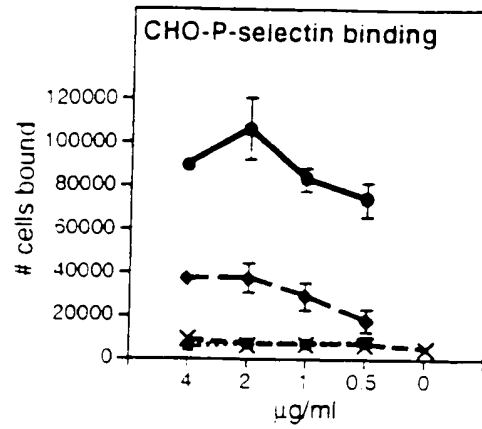


Fig. 14-2

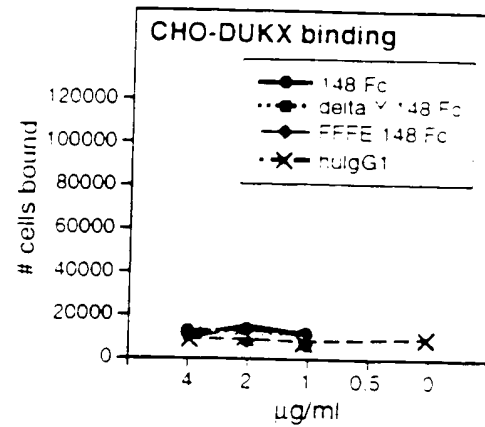


Fig. 15-1

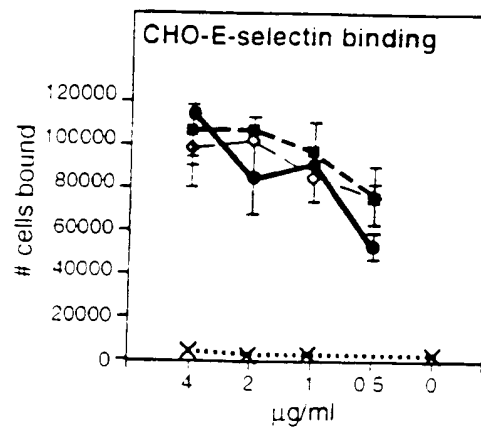


Fig. 15-2

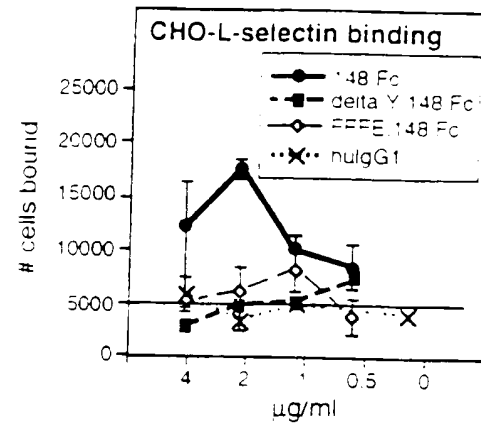


Fig. 16

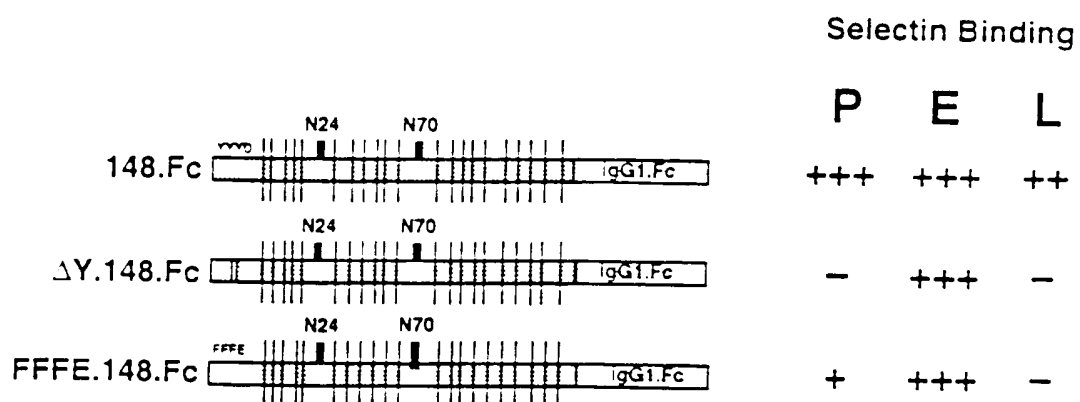


Fig. 17

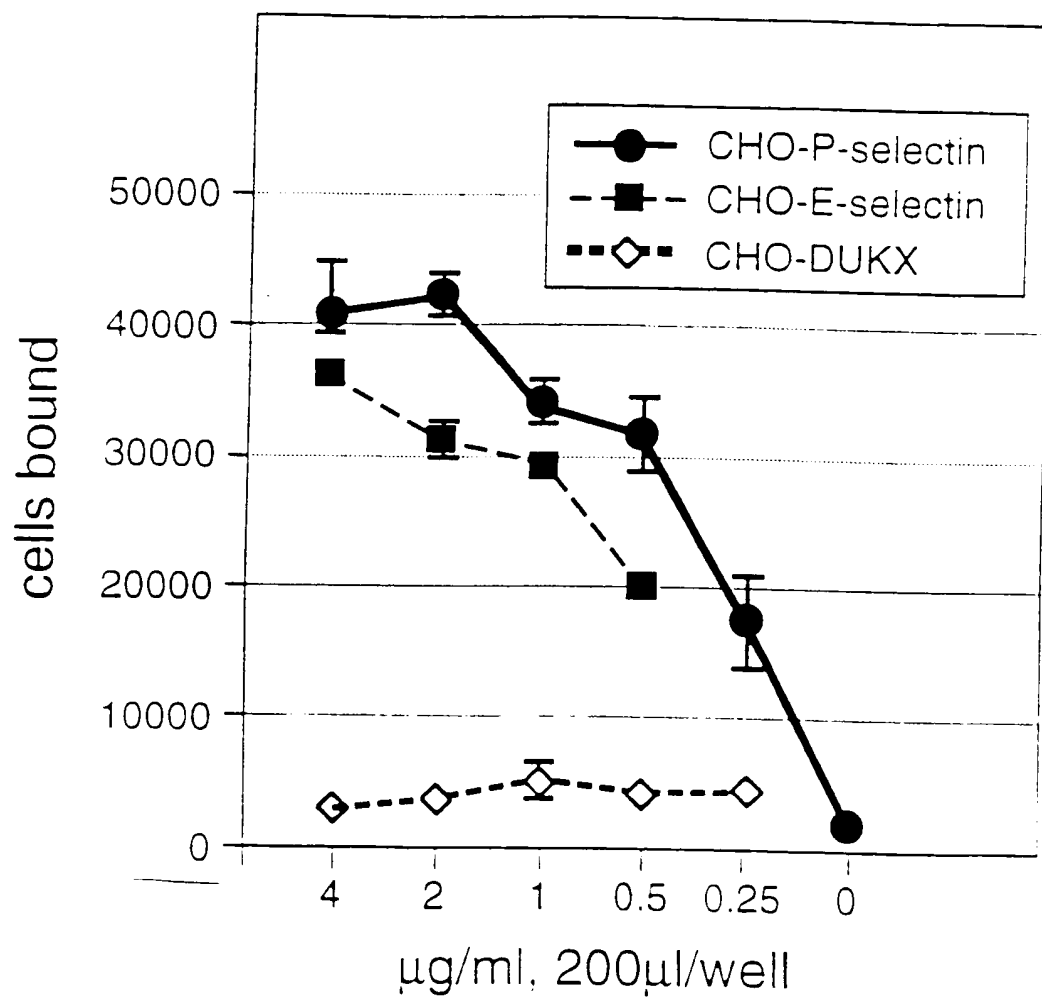
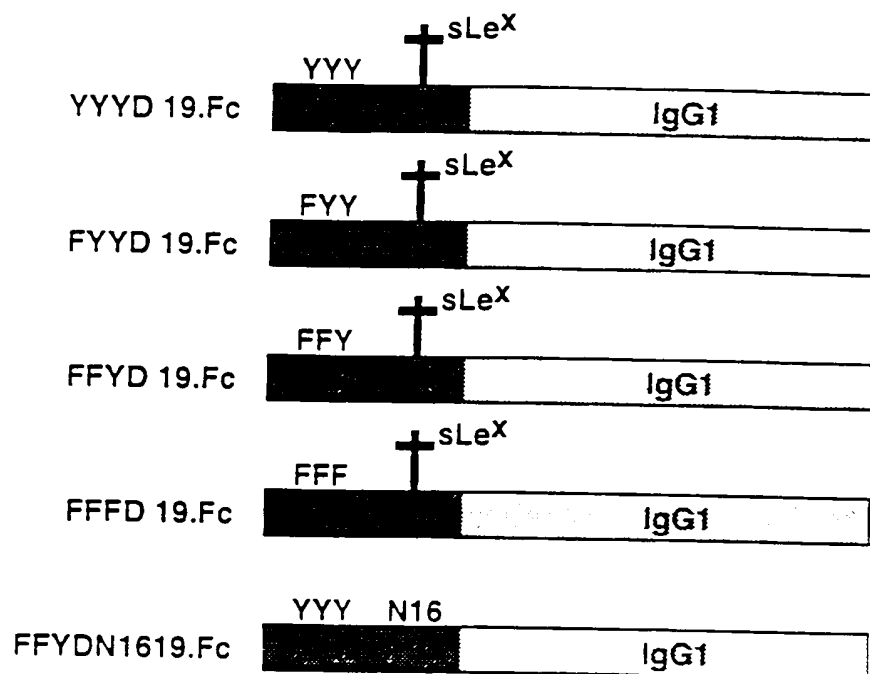


Fig. 18



† - O-linked carbohydrate at Thr16

Fig. 19-1

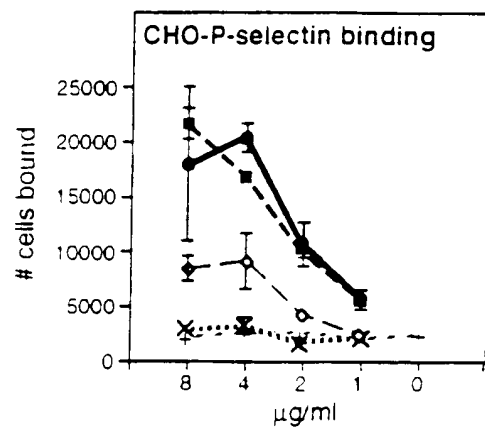


Fig. 19-2

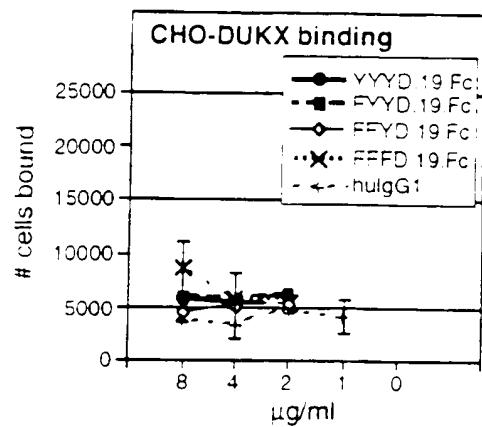


Fig. 20-1

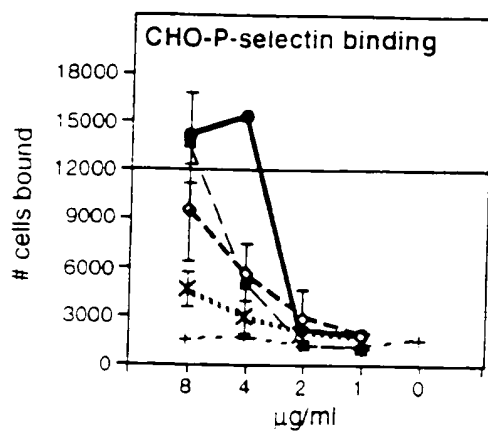


Fig. 20-2

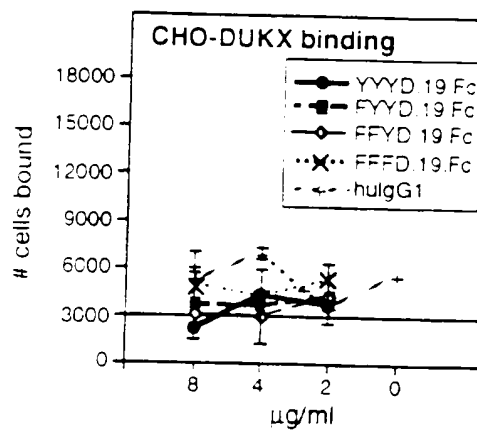


Fig. 21

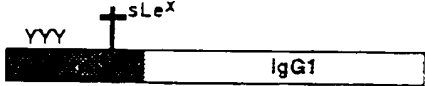
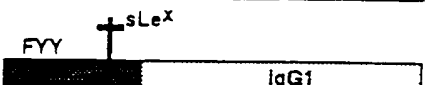
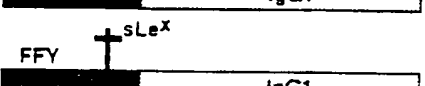
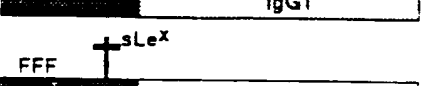
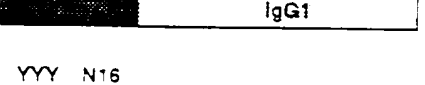
		Selectin Binding		
		P	E	L
YYYD 19.Fc		+++	+	-
FYYD 19.Fc		++	+	-
FFYD 19.Fc		+	+	-
FFFD 19.Fc		+	+	-
FFYDN1619.Fc		-	-	-

Fig. 22

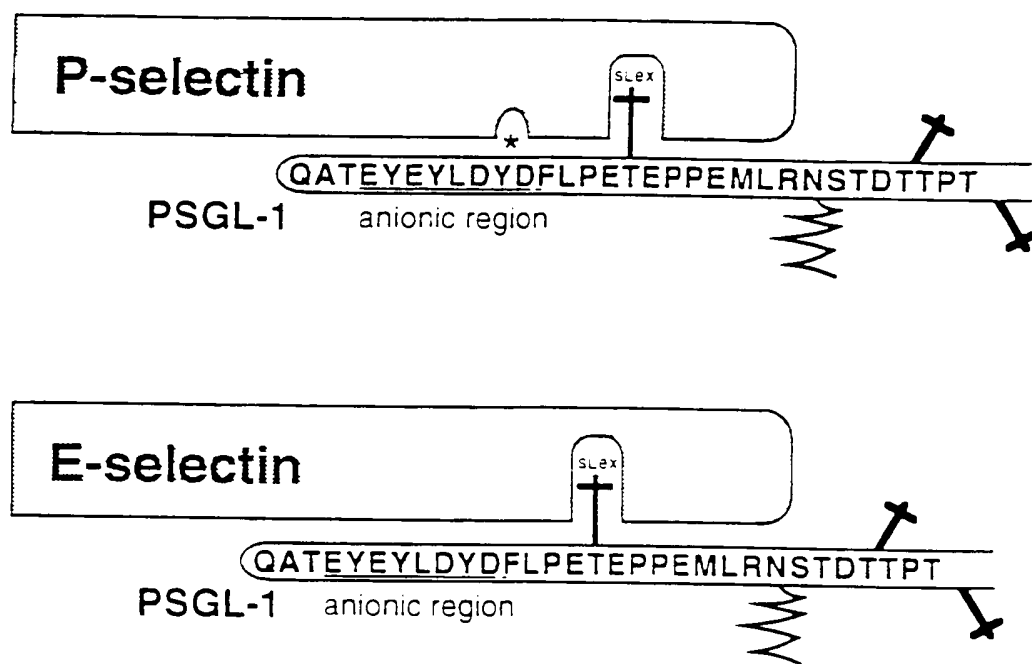


Fig. 23-1

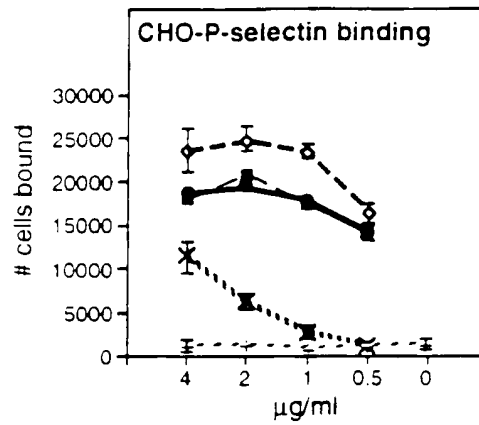


Fig. 23-2

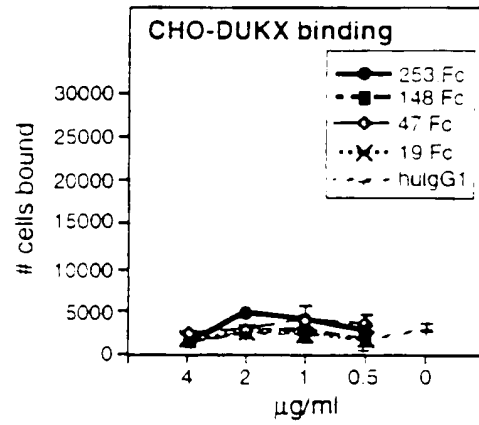


Fig. 24-1

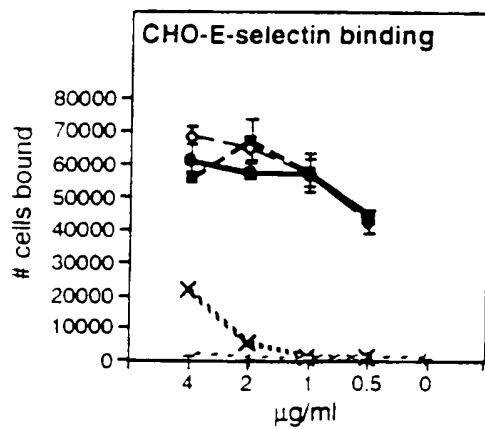


Fig. 24-2

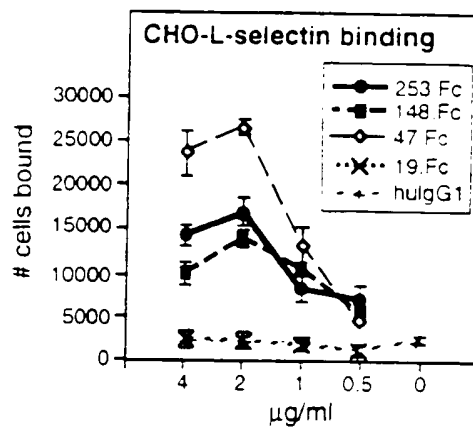
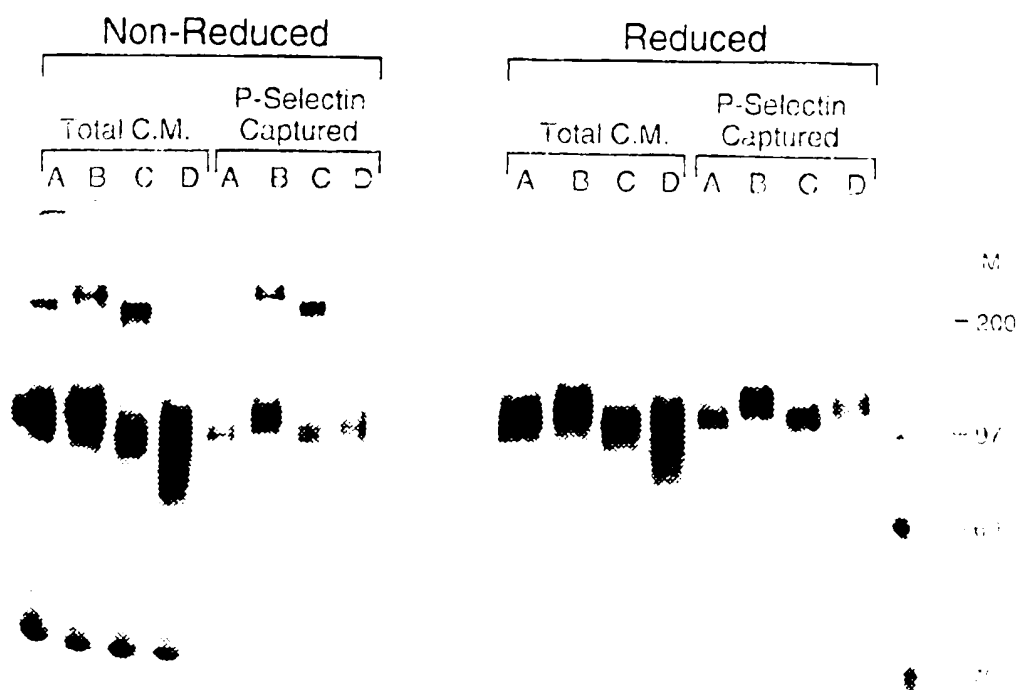


Fig. 25



Key:

A - "I7" sPSGL-1

B - "ΔTM" sPSGL-1

C - "I316" sPSGL-1

D - "Qc" sPSGL-1

Fig. 26

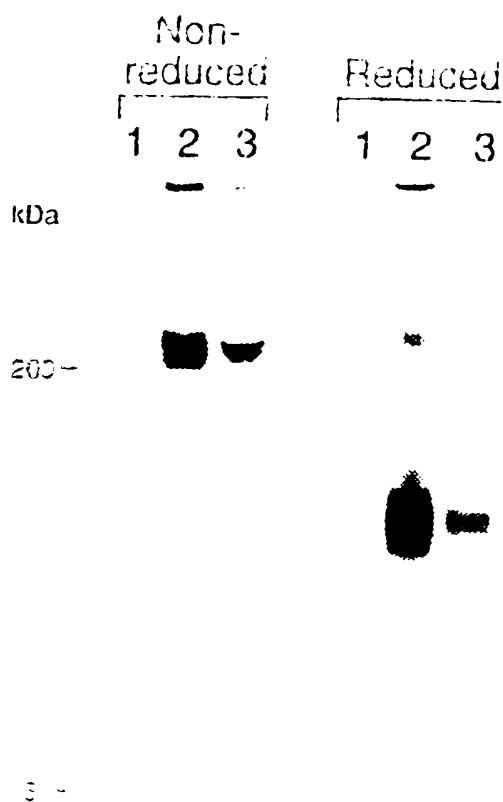


Fig. 27



Fig. 28

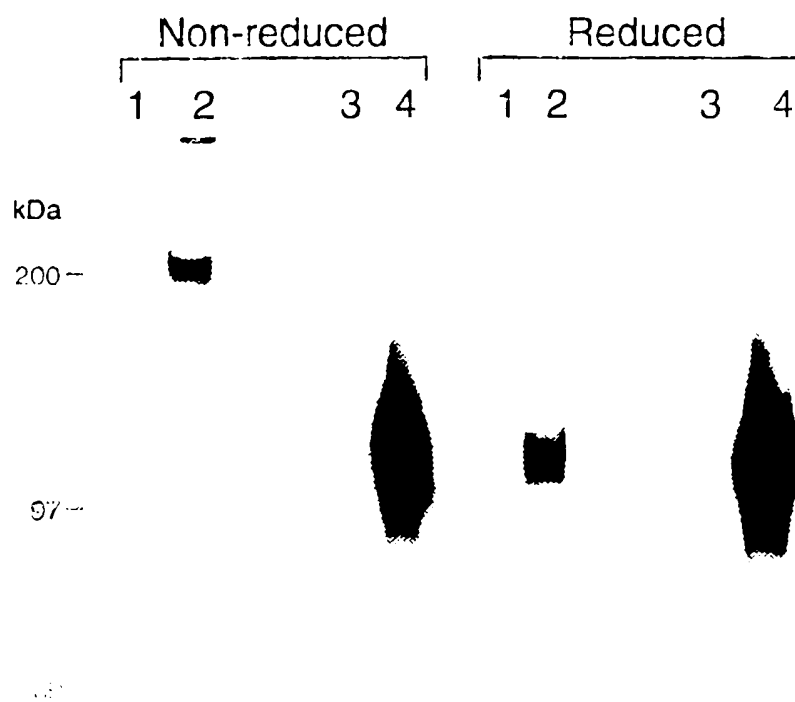


Fig. 29-1

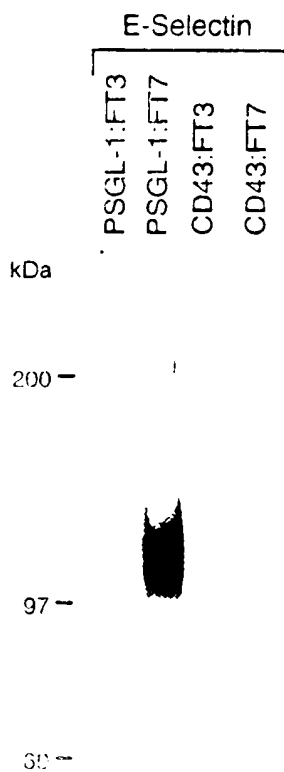


Fig. 29-2

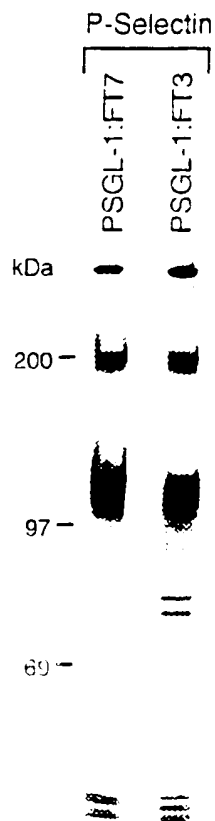
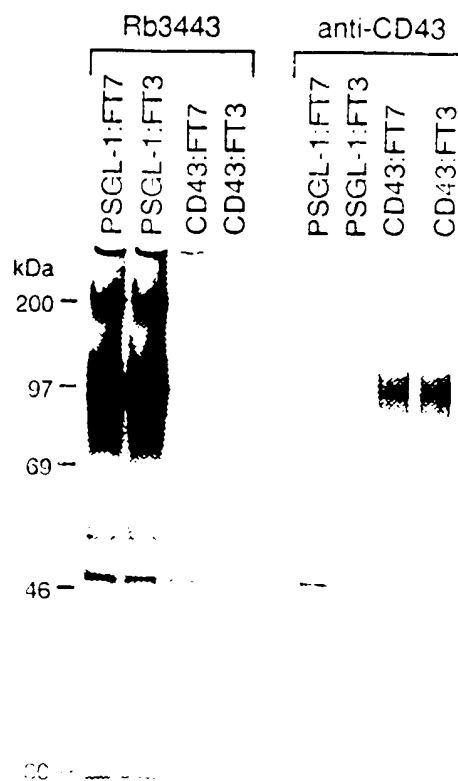


Fig. 29-3



anti-CD43

	PSGL-1:FT7	PSGL-1:FT3	CD43:FT7	CD43:FT3
kDa				
97				
69				

Fig. 30

	<u>E-selectin Inhibition</u>	<u>P-selectin Inhibition</u>
QATEY EYLDYDFLPEC	-	++ (~60 μ M)
TEY EYLDYDF		++
$\begin{array}{c} \text{PO}_3 \quad \text{PO}_3 \\ \quad \\ \text{SYLDYS} \end{array}$	+	+++ (~10 μ M)
$\begin{array}{c} \text{PO}_3 \\ \\ \text{SYLDYS} \end{array}$		+
SYLDYS		-
$\begin{array}{c} \text{PO}_3 \\ \\ \text{SF LDYS} \end{array}$		-
$\begin{array}{c} \text{PO}_3 \quad \text{PO}_3 \\ \quad \\ \text{AcY LDY NH}_2 \end{array}$		+
$\begin{array}{c} \text{PO}_3 \\ \\ \text{Ac LDY NH}_2 \end{array}$		-
$\begin{array}{c} \text{SO}_3 \\ \\ \text{SYLDYS} \end{array}$		-

WO 98/08949

PCT/US97/14159

5

P-SELECTIN LIGAND PROTEINS

15

BACKGROUND OF THE INVENTION

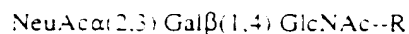
The present invention relates to the field of anti-inflammatory substances which act by inhibiting leukocyte adhesion to endothelial cells. More particularly, the present invention is directed to novel ligands for the mammalian adhesion proteins known as selectins.

20 During inflammation leukocytes adhere to the vascular endothelium and enter subendothelial tissue, an interaction which is mediated by specific binding of the selectin or LEC-CAM class of proteins to ligands on target cells. Such selectin-mediated cellular adhesion also occurs in thrombotic disorders and parasitic diseases and may be implicated in metastatic spread of tumor cells.

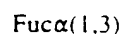
25 The selectin proteins are characterized by a N-terminal lectin-like domain, an epidermal growth factor-like domain, and regions of homology to complement binding proteins. Thus far three human selectin proteins have been identified, E-selectin (formerly ELAM-1), L-selectin (formerly LAM-1) and P-selectin (formerly PADGEM or GMP-140). E-selectin is induced on endothelial cells several hours after activation by cytokines, mediating the calcium-dependent interaction between neutrophils and the endothelium. L-selectin is the lymphocyte homing receptor, and P-selectin rapidly appears on the cell surface of platelets when they are activated, mediating calcium-dependent adhesion of neutrophils or monocytes to platelets. P-selectin is also found in the Weibel-Palade bodies of endothelial cells, upon its release from these vesicles P-selectin mediates early binding of neutrophils to histamine- or thrombin-stimulated endothelium.

35 Selectins are believed to mediate adhesion through specific interactions with ligands present on the surface of target cells. Generally the ligands of selectins are comprised at least in

5 part of a carbohydrate moiety. For example, E-selectin binds to carbohydrates having the terminal structure

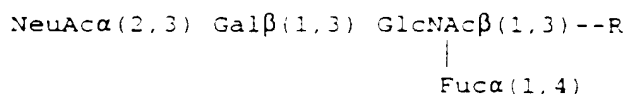


10



and also to carbohydrates having the terminal structure

15

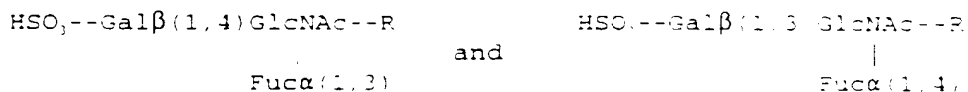


20

where R = the remainder of the carbohydrate chain. These carbohydrates are known blood group antigens and are commonly referred to as sialyl Lewis^x and sialyl Lewis^y, respectively. The presence of the sialyl Lewis^x antigen alone on the surface of an endothelial cell may be sufficient to promote binding to an E-selectin expressing cell. E-selectin also binds to carbohydrates having

25 the terminal structures

30



35 As with E-selectin, each selectin appears to bind to a range of carbohydrates with varying affinities. The strength of the selectin mediated adhesive event (binding affinity) may also depend on the density of the carbohydrate and on the density of the selectin on the cell surface.

P-selectin binds to carbohydrates containing the non-sialated form of the Lewis^x blood group antigen and with higher affinity to sialyl Lewis^x. P-selectin may also recognize sulfatides, which are heterogeneous 3-sulfated galactosyl ceramides, isolated from myeloid and tumor cells

40 by lipid extraction. However, the binding of cells bearing P-selectin to cells bearing P-selectin ligands is abolished when the ligand-bearing cells are treated with proteases, indicating that the P-selectin ligand may be a glycoprotein.

5 Two putative glycoprotein ligands for P-selectin have recently been identified, one of which has been partially purified, (Moore *et al.*, J. Cell Biol. 118, 445-456 (1992)). However, neither amino acid composition nor the amino acid sequence of these glycoproteins are disclosed

SUMMARY OF THE INVENTION

10 In one embodiment, the present invention provides a composition comprising an isolated DNA encoding a P-selectin ligand protein, said protein comprising the amino acid sequence set forth in SEQ ID NO:2 from amino acid 1 to amino acid 402. Also provided is a composition comprising an isolated DNA encoding a soluble P-selectin ligand protein, said protein comprising the amino acid sequence set forth in SEQ ID NO 2 from amino acid 1 to amino acid 310. The
15 invention further provides a composition comprising an isolated DNA encoding a mature P-selectin ligand protein, said protein comprising the amino acid sequence set forth in SEQ ID NO:2 from amino acid 42 to amino acid 402. In another embodiment, the invention provides a composition comprising an isolated DNA encoding a soluble mature P-selectin ligand protein, said protein comprising the amino acid sequence set forth in SEQ ID NO:2 from amino acid 42
20 to amino acid 310. In another embodiment, the invention provides a composition comprising an isolated DNA encoding a P-selectin ligand protein, said protein comprising the amino acid sequence set forth in SEQ ID NO:4. The invention further provides a composition comprising an expression vector comprising any one of the isolated DNAs of the invention, said DNA being operably linked to an expression control sequence; a host cell transformed with the expression
25 vector containing any one of the DNAs described above; and a process for producing the P-selectin ligand protein, which comprises:

(a) culturing a host cell transformed with an expression vector containing any one of the DNAs of the invention in a suitable culture medium; and

(b) purifying the P-selectin ligand protein from the culture medium.

30 In another embodiment, the invention provides a composition comprising a protein comprising the amino acid sequence set forth in SEQ ID NO 2 from amino acid 21 to amino acid 402, said protein being substantially free from other mammalian proteins. The invention further comprises a soluble P-selectin ligand protein comprising the amino acid sequence set forth in SEQ ID NO:2 from amino acid 21 to amino acid 310, said protein being substantially free from other
35 mammalian proteins. In another embodiment, the invention comprises a P-selectin ligand protein comprising the amino acid sequence set forth in SEQ ID NO:2 from amino acid 1 to amino acid 402, said protein being substantially free from other mammalian proteins. The invention also provides a composition comprising a mature P-selectin ligand protein comprising the amino acid

5 sequence set forth in SEQ ID NO:2 from amino acid 42 to amino acid 402, said protein being substantially free from other mammalian proteins. Further provided is a composition comprising a soluble mature P-selectin ligand protein comprising the amino acid sequence set forth in SEQ ID NO:2 from amino acid 42 to amino acid 310, said protein being substantially free from other mammalian proteins. In another embodiment, the invention provides a composition comprising
10 a protein comprising the amino acid sequence set forth in SEQ ID NO:4.

 In yet another embodiment, the invention provides compositions comprising antibodies specific for P-selectin ligand proteins.

 In another embodiment, the invention provides a method of identifying an inhibitor of P-selectin-mediated intercellular adhesion which comprises:

15 (a) combining a P-selectin protein with a P-selectin ligand protein comprising an amino acid sequence selected from the group consisting of the amino acid sequence set forth in SEQ ID NO:2 from amino acid 1 to amino acid 402, the amino acid sequence set forth in SEQ ID NO:2 from amino acid 42 to amino acid 402, the amino acid sequence set forth in SEQ ID NO:2 from amino acid 42 to amino acid 310, and the amino acid sequence set forth in SEQ ID
20 NO:4, said combination forming a first binding mixture;

 (b) measuring the amount of binding between the P-selectin protein and the P-selectin ligand protein in the first binding mixture;

 (c) combining a compound with the P-selectin protein and the P-selectin ligand protein to form a second binding mixture;

25 (d) measuring the amount of binding in the second binding mixture; and

 (e) comparing the amount of binding in the first binding mixture with the amount of binding in the second binding mixture;

 wherein the compound is capable of inhibiting P-selectin-mediated intercellular adhesion when a decrease in the amount of binding of the second binding mixture occurs.

30 In another embodiment, the invention provides a method of identifying an inhibitor of E-selectin-mediated intercellular adhesion which comprises:

 (a) combining a E-selectin protein with a P-selectin ligand protein comprising an amino acid sequence selected from the group consisting of the amino acid sequence set forth in SEQ ID NO:2 from amino acid 1 to amino acid 402, the amino acid sequence set forth in SEQ
35 ID NO:2 from amino acid 42 to amino acid 402, the amino acid sequence set forth in SEQ ID NO:2 from amino acid 42 to amino acid 310, and the amino acid sequence set forth in SEQ ID NO:4, said combination forming a first binding mixture;

- 5 (b) measuring the amount of binding between the E-selectin protein and the P-selectin ligand protein in the first binding mixture;
- (c) combining a compound with the E-selectin protein and the P-selectin ligand protein to form a second binding mixture;
- (d) measuring the amount of binding in the second binding mixture; and
- 10 (e) comparing the amount of binding in the first binding mixture with the amount of binding in the second binding mixture;

wherein the compound is capable of inhibiting E-selectin-mediated intercellular adhesion when a decrease in the amount of binding of the second binding mixture occurs. These methods may also be used to look for L-selectin inhibitors by substituting L-selectin for E-selectin.

- 15 The invention also encompasses processes for producing P-selectin ligand proteins which comprise (a) co-transforming a host cell with a DNA encoding a P-selectin ligand protein and a DNA encoding a fucosyltransferase capable of synthesizing sialyl Lewis X (sLe^x) or sialyl Lewis A (sLe^a) (such as an (α 1,3/ α 1,4) fucosyltransferase or an (α 1, 3) fucosyltransferase), each of said DNAs being operably linked to an expression control sequence; (b) culturing the host cell in
- 20 suitable culture medium; and (c) purifying the P-selectin ligand protein from the culture medium. In certain other embodiments, the host cell is also co-transformed with a DNA encoding a paired basic amino acid converting enzyme and/or a DNA encoding a GlcNAc transferase (preferably a "core2 transferase"). In preferred embodiments, the P-selectin ligand protein is a full-length or soluble form.

- 25 In other embodiments, the present invention includes a P-selectin ligand protein having P-selectin ligand protein activity. In preferred embodiments, the ligand protein is a protein comprising the sequence from amino acid 42 to amino acid 60 of SEQ ID NO. 2, consisting essentially of the sequence from amino acid 42 to amino acid 60 of SEQ ID NO. 2, comprising the sequence from amino acid 42 to amino acid 88 of SEQ ID NO. 2, consisting essentially of the
- 30 sequence from amino acid 42 to amino acid 88 of SEQ ID NO. 2, consisting essentially of the sequence from amino acid 42 to amino acid 118 of SEQ ID NO. 2, or consisting essentially of the sequence from amino acid 42 to amino acid 189 of SEQ ID NO. 2. In other preferred embodiments, at least one of the asparagine residues at positions 65, 111 and 292 of SEQ ID NO. 2 have been deleted or replaced. Certain preferred embodiments of the ligand protein comprises
- 35 at least one of the tyrosine residues at positions 46, 48 and 51 of SEQ ID NO. 2. DNAs encoding these P-selectin ligand proteins, host cells transformed with such DNAs, process for producing protein by culturing such host cells, pharmaceutical compositions comprising the proteins, methods of identifying selectin binding inhibitors using the proteins, antibodies to the proteins

5 and methods of inhibiting selectin mediated binding using the proteins are also encompassed by the invention

In yet other embodiments the invention provides an isolated DNA encoding a fusion protein comprising (a) a first amino acid sequence comprising amino acid 42 to amino acid 60 of SEQ ID NO:2, and (b) a second amino acid sequence derived from the
10 sequence of a protein other than P-selectin ligand. Preferably, an expression control sequence is operably linked to the nucleotide sequence. Host cells transformed with such DNAs are also provided. The invention also provides a process for producing a fusion protein, which comprises: (a) culturing the host cell under condition suitable for expression of the fusion protein; and (b) purifying the fusion protein from the culture
15 medium. Fusion proteins produced according to such process are also provide.

In certain preferred embodiments, the first amino acid sequence of such fusion protein comprises amino acid 42 to amino acid 402 of SEQ ID NO:2, amino acid 42 to amino acid 310 of SEQ ID NO:2, amino acid 42 to amino acid 88 of SEQ ID NO:2, amino acid 42 to amino acid 118 of SEQ ID NO:2, or amino acid 42 to amino acid 189
20 of SEQ ID NO:2.

In other preferred embodiments, the DNA comprises the nucleotide sequence of SEQ ID NO:35 from nucleotide 123 to nucleotide 939, the nucleotide sequence of SEQ ID NO:35, the nucleotide sequence of SEQ ID NO:37 from nucleotide 123 to nucleotide 807, the nucleotide sequence of SEQ ID NO:37, the nucleotide sequence of SEQ ID
25 NO:39 from nucleotide 123 to nucleotide 1311, the nucleotide sequence of SEQ ID NO:39, the nucleotide sequence of SEQ ID NO:41 from nucleotide 123 to nucleotide 792, or the nucleotide sequence of SEQ ID NO:41.

The present invention also provides a fusion protein comprising (a) a first amino acid sequence comprising amino acid 42 to amino acid 60 of SEQ ID NO:2, and (b) a
30 second amino acid sequence derived from the sequence of a protein other than P-selectin ligand. Preferably, the first amino acid sequence comprises amino acid 42 to amino acid 402 of SEQ ID NO:2, amino acid sequence comprises amino acid 42 to amino acid 310 of SEQ ID NO:2, amino acid 42 to amino acid 88 of SEQ ID NO:2, amino acid 42 to amino acid 118 of SEQ ID NO:2, or amino acid 42 to amino acid 189 of SEQ ID NO:2.

35 In certain particularly preferred embodiments, the fusion protein comprises the amino acid sequence of SEQ ID NO:36 from amino acid 42 to amino acid 313, the

5 amino acid sequence of SEQ ID NO:36, the amino acid sequence of SEQ ID NO:38 from amino acid 42 to amino acid 269, the amino acid sequence of SEQ ID NO:38, the amino acid sequence of SEQ ID NO:40 from amino acid 42 to amino acid 437, the amino acid sequence of SEQ ID NO:40, the amino acid sequence of SEQ ID NO:42 from amino acid 42 to amino acid 264, or the amino acid sequence of SEQ ID NO:42.

10 In other preferred embodiments, the second amino acid sequence is linked to the C-terminus or the N-terminus of the first amino acid sequence, with or without being linked by a linking sequence.

In yet other embodiments, the second amino acid sequence is derived from a protein selected from the group consisting of an antibody, a cytokine, a growth factor, a differentiation factor, a hormone, an enzyme, a receptor or fragment thereof and a ligand. Preferably, the second amino acid sequence is derived from the sequence of an antibody, from the Fc portion of an antibody, or is a mutation of a sequence derived from an antibody.

20 In yet further embodiments, the present invention provides for a composition comprising (a) a first peptide comprising amino acid 42 to amino acid 60 of SEQ ID NO:2, and (b) a second peptide derived from the sequence of a protein other than P-selectin ligand, wherein the first peptide and the second peptide are chemically linked by a moiety other than a peptide bond. Any P-selectin ligand protein of the invention can be used in such a composition.

25 BRIEF DESCRIPTION OF THE FIGURES

Fig. 1 is a graph comparing the binding of P-selectin ligand proteins expressed with and without core2.

Fig. 2 is an autoradiograph of immunoprecipitations of P-selectin ligand protein expressed with and without core2.

30 Fig. 3 depicts the results of flow cytometry analysis of the binding of P-selectin ligand protein (expressed with and without core2) to P-selectin/IgG chimera (LEC-γ1) and anti-P-selectin ligand protein monoclonal antibody (MAb 275).

Fig. 4 is an autoradiograph of proteins, including P-selectin ligand protein, which bound to P- and E-selectin/IgG chimeras.

35 Fig. 5 is a schematic representation of structural features of the full length P-selectin ligand protein of SEQ ID NO: 2

5 Fig. 6 is a schematic representation of several P-selectin ligand protein fragments constructed for the purpose of examining the role of N-linked glycosylation sites in binding of the P-selectin ligand proteins to selectins.

 Fig. 7 depicts the results of experiments to determine the role of N-linked glycosylation sites in binding of the P-selectin ligand proteins to selectins.

10 Fig. 8 is a schematic representation of several P-selectin ligand protein fragments constructed for the purpose of examining the role of sulfated tyrosine residues in binding of the P-selectin ligand proteins to selectins.

 Figs. 9-11 depicts the results of experiments to determine the role of sulfated tyrosine residues in binding of the P-selectin ligand proteins to selectins.

15 Fig. 12 is a schematic representation of several P-selectin ligand protein fragments constructed for the purpose of examining the effects of various deletions on the binding of the P-selectin ligand proteins to selectins.

 Fig. 13 is a schematic depiction of the quantitative plate binding assay of Example 4(c)

20 Figs. 14-17 depict the results of experiments comparing the binding of various deleted and altered P-selectin ligand proteins to selectins.

 Fig. 18 is a schematic representation of several P-selectin ligand protein fragments constructed for the purpose of examining the effects of alteration of tyrosine residues in the anionic region of the P-selectin ligand proteins on selectin binding.

25 Figs. 19-21 depict the results of experiments comparing the binding of various deleted and altered P-selectin ligand proteins to selectins.

 Fig. 22 depicts a proposed model for binding of P-selectin ligand proteins to P- and E-selectin.

 Figs. 23 and 24 depict the results of experiments comparing the binding of various deleted and altered P-selectin ligand proteins to selectins.

30 Some of the foregoing figures employ a convention for numbering amino acids within the depicted constructs which is different than the residue numbering employed in SEQ ID NO:2. In the figures, residues are numbered using the first amino acid of soluble mature P-selectin ligand as a starting point. Hence, the residue numbers used in the figures are 41 less than those of SEQ ID NO:2. For example, residue 19 in the figures corresponds to residue 60 in SEQ ID NO:2.

35 Fig. 25 is an analysis of the expression products of CHO cells, already expressing 3/4 fucosyltransferase and Core2 transferase, which were transfected with psPSL.T7, ΔTM, 1316 or psPSL.QC and amplified using methotrexate. Conditioned media was either analyzed directly or

5 first precipitated with LEC- γ I and then analyzed by SDS-PAGE under non-reducing and reducing conditions

Fig. 26. SDS-PAGE separation of myeloid cell membrane proteins affinity captured by P-and E-selectin. Membrane lysates were prepared from U937 cells metabolically labeled with 3 H-glucosamine and subjected to affinity precipitation with immobilized P-and E-selectin and control human IgG₁. Eluted proteins were treated with ("reduced") or without ("non-reduced") DTT prior to gel electrophoresis. Lanes: 1, affinity capture by human IgG₁; 2, affinity capture by P-selectin; 3, affinity capture by E-selectin.

Fig. 27. Sequential affinity capture experiments. 3 H-labeled U937 lysate species were affinity captured by P-or E-selectin, eluted, and then subjected to immunoprecipitation with anti-PSGL-1 antiserum Rb3443. Lanes: 1 and 2, control immunoprecipitations of fresh myeloid cell membrane lysates using pre-immune rabbit serum (lane 1) and Rb3443 (lane 2); 3-5, immunoprecipitation with Rb3443 of myeloid cell membrane lysates previously affinity captured and eluted from P-selectin (lane 3), E-selectin (lane 4), and human IgG₁ (lane 5).

Fig. 28. Comparison of CD43 and PSGL-1 content of myeloid cell membrane extracts. Labeled U937 cell extracts were immunoprecipitated with anti-PSGL-1 rabbit polyclonal antibody Rb3443 or an anti-CD43 mouse MAb and then subjected to SDS-PAGE/autoradiography. Lanes: 1, immunoprecipitation with control pre-immune rabbit serum; 2, immunoprecipitation with Rb3443; 3, immunoprecipitation with control isotype-matched mouse antibody; 4, immunoprecipitation with anti-CD43 antibody.

Fig. 29. COS transfection experiments. COS M6 cells transfected with plasmids encoding PSGL-1 or CD43 as well as Fuc-TIII or Fuc-TVII were metabolically labeled with 35 S-methionine, and membranes were prepared for affinity capture experiments as described in Materials and Methods. The cDNAs employed in the transfections are indicated above the lanes. Precipitations were performed using (A) E-selectin, (B) P-selectin, and (C) anti-PSGL-1 antiserum Rb3443 and anti-CD43 MAb.

Fig. 30 summarizes the results of screening of various P-selectin ligand proteins for inhibition of P- and E-selectin binding (see Example 13).

DETAILED DESCRIPTION OF THE INVENTION

The present inventors have for the first time identified and isolated a novel DNA which encodes a protein which acts as a ligand for P-selectin on human endothelial cells and platelets. The sequence of the DNA is set forth in SEQ ID NO 1. The complete amino acid sequence of the P-selectin ligand protein (i.e., the mature peptide plus the leader sequence) is characterized by the amino acid sequence set forth in SEQ ID NO 2 from amino acid 1 to amino acid 402.

The P-selectin ligand of the invention is a glycoprotein which may contain one or more of the following terminal carbohydrates:



• •

5 ID NO:2) is a homodimeric protein having an apparent molecular of 220 kD as shown by non-reducing SDS-polyacrylamide gel electrophoresis.

The structure of the full-length P-selectin ligand protein is schematically represented in Fig. 5. Three regions of the P-selectin ligand protein of SEQ ID NO:2 are: an extracellular domain (from about amino acid 21 to 310 of SEQ ID NO:2), a transmembrane domain (from
10 about amino acid 311 to 332 of SEQ ID NO:2), and an intracellular, cytoplasmic domain (from about amino acid 333 to 402 of SEQ ID NO:2). The extracellular domain contains three consensus tripeptide sites (Asn-X-Ser/Thr) of potential N-linked glycosylation beginning at Asn residues 65, 111, and 292. The extracellular domain further contains three potential sites of tyrosine sulfation at residues 46, 48, and 51. The region comprised of residues 55-267 contains
15 a high percentage of proline, serine, and threonine including a subdomain of fifteen decameric repeats of the ten amino acid consensus sequence Ala-Thr/Met-Glu-Ala-Gln-Thr-Thr-X-Pro/Leu-Ala/Thr, wherein X can be either Pro, Ala, Gln, Glu, or Arg. Regions such as these are characteristic of highly O-glycosylated proteins.

COS or CHO cells co-transfected with a gene encoding the P-selectin ligand protein and
20 a gene encoding fucosyltransferase (hereinafter FT), preferably an (α 1.3/ α 1.4) fucosyltransferase ("3/4FT"), are capable of binding to CHO cells expressing P-selectin on their surface, but are not capable of binding to CHO cells which do not express P-selectin on their surface. In order to bind to P-selectin, either in purified form or expressed on the surface of CHO cells, the gene encoding the P-selectin ligand protein must be co-transfected with the gene encoding an FT, since
25 transfection of either gene in the absence of the other either abolishes or substantially reduces the P-selectin binding activity. The binding of the P-selectin ligand protein of the invention to P-selectin can be inhibited by EDTA or by a neutralizing monoclonal antibody specific for P-selectin. The binding of the P-selectin ligand protein of the invention to P-selectin is not inhibited by a non-neutralizing monoclonal antibody specific for P-selectin or by an isotype control. These
30 results characterize the binding specificity of the P-selectin ligand protein of the invention.

For the purposes of the present invention, a protein is defined as having "P-selectin ligand protein activity", i.e., variably referred to herein as a "P-selectin ligand protein", or as a "P-selectin ligand glycoprotein" or simply as a "P-selectin ligand", when it binds in a calcium-dependent
35 manner to P-selectin which is present on the surface of cells as in the CHO-P-selectin binding assay of Example 4, or to P-selectin which is affixed to another surface, for example, as the chimeric P-selectin-IgGyl protein of Example 4 is affixed to Petri dishes.

The glycosylation state of the P-selectin ligand protein of the invention was studied using a chimeric, soluble form of the P-selectin ligand protein, described in detail in Example 5(C), and

5 designated sPSL.T7. The sPSL.T7 protein produced from COS cells co-transfected with 3/4FT is extensively modified by post-translational glycosylation, as described in detail in Example 6(C). Thus, it is believed that both N- and O-linked oligosaccharide chains, at least some of which are sialated, are present on the P-selectin ligand protein of the invention.

The P-selectin ligand protein of the invention may also bind to E-selectin and L-selectin.

10 Conditioned medium from COS cells which have been co-transfected with the DNA encoding sPSL.T7 or P-selectin ligand-Ig fusions and with the DNA encoding 3/4FT, when coated on wells of plastic microtiter plates, causes CHO cells which express E-selectin to bind to the plates; however CHO cells which do not express E-selectin do not bind to such plates. The binding of CHO cells which express E-selectin to microtiter plates coated with conditioned medium from

15 COS cells which have been co-transfected with the DNA encoding sPSL.T7 and with the DNA encoding 3/4FT is abolished in the presence of EDTA or of a neutralizing antibody specific for E-selectin. Conditioned medium from COS cells transfected only with the sPSL.T7 DNA does not cause binding of CHO cells which express E-selectin when coated on wells of microtiter plates. For these reasons, the P-selectin ligand protein of the invention is believed to be useful

20 as an inhibitor of E-selectin-mediated intercellular adhesion in addition to P-selectin-mediated intercellular adhesion.

Antibodies raised against COS-produced soluble P-selectin ligand protein are immunoreactive with the major HL-60 glycoprotein that specifically binds P-selectin as determined by affinity capture using an immobilized Fc chimera of P-selectin. U937 cells bear

25 a similar immunoreactive glycoprotein ligand. Thus, a single glycoprotein species is observed upon EDTA elution of immobilized P-selectin previously incubated with detergent extracts of ³H-glucosamine labeled U937 cells. This major species exhibits an apparent molecular weight by SDS-PAGE of 220 kD under non-reducing conditions and 100 kD under reducing conditions. As with the comparable species isolated from HL-60 cells, this U937 ligand is immunoreactive

30 with a polyclonal antibody raised against COS recombinant P-selectin ligand protein. In addition, affinity capture of E-selectin ligands from U937 cell and cell membrane preparations, using an immobilized Fc chimera of E-selectin, yield a single major species with identical mass and electrophoretic behavior as the major U937 P-selectin ligand. Thus, E- and P-selectin recognize the same major glycoprotein ligand in U937 cells, a glycoprotein ligand immunoreactive with an

35 anti-P-selectin ligand protein antibody and possessing the same apparent mass and electrophoretic behavior as full length, recombinant P-selectin ligand protein.

Fragments of the P-selectin ligand protein which are capable of interacting with P-selectin or which are capable of inhibiting P-selectin-mediated intercellular adhesion are also encompassed

by the present invention. Such fragments comprise amino acids 21 to 54 of SEQ ID NO:2, a region of the P-selectin ligand protein having a low frequency of serine and threonine residues, amino acids 55 to 127 of SEQ ID NO:2, having a high frequency of proline, serine, and threonine in addition to two consensus sequences for asparagine-linked glycosylation (Asn-X-Ser/Thr), another larger fragment, amino acids 128 to 267 of SEQ ID NO:2, having both a high frequency of proline, serine, and threonine and containing fifteen repeats of the following ten amino acid consensus sequence: Ala-(Thr/Met)-Glu-Ala-Gln-Thr-Thr-(Pro/Arg/Gln/Ala/Glu)-(Leu/Pro)-(Ala/Thr) (smaller fragments within this large fragment may also retain the capacity to interact with P-selectin or act as inhibitors of P-selectin-mediated intercellular adhesion); the region containing a consensus sequence for asparagine-linked glycosylation and comprising amino acids 268 to 308 of SEQ ID NO:2; the hydrophobic region of the protein represented by amino acids 309 to 333 of SEQ ID NO:2; and the amphophilic region of the P-selectin ligand protein from amino acids 334 to 402 of SEQ ID NO:2. Additional fragments may comprise amino acid 43 to amino acid 56 of SEQ ID NO:2 or amino acid 42 to amino acid 60 of SEQ ID NO:2, with one or more sulfated or phosphorylated (Domcheck *et al.*, *Biochemistry* 31:9865-9870 (1992)) tyrosines at amino acid 46, amino acid 48, and/or amino acid 51. Fragments of the P-selectin ligand protein may be in linear form or they may be cyclized using known methods, for example, as described in H.U. Saragovi, *et al.*, *Bio/Technology* 10, 773-778 (1992) and in R.S. McDowell, *et al.*, *J. Amer. Chem. Soc.* 114, 9245-9253 (1992), both of which are incorporated herein by reference. For the purposes of the present invention, all references to "P-selectin ligand protein" herein include fragments capable of binding to P-selectin.

Such fragments may be fused to carrier molecules such as immunoglobulins, to increase the valency of P-selectin ligand binding sites. For example, soluble forms of the P-selectin ligand protein such as the fragments from amino acid 42 to amino acid 295 or from amino acid 42 to amino acid 88 of SEQ ID NO:2 may be fused through "linker" sequences to the Fc portion of an immunoglobulin (native sequence or mutated sequences for conferring desirable qualities (such as longer half-life or reduced immunogenicity) to the resulting chimera). For a bivalent form of the P-selectin ligand protein, such a fusion could be to the Fc portion of an IgG molecule as in Example 5(D) and in SEQ ID NO:6. Other immunoglobulin isotypes may also be used to generate such fusions. For example, a P-selectin ligand protein - IgM fusion would generate a decavalent form of the P-selectin ligand protein of the invention.

Fusions of any of the P-selectin ligand proteins of the present invention to amino acid sequences derived from other proteins may also be constructed. Preferred P-selectin ligand proteins for such purpose include the fragments from amino acid 42 to amino acid 295 or from

5 amino acid 42 to amino acid 88 of SEQ ID NO 2. Desirable fusion proteins may incorporate amino acid sequence from proteins having a biological activity different from that of P-selectin ligand, such as, for example, cytokines, growth and differentiation factors (such as bone morphogenetic proteins (e.g., BMPs), hormones, enzymes, receptor components or fragments and other ligands. Also, P-selectin ligand protein can be chemically coupled to other proteins or
10 pharmaceutical agents. In such usage, the P-selectin ligand protein, by virtue of the ability to interact with selectin molecules, alters the pharmacokinetics and/or biodistribution of the fused or coupled agent thereby enhancing its therapeutic efficacy. For example, fusion of a P-selectin ligand protein sequence to a cytokine sequence can direct the cytokine's activity to an area of inflammation. In such instance, the P-selectin ligand protein portion of the fusion protein will
15 bind to selectins expressed at the site of inflammation. This binding will cause the cytokine portion of the fusion protein to become localized and available to bind its cognate receptor or any proximal cell surface. Other ligands could similarly be used in such fusion proteins to attract cells expressing their corresponding receptors to a site of P-selectin expression. Preferred examples of such fusions are described in Example 15.

20 In any fusion protein incorporating a P-selectin ligand protein, the amino acid sequence derived from a protein or proteins other than P-selectin ligand can be linked to either the C-terminus or N-terminus of the P-selectin ligand-derived sequence. The linkage may be direct (i.e., without an intervening linking sequence not derived from either protein) or through a linking sequence.

25 Methods of treating a mammalian subject using such fusion proteins are also contemplated by the present invention. In such instances, the fusion protein is used to treat a condition which is affected by the protein to which the P-selectin ligand protein is fused. For example, a fusion of a P-selectin ligand protein to IL-11 could be used to localize the activity of IL-11 to bone marrow endothelial cells which express selectins on their surface. Once localized,
30 the IL-11 portion of the fusion protein will stimulate megakaryocyte progenitors. Similarly, a fusion of a P-selectin ligand protein to a BMP could be used to stimulate bone or cartilage formation in an area of injury. Injured tissues express P-selectin, which will bind the fusion protein. Once localized, the BMP portion of the fusion protein will stimulate bone or cartilage production in the area of injury.

35 As detailed in the Examples below, the P-selectin ligand protein of the invention was initially obtained using an expression cloning approach (Clark *et al.*, U.S. 4,675,285). A cDNA library was constructed from the human promyelocytic cell line HL-60 (S.J. Collins, *et al.*, *Nature* 270, 347-349 (1977), ATCC No. CCL 240). This library was cotransfected into COS cells with

5 a DNA encoding a 3/4FT, and the cotransfectants were screened for binding to a chimeric molecule consisting of the extracellular portion of P-selectin and the Fc portion of a human IgG γ 1 monoclonal antibody. Cotransfectants which bound to the chimeric P-selectin were enriched for cDNAs encoding the P-selectin ligand protein. This screening process was repeated several times to enrich the plasmid population further for cDNAs encoding the P-selectin ligand protein. In a
10 second cloning stage, the enriched plasmid population was again cotransfected into COS cells with the 3/4FT gene and screened for binding to a fluorescently labeled CHO cell line which expressed P-selectin on the cell surface. A single cDNA clone was obtained from this approach and was designated pMT21.PL85. The pMT21.PL85 plasmid was deposited with the American Type Culture Collection on October 16, 1992 and given the accession number ATCC 69096.

15 One novel DNA of the present invention is set forth in SEQ ID NO:1. The DNA of the present invention may encode a variety of forms of the P-selectin ligand protein. For example, in one embodiment, the DNA of the invention encodes the entire P-selectin ligand protein having the amino acid sequence set forth in SEQ ID NO:2 from amino acid 1 to amino acid 402. In another embodiment, the DNA of the invention encodes a form of the P-selectin ligand protein
20 which lacks the signal sequence and which is characterized by the amino acid sequence set forth in SEQ ID NO:2 from amino acid 21 to amino acid 402. In yet another embodiment, the DNA of the invention encodes the mature P-selectin ligand protein characterized by the amino acid sequence set forth in SEQ ID NO:2 from amino acid 42 to amino acid 402. Another embodiment of the DNA of the invention encodes a soluble form of the P-selectin ligand protein characterized
25 by the amino acid sequence set forth in SEQ ID NO:2 from amino acid 1 to amino acid 310. The DNA of the invention is also embodied in a DNA encoding a soluble form of the mature P-selectin ligand protein, said protein being characterized by the amino acid sequence set forth in SEQ ID NO:2 from amino acid 42 to amino acid 310. The DNA of the invention is further embodied in a DNA sequence encoding a soluble form of the P-selectin ligand protein which
30 lacks the signal sequence, said protein being characterized by the amino acid sequence set forth in SEQ ID NO:2 from amino acid 21 to amino acid 310. The DNA of the present invention is free from association with other human DNAs and is thus characterized as an isolated DNA. As detailed above, DNAs which encode P-selectin ligand fragments which interact with P-selectin are also included in the present invention.

35 The expression of P-selectin ligand protein mRNA transcripts has been observed in a variety of human cell lines (HL-60, THP-1, U937) and in human monocytes and polymorphonuclear leukocytes by Northern analysis using a P-selectin ligand protein cDNA probe. In all of these cell lines, a major transcript of 2.5 kb was observed. A minor species of

5 approximately 4 kb was observed in the HL60 and U937 cell lines and in polymorphonuclear leukocytes. In contrast, no P-selectin ligand mRNA expression was detected in the human hepatoblastoma cell line HepG2.

The P-selectin ligand protein of the invention is encoded by a single copy gene and is not part of a multi-gene family, as determined by Southern blot analysis. The genomic form of the P-selectin ligand protein of the invention contains a large intron of approximately 9 kb located at nucleotide 54 in the 5' untranslated region. In polymorphonuclear leukocytes and monocytes, the P-selectin ligand protein of the invention is encoded by the DNA sequence set forth in SEQ ID NO:3. In this embodiment, the P-selectin ligand protein contains sixteen repeat regions. The isolated DNA of the invention is correspondingly also embodied in the DNA sequence set forth in SEQ ID NO:3 and is contained on plasmid pPL85R16 which was deposited with the American Type Culture Collection on October 22, 1993 and given the Accession Number ATCC 75577.

The invention also encompasses allelic variations of the isolated DNA as set forth in SEQ ID NO:1 or of the isolated DNA as set forth in SEQ ID NO:3, that is, naturally-occurring alternative forms of the isolated DNA of SEQ ID NO:1 or SEQ ID NO:3 which also encode proteins having P-selectin ligand activity. Also included in the invention are isolated DNAs which hybridize to the DNA set forth in SEQ ID NO:1 or to the DNA set forth in SEQ ID NO:3 under stringent (e.g. 4xSSC at 65°C or 50% formamide and 4xSSC at 42°C), or relaxed (4xSSC at 50°C or 30-40% formamide at 42°C) conditions, and which have P-selectin ligand protein activity. Isolated DNA sequences which encode the P-selectin ligand protein but which differ from the DNA set forth in SEQ ID NO:1 or from the DNA set forth in SEQ ID NO:3 by virtue of the degeneracy of the genetic code and which have P-selectin ligand protein activity are also encompassed by the present invention. Variations in the DNA as set forth in SEQ ID NO:1 or in the DNA as set forth in SEQ ID NO:3 which are caused by point mutations or by induced modifications which enhance the P-selectin ligand activity, half-life or production level are also included in the invention. For the purposes of the present invention all references herein to the "DNA of SEQ ID NO:1" include, in addition to DNAs comprising the specific DNA sequence set forth in SEQ ID NO:1, DNAs encoding the mature P-selectin ligand protein of SEQ ID NO:2, DNAs encoding fragments of the P-selectin ligand protein of SEQ ID NO:2 which are capable of binding to P-selectin; DNAs encoding soluble forms of the P-selectin ligand protein of SEQ ID NO:2; allelic variations of the DNA sequence of SEQ ID NO:1, DNAs which hybridize to the DNA sequence of SEQ ID NO:1 and which encode proteins having P-selectin ligand protein activity, DNAs which differ from the DNA of SEQ ID NO:1 by virtue of degeneracy of the genetic code, and the variations of the DNA sequence of SEQ ID NO:1 set forth above. Similarly,

all references to the "DNA of SEQ ID NO:3" include in addition to the specific sequence set forth in SEQ ID NO:3, DNAs encoding the mature P-selectin ligand protein of SEQ ID NO:4, DNAs encoding fragments of the P-selectin ligand protein of SEQ ID NO:4 which are capable of binding to P-selectin, DNAs encoding soluble forms of the P-selectin ligand protein of SEQ ID NO:4, allelic variations of the DNA of SEQ ID NO:3, DNAs which hybridize to the DNA sequence of SEQ ID NO:3 and which encode proteins having P-selectin ligand protein activity, DNAs which differ from the DNA of SEQ ID NO:3 by virtue of degeneracy of the genetic code, and the variations of the DNA of SEQ ID NO:3 set forth above.

A DNA encoding a soluble form of the P-selectin ligand protein may be prepared by expression of a modified DNA in which the regions encoding the transmembrane and cytoplasmic domains of the P-selectin ligand protein are deleted and/or a stop codon is introduced 3' to the codon for the amino acid at the carboxy terminus of the extracellular domain. For example, hydrophobicity analysis predicts that the P-selectin ligand protein set forth in SEQ ID NO:2 has a transmembrane domain comprised of amino acids 311 to 332 of SEQ ID NO:2 and a cytoplasmic domain comprised of amino acids 333 to 402 of SEQ ID NO:2. A modified DNA as described above may be made by standard molecular biology techniques, including site-directed mutagenesis methods which are known in the art or by the polymerase chain reaction using appropriate oligonucleotide primers. Methods for producing several DNAs encoding various soluble P-selectin ligand proteins are set forth in Example 5.

A DNA encoding other fragments and altered forms of P-selectin ligand protein may be prepared by expression of modified DNAs in which portions of the full-length sequence have been deleted or altered. Substantial deletions of the P-selectin ligand protein sequence can be made while retaining P-selectin ligand protein activity. For example, P-selectin ligand proteins comprising the sequence from amino acid 42 to amino acid 189 of SEQ ID NO:2, the sequence from amino acid 42 to amino acid 118 of SEQ ID NO:2, or the sequence from amino acid 42 to amino acid 89 of SEQ ID NO:2 each retain the P-selectin protein binding activity and the ability to bind to E-selectin. P-selectin ligand proteins in which one or more N-linked glycosylation sites (such as those at amino acids 65, 111 and 292 of SEQ ID NO:2) have been changed to other amino acids or deleted also retain P-selectin protein binding activity and the ability to bind E-selectin. P-selectin ligand proteins comprising from amino acid 42 to amino acid 60 of SEQ ID NO:2 (which includes a highly anionic region of the protein from amino acid 45 to amino acid 58 of SEQ ID NO:2) also retain P-selectin ligand protein activity, however, P-selectin ligand proteins limited to such sequence do not bind to E-selectin. Preferably, a P-selectin ligand protein retains at least one (more preferably at least two and most preferably all three) of the tyrosine

5 residues found at amino acids 46, 48 and 51 of SEQ ID NO. 2, sulfation of which may contribute to P-selectin ligand protein activity. Construction of DNAs encoding these and other active fragments or altered forms of P-selectin ligand protein may be accomplished in accordance with methods known to those skilled in the art.

The isolated DNA of the invention may be operably linked to an expression control
10 sequence such as the pMT2 or pED expression vectors disclosed in Kaufman *et al.*, Nucleic Acids Res. 19, 4485-4490 (1991), in order to produce the P-selectin ligand recombinantly. Many suitable expression control sequences are known in the art. General methods of expressing recombinant proteins are also known and are exemplified in R. Kaufman, Methods in Enzymology 185, 537-566 (1990). As defined herein "operably linked" means enzymatically or
15 chemically ligated to form a covalent bond between the isolated DNA of the invention and the expression control sequence, in such a way that the P-selectin ligand protein is expressed by a host cell which has been transformed (transfected) with the ligated DNA/expression control sequence.

Several endoproteolytic enzymes are known which cleave precursor peptides at the carboxyl side of paired amino acid sequences (e.g., -Lys-Arg- and -Arg-Arg-) to yield mature
20 proteins. Such enzymes are generally known as paired basic amino acid converting enzymes or PACE, and their use in recombinant production of mature peptides is extensively disclosed in WO 92/09698 and U.S. Application Serial No. 07/885,972, both of which are incorporated herein by reference. The PACE family of enzymes are known to increase the efficiency of proteolytic processing of precursor polypeptides in recombinant host cells. As mentioned above, the P-
25 selectin ligand protein of the invention contains such a PACE cleavage site.

The soluble mature P-selectin ligand protein of the present invention may be made by a host cell which contains a DNA sequence encoding any soluble P-selectin ligand protein as described herein and a DNA sequence encoding PACE as described in WO 92/09698 and U.S. Application Serial No. 07/885,972, incorporated herein by reference, or using the DNA sequence
30 of SEQ ID NO:5. Such a host cell may contain the DNAs as the result of co-transformation or sequential transformation of separate expression vectors containing the soluble P-selectin ligand protein DNA and the PACE DNA, respectively. A third DNA which encodes a 3/4FT may also be co-transformed with the DNAs encoding the P-selectin ligand protein and PACE. Alternatively, the host cell may contain the DNAs as the result of transformation of a single
35 expression vector containing both soluble P-selectin ligand protein DNA and PACE DNA. Construction of such expression vectors is within the level of ordinary skill in molecular biology. Methods for co-transformation and transformation are also known.

5 Many DNA sequences encoding PACE are known. For example, a DNA encoding one form of PACE, known as furin, is disclosed in A.M.W. van den Ouweland *et al.*, Nucl. Acids Res. 18, 664 (1990), incorporated herein by reference. A cDNA encoding a soluble form of PACE, known as PACESOL, is set forth in SEQ ID NO 5. DNAs encoding other forms of PACE also exist, and any such PACE-encoding DNA may be used to produce the soluble mature P-selectin
10 ligand protein of the invention, so long as the PACE is capable of cleaving the P-selectin ligand protein at amino acids 38-41. Preferably, a DNA encoding a soluble form of PACE is used to produce the soluble mature P-selectin ligand protein of the present invention.

The DNAs encoding a soluble form of the P-selectin ligand protein and PACE, separately or together, may be operably linked to an expression control sequence such as those contained in
15 the pMT2 or pED expression vectors discussed above, in order to produce the PACE-cleaved soluble P-selectin ligand recombinantly. Additional suitable expression control sequences are known in the art. Examples 3(C) and 3(D) below set forth methods for producing the soluble mature P-selectin ligand protein of the invention.

A number of types of cells may act as suitable host cells for expression of the P-selectin
20 ligand protein. Suitable host cells are capable of attaching carbohydrate side chains characteristic of functional P-selectin ligand protein. Such capability may arise by virtue of the presence of a suitable glycosylating enzyme within the host cell, whether naturally occurring, induced by chemical mutagenesis, or through transfection of the host cell with a suitable expression plasmid containing a DNA sequence encoding the glycosylating enzyme. Host cells include, for example,
25 monkey COS cells, Chinese Hamster Ovary (CHO) cells, human kidney 293 cells, human epidermal A431 cells, human Colo205 cells, 3T3 cells, CV-1 cells, other transformed primate cell lines, normal diploid cells, cell strains derived from in vitro culture of primary tissue, primary explants, HeLa cells, mouse L cells, BHK, HL-60, U937, or HaK cells.

The P-selectin ligand protein may also be produced by operably linking the isolated DNA
30 of the invention and one or more DNAs encoding suitable glycosylating enzymes to suitable control sequences in one or more insect expression vectors, and employing an insect expression system. Materials and methods for baculovirus/insect cell expression systems are commercially available in kit form from, e.g., Invitrogen, San Diego, California, U.S.A. (the MaxBac® kit), and such methods are well known in the art, as described in Summers and Smith, Texas Agricultural
35 Experiment Station Bulletin No. 1555 (1987), incorporated herein by reference. Soluble forms of the P-selectin ligand protein may also be produced in insect cells using appropriate isolated DNAs as described above. A DNA encoding a form of PACE may further be co-expressed in an insect host cell to produce a PACE-cleaved form of the P-selectin ligand protein.

5 Alternatively, it may be possible to produce the P-selectin ligand protein in lower eukaryotes such as yeast or in prokaryotes such as bacteria. Potentially suitable yeast strains include *Saccharomyces cerevisiae*, *Schizosaccharomyces pombe*, *Kluyveromyces* strains, *Candida*, or any yeast strain capable of expressing heterologous proteins. Potentially suitable bacterial strains include *Escherichia coli*, *Bacillus subtilis*, *Salmonella typhimurium*, or any bacterial strain
10 capable of expressing heterologous proteins. If the P-selectin ligand protein is made in yeast or bacteria, it is necessary to attach the appropriate carbohydrates to the appropriate sites on the protein moiety covalently, in order to obtain the glycosylated P-selectin ligand protein. Such covalent attachments may be accomplished using known chemical or enzymatic methods.

 The P-selectin ligand protein of the invention may also be expressed as a product of
15 transgenic animals, e.g., as a component of the milk of transgenic cows, goats, pigs, or sheep which are characterized by somatic or germ cells containing a DNA sequence encoding the P-selectin ligand protein.

 The P-selectin binding activity of a P-selectin protein may be enhanced by co-transformation of a host cell with a GlcNAc transferase, preferably UDP-GlcNAc:Gal β 1-3GalNAc-R(GlcNAc to GalNAc) β 1-6 GlcNAc transferase (EC 2.4.1.102), also known as "core2
20 transferase."

 O-linked glycans present on P-selectin ligand protein have been shown to be important for binding to P-selectin (D. Sako *et al.*, *Cell* **75**, 1179-1186 (1993)). It has been reported that sialyl Le^x on O-linked glycans of myeloid cells are presented on complex, branched structures
25 (Maemura, K. and Fukuda, M., *J. Biol. Chem.* **267**, 24379-24386 (1992)). The enzyme responsible for generating such oligosaccharide structures is "core2". The core2 enzyme activity is found at very low levels in COS cells and at trace levels in CHO cells. Host cells co-transformed with DNAs encoding a P-selectin ligand protein, an (α 1.3/ α 1.4) fucosyltransferase and core2 can produce P-selectin ligand protein exhibiting 20-30 fold enhanced binding to P-
30 selectin.

 In certain preferred embodiments, P-selectin ligand protein is produced by co-transfecting a host cell with DNAs encoding soluble P-selectin ligand protein, 3/4FT, core2 and PACE

 The P-selectin ligand protein of the invention may be prepared by culturing transformed host cells under culture conditions necessary to express a P-selectin binding glycoprotein. The
35 resulting expressed glycoprotein may then be purified from culture medium or cell extracts. Soluble forms of the P-selectin ligand protein of the invention can be purified by affinity chromatography over Lentil lectin-Sepharose® and subsequent elution with 0.5M α -methylmannoside. The eluted soluble P-selectin ligand protein can then be further purified and

concentrated by a 0-70% ammonium sulfate precipitation step. The protein is then recovered, resuspended, and further purified by size exclusion chromatography over a TSK G4000SW_{XL}. Alternatively, full length forms of the P-selectin ligand protein of the invention can be purified by preparing a total membrane fraction from the expressing cell and extracting the membranes with a non-ionic detergent such as Triton X-100. The detergent extract can then be passed over an affinity column comprised of immobilized P-selectin, and the P-selectin ligand protein can be eluted from the column with 10mM EDTA in a buffer containing 0.1% detergent. The material eluted from the affinity column can then be dialyzed to remove EDTA and further purified over a Lentil lectin-Sepharose® affinity column, again eluting with 0.5M α -methyl-mannoside.

Alternatively, the P-selectin ligand protein of the invention is concentrated using a commercially available protein concentration filter, for example, an Amicon or Millipore Pellicon ultrafiltration unit. Following the concentration step, the concentrate can be applied to a purification matrix such as a gel filtration medium. Alternatively, an anion exchange resin can be employed, for example, a matrix or substrate having pendant diethylaminoethyl (DEAE) groups. The matrices can be acrylamide, agarose, dextran, cellulose or other types commonly employed in protein purification. Alternatively, a cation exchange step can be employed. Suitable cation exchangers include various insoluble matrices comprising sulfopropyl or carboxymethyl groups. Sulfopropyl groups are preferred (e.g., S-Sepharose® columns). The purification of the P-selectin ligand protein from culture supernatant may also include one or more column steps over such affinity resins as concanavalin A-agarose, heparin-toyopearl® or Cibacrom blue 3GA Sepharose®; or by hydrophobic interaction chromatography using such resins as phenyl ether, butyl ether, or propyl ether; or by immunoaffinity chromatography.

Finally, one or more reverse-phase high performance liquid chromatography (RP-HPLC) steps employing hydrophobic RP-HPLC media, e.g., silica gel having pendant methyl or other aliphatic groups, can be employed to further purify the P-selectin ligand protein. Some or all of the foregoing purification steps, in various combinations, can also be employed to provide a substantially homogeneous isolated recombinant protein. The P-selectin ligand protein thus purified is substantially free of other mammalian proteins and is defined in accordance with the present invention as "isolated P-selectin ligand protein".

Isolated P-selectin ligand protein may be useful in treating conditions characterized by P-, E- or L-selectin mediated intercellular adhesion. Such conditions include, without limitation, myocardial infarction, bacterial or viral infection, metastatic conditions, inflammatory disorders such as arthritis, gout, uveitis, acute respiratory distress syndrome, asthma, emphysema, delayed type hypersensitivity reaction, systemic lupus erythematosus, thermal injury such as burns or

5 trostbite, autoimmune thyroiditis, experimental allergic encephalomyelitis, multiple sclerosis, multiple organ injury syndrome secondary to trauma, diabetes, Reynaud's syndrome, neutrophilic dermatosis (Sweet's syndrome), inflammatory bowel disease, Grave's disease, glomerulonephritis, gingivitis, periodontitis, hemolytic uremic syndrome, ulcerative colitis, Crohn's disease, necrotizing enterocolitis, granulocyte transfusion associated syndrome, cytokine-induced toxicity, and the like. Isolated P-selectin ligand protein may also be useful in organ transplantation, both to prepare organs for transplantation and to quell organ transplant rejection. Accordingly, P-selectin ligand protein may be administered to a living or non-living organ donor, prior to organ removal. In addition, P-selectin ligand protein may be administered "*ex-vivo*" to the donor organ concomitantly with organ preservation solution, prior to, and/or subsequent to surgical anastomosis with the recipient. Isolated P-selectin ligand protein may be used to treat hemodialysis and leukophoresis patients. Additionally, isolated P-selectin ligand protein may be used as an antimetastatic agent. Isolated P-selectin ligand protein may be used itself as an inhibitor of P-, E- or L-selectin-mediated intercellular adhesion or to design inhibitors of P-, E- or L-selectin-mediated intercellular adhesion. The present invention encompasses both pharmaceutical compositions containing isolated P-selectin ligand protein and therapeutic methods of treatment or use which employ isolated P-selectin ligand protein.

Isolated P-selectin ligand protein, purified from cells or recombinantly produced, may be used as a pharmaceutical composition when combined with a pharmaceutically acceptable carrier. Such a composition may contain, in addition to P-selectin ligand protein and carrier, diluents, fillers, salts, buffers, stabilizers, solubilizers, and other materials well known in the art. The term "pharmaceutically acceptable" means a non-toxic material that does not interfere with the effectiveness of the biological activity of the active ingredient(s). The characteristics of the carrier will depend on the route of administration. The pharmaceutical composition of the invention may also contain cytokines, lymphokines, or other hematopoietic factors such as M-CSF, GM-CSF, IL-1, IL-2, IL-3, IL-4, IL-5, IL-6, IL-7, IL-8, IL-9, IL-10, IL-11, IL-12, G-CSF, Meg-CSF, stem cell factor, and erythropoietin. The pharmaceutical composition may contain thrombolytic or anti-thrombotic factors such as plasminogen activator and Factor VIII. The pharmaceutical composition may further contain other anti-inflammatory agents. Such additional factors and/or agents may be included in the pharmaceutical composition to produce a synergistic effect with isolated P-selectin ligand protein, or to minimize side effects caused by the isolated P-selectin ligand protein. Conversely, isolated P-selectin ligand protein may be included in formulations of the particular cytokine, lymphokine, other hematopoietic factor, thrombolytic or anti-thrombotic factor, or anti-inflammatory agent to minimize side effects of the cytokine,

5 lymphokine, other hematopoietic factor, thrombolytic or anti-thrombotic factor, or anti-inflammatory agent.

The pharmaceutical composition of the invention may be in the form of a liposome in which isolated P-selectin ligand protein is combined, in addition to other pharmaceutically acceptable carriers, with amphipathic agents such as lipids which exist in aggregated form as micelles, insoluble monolayers, liquid crystals, or lamellar layers which in aqueous solution. Suitable lipids for liposomal formulation include, without limitation, monoglycerides, diglycerides, sulfatides, lysolecithin, phospholipids, saponin, bile acids, and the like. Preparation of such liposomal formulations is within the level of skill in the art, as disclosed, for example, in U.S. Patent No. 4,235,871; U.S. Patent No. 4,501,728; U.S. Patent No. 4,837,028; and U.S. Patent No. 4,737,323, all of which are incorporated herein by reference.

As used herein, the term "therapeutically effective amount" means the total amount of each active component of the pharmaceutical composition or method that is sufficient to show a meaningful patient benefit, i.e., healing of chronic conditions characterized by P-selectin- or E-selectin-mediated cellular adhesion or increase in rate of healing of such conditions. When applied to an individual active ingredient, administered alone, the term refers to that ingredient alone. When applied to a combination, the term refers to combined amounts of the active ingredients that result in the therapeutic effect, whether administered in combination, serially or simultaneously.

In practicing the method of treatment or use of the present invention, a therapeutically effective amount of isolated P-selectin ligand protein is administered to a mammal having a P-selectin-mediated disease state. Isolated P-selectin ligand protein may be administered in accordance with the method of the invention either alone or in combination with other therapies such as treatments employing receptor antagonists, ligand antagonists, cytokines, lymphokines or other hematopoietic factors. When co-administered with one or more cytokines, lymphokines or other hematopoietic factors, isolated P-selectin ligand protein may be administered either simultaneously with the cytokine(s), lymphokine(s), other hematopoietic factor(s), thrombolytic or anti-thrombotic factors, or sequentially. If administered sequentially, the attending physician will decide on the appropriate sequence of administering isolated P-selectin ligand protein in combination with cytokine(s), lymphokine(s), other hematopoietic factor(s), thrombolytic or anti-thrombotic factors.

Administration of isolated P-selectin ligand protein used in the pharmaceutical composition or to practice the method of the present invention can be carried out in a variety of

5 conventional ways, such as oral ingestion, inhalation, or cutaneous, subcutaneous, or intravenous injection. Intravenous administration to the patient is preferred.

When a therapeutically effective amount of isolated P-selectin ligand protein is administered orally, isolated P-selectin ligand protein will be in the form of a tablet, capsule, powder, solution or elixir. When administered in tablet form, the pharmaceutical composition of
10 the invention may additionally contain a solid carrier such as a gelatin or an adjuvant. The tablet, capsule, and powder contain from about 5 to 95% isolated P-selectin ligand protein, and preferably from about 25 to 90% isolated P-selectin ligand protein. When administered in liquid form, a liquid carrier such as water, petroleum, oils of animal or plant origin such as peanut oil, mineral oil, soybean oil, or sesame oil, or synthetic oils may be added. The liquid form of the
15 pharmaceutical composition may further contain physiological saline solution, dextrose or other saccharide solution, or glycols such as ethylene glycol, propylene glycol or polyethylene glycol. When administered in liquid form, the pharmaceutical composition contains from about 0.5 to 90% by weight of isolated P-selectin ligand protein and preferably from about 1 to 50% isolated P-selectin ligand protein.

20 When a therapeutically effective amount of isolated P-selectin ligand protein is administered by intravenous, cutaneous or subcutaneous injection, isolated P-selectin ligand protein will be in the form of a pyrogen-free, parenterally acceptable aqueous solution. The preparation of such parenterally acceptable protein solutions, having due regard to pH, isotonicity, stability, and the like, is within the skill in the art. A preferred pharmaceutical composition for
25 intravenous, cutaneous, or subcutaneous injection should contain, in addition to isolated P-selectin ligand protein an isotonic vehicle such as Sodium Chloride Injection, Ringer's Injection, Dextrose Injection, Dextrose and Sodium Chloride Injection, Lactated Ringer's Injection, or other vehicle as known in the art. The pharmaceutical composition of the present invention may also contain stabilizers, preservatives, buffers, antioxidants, or other additive known to those of skill in the art.

30 The amount of isolated P-selectin ligand protein in the pharmaceutical composition of the present invention will depend upon the nature and severity of the condition being treated, and on the nature of prior treatments which the patient has undergone. Ultimately, the attending physician will decide the amount of isolated P-selectin ligand protein with which to treat each individual patient. Initially, the attending physician will administer low doses of isolated P-
35 selectin ligand protein and observe the patient's response. Larger doses of isolated P-selectin ligand protein may be administered until the optimal therapeutic effect is obtained for the patient, and at that point the dosage is not increased further. It is contemplated that the various

5 pharmaceutical compositions used to practice the method of the present invention should contain about 0.1 μ g to about 100 mg of isolated P-selectin ligand protein per kg body weight.

The duration of intravenous therapy using the pharmaceutical composition of the present invention will vary, depending on the severity of the disease being treated and the condition and potential idiosyncratic response of each individual patient. It is contemplated that the duration
10 of each application of the isolated P-selectin ligand protein will be in the range of 12 to 24 hours of continuous intravenous administration. Ultimately the attending physician will decide on the appropriate duration of intravenous therapy using the pharmaceutical composition of the present invention.

Isolated P-selectin ligand protein of the invention may also be used to immunize animals
15 to obtain polyclonal and monoclonal antibodies which specifically react with the P-selectin ligand protein and which may inhibit P-selectin-mediated cellular adhesion. Such antibodies may be obtained using the entire P-selectin ligand protein as an immunogen, or by using fragments of P-selectin ligand protein such as the soluble mature P-selectin ligand protein. Smaller fragments of the P-selectin ligand protein may also be used to immunize animals, such as the fragments set
20 forth below: amino acid 42 to amino acid 56 of SEQ ID NO:2 and amino acid 127 to amino acid 138 of SEQ ID NO:2. An additional peptide immunogen comprises amino acid 238 to amino acid 248 of SEQ ID NO:2, with an alanine residue added to the amino terminus of the peptide. Another peptide immunogen comprises amino acid 43 to amino acid 56 of SEQ ID NO:2 having a sulfated tyrosine in any or all of positions 46, 48 or 51. The peptide immunogens additionally
25 may contain a cysteine residue at the carboxyl terminus, and are conjugated to a hapten such as keyhole limpet hemocyanin (KLH). Additional peptide immunogens may be generated by replacing tyrosine residues with sulfated tyrosine residues. Methods for synthesizing such peptides are known in the art, for example, as in R.P. Merrifield, *J. Amer. Chem. Soc.* 85, 2149-2154 (1963); J.L. Krstenansky, *et al.*, *FEBS Lett.* 211, 10 (1987).

30 Monoclonal antibodies binding to P-selectin ligand glycoprotein or to complex carbohydrate moieties characteristic of the P-selectin ligand glycoprotein may be useful diagnostic agents for the immunodetection of inflammatory diseases and some forms of cancer. Some cancerous cells, such as small cell lung carcinomas, may express detectable levels of the P-selectin ligand protein. This abnormal expression of the P-selectin ligand protein by cancer cells may play
35 a role in the metastasis of these cells.

Neutralizing monoclonal antibodies binding to P-selectin ligand glycoprotein or to complex carbohydrates characteristic of P-selectin ligand glycoprotein may also be useful therapeutics for both inflammatory diseases and also in the treatment of some forms of cancer

5 where abnormal expression of P-selectin ligand protein is involved. These neutralizing monoclonal antibodies are capable of blocking the selectin mediated intercellular adherence function of the P-selectin ligand protein. By blocking the binding of P-selectin ligand protein, the adherence of leukocytes to sites of inappropriate inflammation is either abolished or markedly reduced. In the case of cancerous cells or leukemic cells, neutralizing monoclonal antibodies
10 against P-selectin ligand protein may be useful in detecting and preventing the metastatic spread of the cancerous cells which may be mediated by the P-selectin ligand protein. In addition, the monoclonal antibodies bound to these cells may target the cancerous cells for antibody-dependent cell mediated cytotoxicity (ADCC), thus helping to eliminate the cancerous cells. Human antibodies which react with the P-selectin ligand protein may be produced in transgenic animals
15 which contain human immunoglobulin encoding genes in their germ lines. Example 7 below sets forth production of a rabbit polyclonal antibody specific P-selectin ligand protein fragments.

P-selectin ligand protein of the invention may also be used to screen for agents which are capable of binding to P-selectin ligand protein and thus may act as inhibitors of P-selectin- or E-selectin-mediated intercellular adhesion. Binding assays using a desired binding protein,
20 immobilized or not, are well known in the art and may be used for this purpose using the P-selectin ligand protein of the invention. Appropriate screening assays may be cell-based, as in Examples 3 and 9 below. Alternatively, purified protein based screening assays may be used to identify such agents. For example, P-selectin ligand protein may be immobilized in purified form on a carrier and binding to purified P-selectin may be measured in the presence and in the absence
25 of potential inhibiting agents. A suitable binding assay may alternatively employ purified P-selectin immobilized on a carrier, with a soluble form of P-selectin ligand protein of the invention.

Any P-selectin ligand protein may be used in the screening assays described above. For example, the full-length P-selectin ligand protein set forth in SEQ ID NO:2 from amino acid 1 to amino acid 402 may be used to screen for inhibitors; or the mature P-selectin ligand protein set
30 forth in SEQ ID NO:2 from amino acid 42 to amino acid 402 may be used to screen for inhibitors, or the soluble mature P-selectin ligand protein set forth in SEQ ID NO:2 from amino acid 42 to amino acid 310 may be used to screen for inhibitors. Alternatively, the P-selectin ligand protein of SEQ ID NO:4 from amino acid 1 to amino acid 412, or a mature form of the P-selectin ligand protein as set forth in SEQ ID NO:4 from amino acid 42 to amino acid 412, or a soluble mature
35 form of the P-selectin ligand protein set forth in SEQ ID NO:4 from amino acid 42 to amino acid 320 may be used to screen for inhibitors of intercellular adhesion in accordance with the present invention.

5 In such a screening assay, a first binding mixture is formed by combining P-selectin or E-selectin and P-selectin ligand protein, and the amount of binding in the first binding mixture (B_0) is measured. A second binding mixture is also formed by combining P-, E- or L-selectin, P-selectin ligand protein, and the compound or agent to be screened, and the amount of binding in the second binding mixture (B) is measured. The amounts of binding in the first and second
10 binding mixtures are compared, for example, by performing a B/B_0 calculation. A compound or agent is considered to be capable of inhibiting P-, E- or L-selectin mediated intercellular adhesion if a decrease in binding in the second binding mixture as compared to the first binding mixture is observed. The formulation and optimization of binding mixtures is within the level of skill in the art, such binding mixtures may also contain buffers and salts necessary to enhance or to
15 optimize binding, and additional control assays may be included in the screening assay of the invention.

Compounds found to reduce by at least about 10%, preferably greater than about 50% or more of the binding activity of P-selectin ligand protein to P-, E- or L-selectin may thus be identified and then secondarily screened in other selectin binding assays, including assays binding
20 to L-selectin and *in vivo* assays. By these means compounds having inhibitory activity for selectin-mediated intercellular adhesion which may be suitable as anti-inflammatory agents may be identified.

EXAMPLE 1

25 CLONING OF THE P-SELECTIN LIGAND PROTEIN GENE

A. Construction of the HL60 cDNA library

An HL60 cDNA library was constructed for expression cloning the P-selectin ligand. PolyA⁺ RNA was isolated from total RNA from the human promyelocytic cell line HL60 (S.J. Collins, *et al.*, *supra*) using a Fast Track mRNA Isolation Kit (Invitrogen, San Diego, CA).
30 Double stranded cDNA was synthesized from the polyA⁺ RNA fraction and blunt-end ligated with EcoRI adaptors (5'-AATTCGTCGACTCTAGAG-3', SEQ ID NO 7; 5'-CTCTAGAGTCGACGG-3', SEQ ID NO 8). The cDNA was ligated into the expression vector pMT21 (R. Kaufman *et al.*, J. Mol. Cell. Biol. 9, 946-958 (1989) that had been incubated sequentially with EcoRI
35 endonuclease and calf intestinal alkaline phosphatase and gel purified. The ligation product was electroporated in 2 μ l aliquots into competent *E. coli* DH5 α cells and grown in 1 ml of SOB medium (J. Sambrook *et al.*, Molecular Cloning: A Laboratory Manual, New York, Cold Spring Harbor Laboratory Press, p1 90 (1989)) which has been supplemented with 10 mM MgCl₂, 10

5 mM MgSO₄, and 2% glycerol for one hour at 37°C. In order to divide the library into smaller subsets, an aliquot from each ml of bacterial suspension was plated onto agar plates in the presence of ampicillin, and the number of colonies per ml was calculated. Assuming that each colony represented one cDNA clone, 600,000 clones were generated and divided into subsets of approximately 16,000 clones per pool. Each of the 38 pools were grown overnight in L-broth in the presence of ampicillin and the plasmids were purified over a CsCl gradient.

B. Screening for the P-selectin ligand protein gene

In the first stage, the LEC-γ1 binding assay of Example 4(A) was utilized to pan the HL60 cDNA library and thereby to enrich for the plasmid of interest. Six μg of each HL60 cDNA library pool was co-transfected with 2 μg of α 3/4FT gene (Example 2) into COS cells. Approximately 45 hours post-transfection, the COS cells were lifted from the plates by incubating the cells in 1 mM EGTA for 15 min. at 37°C, followed by scraping with cell lifters. The cells were washed twice in Hanks buffered saline solution containing 1 mM calcium (HBSS). The cells were resuspended in 4 ml of HBSS. The resuspended transfected COS cells were screened using the LEC-γ1 binding assay described in Example 4(A).

The plasmids from adherent COS cells were recovered from a Hirts extract [B. Hirts, J. Mol. Biol., 26, 365-369 (1967)] and then electroporated into *E. coli* DH5α cells for amplification. The enriched population of plasmids was purified over a CsCl gradient and re-transfected along with the 3/4FT gene (Example 2) into COS cells. The transfection, screening, and plasmid amplification process was repeated for a total of three times before a pool that bound to the LEC-γ1-coated plates was visually detected. The positive plasmid pool was subsequently broken down into subsets. This involved electroporating the Hirts extract from the positive pool into *E. coli* DH5α cells and quantitating colonies per ml as described above. Various pool sizes were produced by plating out a predetermined number of colonies on agar plates in the presence of ampicillin. Duplicate plates were prepared by performing nitrocellulose lifts and storing the filters on new agar plates. The duplicate plates served as reference plates for selecting individual or groups of colonies from any pool identified as being positive.

In the second stage of cloning, COS cells were co-transfected with the sublibrary pools and the 3/4FT gene by the same procedure used in the initial steps of screening. Forty-eight hours post-transfection, the transfected cells were screened using the fluorescent CHO.P-selectin assay of Example 4(B). Positive pools were further subdivided, as described above, until finally individual colonies were screened and positive clones identified. Using this method, a single positive clone, pMT21 PL85, was found to encode the P-selectin ligand protein. The DNA

5 sequence of the P-selectin ligand contained in pMT21 PL85 is set forth in SEQ ID NO:1, and the binding characteristics of the P-selectin ligand protein encoded by pMT21 PL85 are set forth in Example 4(C) below.

10 EXAMPLE 2

CLONING THE α 1,3/1,4 FUCOSYLTRANSFERASE GENE

The α 1,3/1,4 fucosyltransferase gene (3/4FT) was cloned from total human genomic DNA (Clontech Laboratories) by means of PCR. The sense oligonucleotide primer contained an XbaI site and the 5' terminus of the gene (5'-
15 TAGCATACGCTCTAGAGCATGGATCCCCTGGGTGCAGCCAAGC-3', SEQ ID NO:9), and the antisense oligonucleotide primer contained an EcoRI site and the 3' terminus of the gene (5'-CCGGAATTCTCAGGTGAACCAAGCCGC-3', SEQ ID NO:10). The PCR product was sequentially digested with XbaI and EcoRI and purified by standard gel purification methods. This gene was then ligated with vector pMT3Sv2ADA (R. Kaufman, Methods in Enzymology, *supra*) that had
20 also been sequentially digested with XbaI and EcoRI and purified by standard gel purification methods. Competent HB101 cells (Biorad) were transformed with this ligation product and then plated on agar plates in the presence of ampicillin. Nitrocellulose filter lifts of ampicillin-resistant transformants were probed with a radiolabelled oligonucleotide (5'-
25 AAGTATCTGTCCAGGGCTTCCAGGT-3', SEQ ID NO:11) complementary to the nucleotide region 506-530 in the middle of the gene (J. Sambrook *et al.*, *supra*).

Plasmid DNA minipreps were prepared from twelve positive clones. The purified DNA was then digested with EcoRI and XbaI to identify the correct clone with the proper size insert. This clone (pEA.3/4FT) was then grown up large scale and the DNA isolated by CsCl density gradient banding (J. Sambrook *et al.*, *supra*). DNA sequencing confirmed the identity of the
30 3/4FT gene. The functionality of the gene was assessed in a cell-cell binding assay as follows. COS-1 monkey cells [(clone M6; M. Horwitz *et al.*, Mol. Appl. Genet., 2:147-149, (1983)] were transfected with 3/4FT using DEAE dextran followed by DMSO shock treatment and chloroquine incubation [L. Sompeyrac and K. Dana, Proc. Natl. Acad. Sci., 78:7575-7578 (1981); M. Lopata *et al.*, Nucleic Acids Res., 12:5707-5717, (1984); H. Luthman and G. Magnuson, Nucleic Acids
35 Res., 11:1295-1308, (1983)]. The transfected COS cells were suspended and quantitated for binding to a CHO line expressing E-selectin [G. Larsen *et al.*, J. Biol. Chem. 267:11104-11110, (1992)]. This assay confirmed that the COS cells transfected with 3/4FT can express the sialylated Lewis^x epitope on the cell surface.

EXAMPLE 3**EXPRESSION OF THE P-SELECTIN LIGAND PROTEIN****A. Expression of the P-selectin Ligand in LEC11 cells**

10 Functional P-selectin ligand was expressed in the SLe^x-positive Chinese hamster ovary (CHO) cell line LEC11 (Campbell, C. and Stanley, P. Cell 35:303-309 (1983) as follows: approximately 8 µg of plasmid containing the P-selectin ligand gene (pMT21:PL85, Example 1) was transfected into LEC11 cells. At 68 hours post-transfection, the cells were treated with 2.5 mM sodium butyrate for 4 hours. The cells were observed to induce P-selectin adhesion, as
15 determined using the 6-CFD labeled CHO:P-selectin cell binding assay (described in Example 4, section B). In contrast, neither LEC11 cells alone nor LEC11 cells transfected with a control plasmid induced P-selectin adhesion.

B. Expression of Soluble P-Selectin Ligand in COS cells

20 COS cells were transfected with 8 µg pED.sPSL.T7 (see Example 5C) and 4 µg pEA.3/4 FT plasmid of Example 2, 8 µg pED.sPSL.T7 alone, or 8 µg plasmid vector (pMT21) and 4 µg pEA.3/4 FT gene. Forty-five hr post-transfection, the cells were rinsed twice in PBS and incubated overnight at 37°C in serum-free DMEM minus phenol red (JRH Biosciences) supplemented with 2 mM L-glutamine, 100 U/ml penicillin and 100 µg/ml streptomycin.
25 Phenylmethylsulfonyl fluoride, aprotinin and NaN₃ were added to final concentrations of 1mM, 2 µg/ml and 0.02%, respectively, and the conditioned medium was centrifuged to remove all debris.

For immunoprecipitation experiments, the labeled soluble P-selectin ligand protein was produced by co-transfecting COS cells with pED.sPSL.T7 and pEA.3/4 FT. At forty-five hr post-transfection, the COS cells were labeled with 250 µCi/ml ³⁵S methionine (NEN) for 5 hours and
30 the medium was collected. Expression of sPSL.T7 protein was confirmed by immunoprecipitation with anti-T7 antibodies.

C. Expression of PACE-cleaved P-selectin ligand in COS Cells

35 COS cells were co-transfected with the pED.sPSL.T7 plasmid of Example 5(C), the pEA.3/4FT cDNA of Example 2, and a plasmid containing the PACE cDNA as set forth in SEQ ID NO:5. A parallel control co-transfection was done using only the pED.sPSL.T7 plasmid and the pEA.3/4FT plasmid. After 45 hours, conditioned medium from these transfected COS cells

was coated onto plastic dishes and binding to CHO:P-selectin cells (Example 4) was determined. An approximately two-fold increase in bound CHO:P-selectin cells was observed for dishes coated with medium containing the P-selectin ligand co-expressed with PACE, as compared with medium containing P-selectin ligand which had not been co-expressed with PACE. Amino acid sequencing of the N-terminus of purified sPSL.T7 protein from the PACE co-transfection showed that all of the ligand had been cleaved at the PACE consensus site (amino acids 38-41 of SEQ ID NO:1). Radiolabelling of co-transfected COS cells with ³⁵S-methionine and subsequent SDS-polyacrylamide gel electrophoresis and autoradiography showed that comparable quantities of the P-selectin ligand had been secreted in both co-transfections.

D. Expression of the P-selectin Ligand Protein in CHO Cells

A full-length form (amino acids 1-402) of the P-selectin ligand protein was expressed in the CHO(DUKX) cell line (Urlaub & Chasin, Proc. Natl. Acad. Sci. USA 77, 4216-4220 (1980)) as follows: approximately 25 µg of the pMT21:PL85 plasmid and approximately 8 µg of the pED.3/4FT (produced by restriction of pEA.3/4FT with EcoRI and XbaI and insertion of the resulting fragment into the pED plasmid) were co-transfected into CHO(DUKX) cells using the calcium phosphate method. Transfectants were selected for resistance to methotrexate. After two weeks, individual colonies were screened for SLe^x expression by using a conjugate of an anti SLe^x antibody (CSLEX-1, U.S. 4,752,569) and sheep red blood cells (sRBC) prepared by the chromic chloride method (Goding, J. W., J. Immunol. Methods 10:61-66 (1976)) as follows: sRBC were washed with 0.15M NaCl until the wash became clear and then a 50% suspension of sRBC was prepared in 0.15M NaCl. One ml of 0.01% chromic chloride solution was added dropwise while vortexing to 0.2 ml of a sRBC suspension containing 50 µg of CSLEX-1. After incubating at 37°C for 30 minutes, 10 ml of phosphate buffered saline (PBS) solution was added to the reaction. The conjugate was washed once before resuspending into 10 ml of PBS. The plates containing transfectants were washed with PBS and then 3 ml of PBS and one ml of the sRBC/CSLEX-1 conjugate was added to each plate. Positive colonies were red on a transilluminator and were picked into alpha medium with 10% fetal bovine serum. After two weeks, colonies were subjected to stepwise amplification using methotrexate at concentrations of 2, 10, 25, 100, 250 nM. The stable cell line obtained was designated CD-PSGL-1 (R3.4). Expression of the P-selectin ligand protein was confirmed by immunoprecipitation studies using the polyclonal anti-P-selectin ligand protein antibody of Example 7(A). The functionality of the P-selectin ligand protein produced by the CD-PSGL-1 (R3.4) cell line was tested by assaying the transfectants for binding to LEC-γ1 as in Example 4(A).

7 The sPSL.T7 protein was expressed in a stable CHO-PACE line which was already
expressing the cDNA encoding PACE as set forth in SEQ ID NO:5 under adenosine deaminase
selection (Kaufman, *et al.*, PNAS (USA) 83:3136-3140 (1986)). The psPSL.T7 (25 µg) and
pED.3/4FT (8 µg) plasmids were cotransfected into CHO-PACE cells using the calcium
phosphate method. Transfectants were selected for resistance to methotrexate, and individual
10 colonies which bound to the sRBC/CSLEX-1 conjugate were picked. After two weeks in culture,
the colonies were subjected to stepwise amplification as described above. The stable cell line
obtained was designated CP/PSL-T7 (R4.1). Expression of sPSL.T7 protein was confirmed by
standard immunoprecipitation methods using either a T7 specific monoclonal antibody or the
LEC-γ1 chimera of Example 4(A). In a similar fashion, a stable cell line expressing the mature
15 full length form (amino acids 42-402) of the P-selectin ligand protein was obtained by co-
transfection of pMT21.PL85 and pED.3/4FT into the CHO-PACE line.

Stable cell lines expressing the sPSL.Q protein of Example 5(B) and the sPSL.Fc protein
of Example 5(D) were constructed as follows: plasmids pED.sPSL.Q (25 µg) or pED.sPSL.Fc
(25 µg) were cotransfected with approximately 25 µg of the pED.3/4FT plasmid described above
20 and approximately 20 µg of a plasmid containing the PACE cDNA as set forth in SEQ ID NO:5)
as well as the neomycin resistance gene into CHO(DUKX) cells using the calcium phosphate
method. Transfectants were selected for resistance to methotrexate and the G418 antibiotic.
Approximately two weeks later, individual colonies were screened for SLe^x expression using
sRBC/CSLEX-1 conjugate binding. The positive colonies were picked in G418 medium at 1
25 mg/ml concentration. After 2-3 weeks in culture, cells were amplified with methotrexate in a
stepwise selection. The stable cell lines obtained were designated CD-sPSL.Q (R8.2) and CD-
sPSL.Fc (R8.1), respectively. The expression of sPSL.Q and sPSL.Fc protein was confirmed by
standard immunoprecipitation method using the anti P-selectin ligand protein polyclonal antibody
of Example 7(A).

30

EXAMPLE 4

ASSAYS OF P-SELECTIN-MEDIATED INTERCELLULAR ADHESION

35 A. LEC-γ1 Binding Assay

A DNA encoding a chimeric form of P-selectin conjugated to the Fc portion of a human
IgGγ1 (LEC-γ1) was constructed using known methods (Aruffo *et al.* Cell 67: 35-44 (1991)), and

ably transfected into dhfr CHO cells (CHO DUKX) for high level production of the chimeric LEC- γ 1 protein, which was purified for use in the binding assay set forth below.

Petri dishes were coated first with a polyclonal anti-human IgG γ 1 Fc antibody and then with LEC- γ 1. This method orients the LEC- γ 1 construct such that the P-selectin portion of the chimeric molecule is presented on the surface of the plates. Adhesion of HL60 cells to the oriented LEC- γ 1 was quantitated in the presence and absence of calcium. HL60 adhesion was shown to be calcium dependent, confirming that the chimeric molecule had retained functional binding of P-selectin to its ligand on HL60 cells. The binding of HL60 cells to oriented LEC- γ 1 was also shown to be blocked by a neutralizing monoclonal antibody to P-selectin, demonstrating the specificity of P-selectin binding.

B. Fluorescent CHO-P-selectin Binding Assay

The assay employed a fluorescently labeled CHO:P-selectin cell line (Larsen *et al.*, J. Biol. Chem. 267, 11104-11110 (1992)) that can bind to and form clusters on the surface of COS cells that are co-transfected with the P-selectin ligand gene and the 3/4 FT gene. The CHO:P-selectin cells were suspended at 1.5×10^6 cells/ml in 1% fetal bovine serum in DME medium and labeled by adding 6-carboxyfluorescein diacetate (6-CFD) to a final concentration of 100 μ g/ml. After incubation at 37°C for 15 minutes, the cells were washed in medium and resuspended at 1×10^5 cells/ml. Five ml of the labeled cells were added to each washed COS transfectant-containing plate to be assayed and incubated at room temperature for 10 minutes. Nonadherent cells were removed by four washes with medium. The plates were then scanned by fluorescence microscopy for rosettes of adherent CHO:P-selectin cells.

C. Quantitative adhesion assay using radioactively labeled CHO:P-selectin cells

COS cells were co-transfected with the pMT21 PL85 plasmid of Example 1 and the pEA.3/4FT plasmid of Example 2 by the same procedure used in the initial stages of screening. As controls, COS cells were transfected with pMT21 PL85 alone, or with pEA.3/4FT alone, or with a similar plasmid containing no insert ("mock"). 24 hours post-transfection, the transfected cells were trypsinized and distributed into Costar 6-well tissue culture plates. CHO P-selectin cells were labeled for 16 hours with 3 H-thymidine using known methods and preincubated at 0.5×10^6 cells/ml for 30 minutes at 4°C in α medium containing 1% BSA (control); α medium containing 1% BSA, 5 mM EDTA and 5 mM EGTA; α medium containing 1% BSA and 10 μ g/ml of a neutralizing anti P-selectin monoclonal antibody; and α medium containing 1% BSA and a non-neutralizing anti-P-selectin monoclonal antibody. The preincubated cells were then

added to the wells containing the transfected COS cells. After a 10 minute incubation, unbound cells were removed by 4 changes of medium. The bound CHO-P-selectin cells were released by trypsinization and quantified by scintillation counting.

COS cells co-transfected with P-selectin ligand and the 3/4FT induced approximately 5.4-fold more binding of CHO-P-selectin cells relative to COS mock cells; assay in the presence of EGTA and EDTA reduced binding to the level of the mock transfected COS cells. Likewise, incubation with neutralizing anti-P-selectin antibody also eliminated specific binding, whereas non-neutralizing antibody had no effect. In contrast, the binding of CHO-P-selectin to COS cells transfected with P-selectin ligand alone was not statistically different than binding to the mock-transfected COS in both the presence or absence of EDTA and EGTA, or anti-P-selectin antibodies. The binding of CHO-P-selectin cells to COS cells transfected with 3/4 FT alone was approximately 2-fold greater than to the mock-transfected COS, but was unaffected by the presence or absence of EDTA and EGTA.

EXAMPLE 5

CONSTRUCTION OF SOLUBLE P-SELECTIN LIGANDS

The EcoRI adaptors used to generate the cDNA library from HL60 cells in Example 1 contain an XbaI restriction site (TCTAGA) just 5' of the beginning of SEQ ID NO:1 as it is located in the pMT21:PL85 plasmid. In order to generate soluble forms of the PSL, the pMT21:PL85 plasmid was restricted with XbaI and with HincII (which cleaves after nucleotide 944 of SEQ ID NO:1). The approximately 950 bp fragment thus generated, containing all of the encoded extracellular segment of the ligand up to and including the codon for valine 295, was isolated and used to generate DNAs encoding soluble forms of the P-selectin ligand protein as set forth in sections A through D below.

A. Construction of psPSL.QC

The fragment was purified and ligated into mammalian expression vector pED between the XbaI and EcoRI sites, along with double stranded synthetic oligonucleotide DNA that recreated the codons from Asn 296 to Cys 310 and introduced a novel stop codon immediately following Cys 310. The sequence of the oligos is as follows:

5'-AACTACCCAGTGGGAGCACCAGACCACATCTCTGTGAAGCAGTGCTAG (SEQ ID NO:12)
5'-AATTCTAGCACTGCTTCACAGAGATGTGGTCTGGTGCCTCCCACTGGGTAGTT (SEQ ID NO:13)

5 The resulting plasmid was designated pED.sPSL.QC, and the protein expressed from the plasmid was designated sPSL.QC.

B. Construction of psPSL.Q

10 The fragment was purified and ligated into the pED plasmid (Kaufman *et al.*, 1991) between the XbaI and EcoRI sites, along with the double stranded synthetic oligonucleotide DNA that recreated the codons from Asn 296 to Gln 309 and introduced a novel stop codon immediately following Gln 309. The sequence of the oligos is as follows:

5'-AACTACCCAGTGGGAGCACCAGACCACATCTCTGTGAAGCAGTAG (SEQ ID NO:14)

5'-AATTCTACTGCTTCACAGAGATGTGGTCTGGTGTCTCCCACTGGGTTAGTT (SEQ ID NO:15)

15 The resulting plasmid was designated pED.sPSL.Q, and the protein expressed from the plasmid was designated sPSL.Q.

C. Construction of psPSL.T7

20 Oligonucleotides encoding 14 amino acids including an epitope derived from the phage T7 major capsid protein were synthesized, creating a C-terminal fusion of the epitope "tag" with an additional 32 amino acids derived from the vector sequence. Two oligonucleotides having the sequences

5'-CTAGACCCGGGATGGCATCCATGACAGGAGGACAACAAATGGTAGGCCGTAG (SEQ ID NO:16) and

25 5'-AATTCTACGGCCTACCCATTTGTTGTCTCTCTGTCATGGATGCCATCCCGGGT (SEQ ID NO:17)

were duplexed and ligated with the large XbaI-EcoRI fragment of mammalian expression plasmid pED. The resulting plasmid, pED.T7 was restricted with XbaI and SmaI and ligated to the 950 bp XbaI-HincII fragment described above, resulting in plasmid pED.sPSL.T7.

30 The protein resulting from expression of pED.sPSL.T7 was designated sPSL.T7

D. Construction of Soluble P-selectin Ligand--IgGFc Chimera

The plasmid DNA encoding a soluble, extracellular form of the P-selectin ligand protein fused to the Fc portion of human immunoglobulin IgG1 was constructed as follows: the mammalian expression vector pED.Fc contains sequences encoding the Fc region of a human
35 IgG1 with a novel linker sequence enabling the fusion of coding sequences amino terminal to the hinge region via a unique XbaI restriction site. A three fragment ligation was performed: pED.Fc was restricted with XbaI and gel purified in linear form. The 950 bp fragment from pMT21 PL85

5 described above comprised the second fragment. The third fragment consisted of annealed synthetic oligonucleotide DNAs having the following sequence

5' - CTGCGGGCCGCGAGT (SEQ ID NO:18)

5' - CTAGACTGCGGGCCGCGAG (SEQ ID NO:19)

The ligation products were grown as plasmid DNAs and individual clones having the correct
10 configuration were identified by DNA sequencing. The plasmid was designated pED.PSL.Fc. The DNA coding region of the resulting soluble P-selectin ligand /Fc fusion protein is shown in SEQ ID NO:6.

EXAMPLE 6

15 CHARACTERIZATION OF EXPRESSED P-SELECTIN LIGANDS

A. Binding Characterization of Full-Length P-selectin Ligand Protein Expressed on COS Cells

Co-transfection of COS cells with the pEA.3/4FT plasmid of Example 2 and the
20 pMT21:PL85 plasmid of Example 1 yields COS cells which specifically bind to CHO:P-selectin cells. This binding is observed only upon co-transfection of pEA.3/4FT and pMT21:PL85; use of either plasmid alone generates COS cells which do not bind to CHO:P-selectin cells. No binding is observed between the parental CHO(DUKX) cell line which does not express P-selectin and COS cells co-transfected with pEA.3/4FT and pMT21:PL85. The binding between the co-
25 transfected COS cells and CHO:P-selectin cells is sensitive to chelators of divalent ions such as EDTA and EGTA, consistent with the Ca^{++} dependency of P-selectin mediated cellular adhesion. A neutralizing anti-P-selectin monoclonal antibody blocked the binding between the CHO:P-selectin cells and the COS cells which had been co-transfected with pEA.3/4FT and pMT21:PL85, while a non-neutralizing anti-P-selectin monoclonal antibody had no effect on the
30 binding. The antibody results indicate that the functional domain(s) of P-selectin are required for binding to P-selectin ligand protein expressed on the surface of COS cells.

B. Electrophoretic Characterization of Full-Length P-selectin Ligand Expressed in COS Cells

35 Detergent extracts of co-transfected COS cells were prepared as follows: 45 hours post co-transfection, approximately 1.5×10^7 cells were suspended in 5 ml of lysis buffer (10mM Piperazine-N,N'-bis[2-ethanesulfonic acid] (PIPES) pH 7.5, 100 mM KCl, 3 mM $MgCl_2$, 1 mM benzamidine, 0.5 μ g/ml leupeptin, 0.75 μ g/ml pepstatin, 1 mM ethylmaleimide, and 1 μ g/ml aprotinin) and lysed by sonication. Cellular debris was removed by low speed centrifugation (500
40 x g, 10 minutes), and a membrane fraction collected by ultracentrifugation (100,000 x g, 60 min).

5 The high speed membrane pellet was resuspended in an extraction buffer (10 mM 3-[N-Morpholino]propanesulfonic acid] (MOPS) pH 7.5, 0.1 M NaCl, 0.02% NaN₃, 1% Thesit® (Sigma), 1 mM benzamidine, 0.5 µg/ml leupeptin, 0.75 µg/ml pepstatin, 1 mM ethylmaleimide, and 1 µg/ml aprotinin). Samples were then subjected to SDS polyacrylamide gel electrophoresis and transfer to nitrocellulose blots as follows: an aliquot of the detergent extract was suspended
10 in 1% SDS loading buffer and heated for 5 minutes at 100°C before loading onto an 8-16% polyacrylamide gel (reduced) or a 6% gel (non-reduced) and electrophoresed in the Laemmli buffer system. Blots were prepared using Immobilon-P® transfer membranes. The blots were immersed in 10 mM MOPS pH 7.5, 0.1 M NaCl, 0.02% NaN₃, 1 mM MgCl₂, 1 mM CaCl₂, and 10% non-fat milk overnight at 4°C. Blots were rinsed once in the above buffer, minus the milk,
15 and incubated in blotting buffer (10 mM MOPS pH 7.5, 0.1 M NaCl, 1% bovine serum albumin, 0.05% Thesit, 1 mM MgCl₂, 1 mM CaCl₂) for 30 minutes at room temperature.

The blots were then probed for the P-selectin ligand as follows: 50 ng of a P-selectin/Fc chimera was pre-incubated with 3 µCi of ¹²⁵I-Protein A in blotting buffer for 30 minutes at room temperature. Additional excipients (e.g., EDTA, EGTA, monoclonal antibodies) could be added
20 to the pre-incubation mixture at this point to evaluate their effects on binding of the chimera to the P-selectin ligand. The pre-incubated mixture was then incubated with the blots (prepared as above) for 60 minutes at room temperature, and the blots were subsequently washed four times with the same blotting buffer (without bovine serum albumin), air dried, and autoradiographed at -70°C.

25 Under non-reducing conditions, two bands were observed with this technique for membrane extracts prepared from co-transfected COS cells. The major band migrated with an estimated molecular weight of approximately 220 kD, whereas the minor band migrated with a molecular weight of approximately 110 kD. Under reducing conditions, only a single band was observed with a molecular weight of approximately 110 kD, indicating that under non-reducing
30 conditions, the P-selectin ligand exists as a homodimer. The approximate molecular weight of the reduced monomer is greater than that predicted from the deduced amino acid sequence of the cDNA clone (45 kD), indicating that the expressed protein undergoes extensive post-translational modification (see Example 6(C)). The specificity of the P-selectin/Fc chimera was confirmed by the observation that a nonspecific IgG₁ probe yielded no bands on the blots. Additionally, the
35 binding of the P-selectin/Fc chimera to the blots was abolished by EDTA, EGTA, and a neutralizing anti-P-selectin monoclonal antibody. Specific bands on the blots were observed only from membrane extracts of COS cells co-transfected with the pEA.3/4FT and pMT21.PL85

- 5 plasmids. Membrane extracts from control transfections (pEA.3/4FT or pMT21 PL85 alone) failed to yield observable bands on blots.

C. Glycosylation of P-selectin Ligand Protein

10 The presence of covalently attached carbohydrate on recombinant P-selectin ligand and its role in binding to P-selectin was determined as follows: COS cells were co-transfected with pED.sPSL.T7 of Example 5(C) and the pEA.3/4FT plasmid of Example 2. After 48 hours, the cells were pulsed with ³⁵S-methionine. 200 µl of ³⁵S methionine-labeled sPSL.T7 conditioned medium was incubated with 5 µg LEC-γ1 in the presence of 2 mM CaCl₂ and 1 mg/ml bovine serum albumin (BSA). After rotating for 2 hours at 4°C, Protein A-Sepharose beads (Pharmacia) were added for 1 hour at 4°C, pelleted by centrifugation and washed twice in Tris buffered saline (20 mM Tris-HCl, 150 mM NaCl pH 7.5, hereinafter TBS) containing 2 mM CaCl₂ and 1 mg/ml BSA. The pellets were then resuspended and treated with neuraminidase (*Streptococcus pneumoniae*), O-glycanase, and N-glycanase (all from Genzyme) as follows. All glycosidase digestions were done at 37°C overnight. For neuraminidase digestion, the pellet was resuspended in 50 µl 2-(N-morpholino)-ethanesulfonic acid (MES) buffer, pH 6.5 (Calbiochem) and 0.1% SDS, heated at 95°C for 5 minutes, then pelleted. The supernatant was modified to contain 1.4% n-Octyl B-D-glucopyranoside (OGP), 10mM calcium acetate, 20 mM sodium cacodylate and 2.5 mM PMSF, final pH 7.0. Eight µl neuraminidase was added for a final concentration of 1 unit/ml. For neuraminidase/O-glycanase digestion, the sample was prepared as above and along with the neuraminidase, the O-glycanase was also added to a final concentration of 0.1 unit/ml. For N-glycanase digestion, the pellet was resuspended in 54 µl MES buffer and 1% SDS, heated at 95°C for 5 minutes, then pelleted. The supernatant was modified to contain 0.2 M sodium phosphate, 3.5% OGP, and 2.5 mM PMSF, final pH 8.5. N-glycanase was added for a final concentration of 12 units/ml and incubated as above.

30 The effect of glycosidase treatment on sPSL.T7 was assessed in two ways. For this, each digested protein sample was divided into two equal fractions. One fraction was precipitated with the P-selectin polyclonal antibody of Example 7(A), to show the effect of digestion on the electrophoretic mobility. The other fraction was precipitated with the LEC-γ1 chimera of Example 4(A), to assess the remaining P-selectin ligand binding activity after digestion. The immunoprecipitated samples were analyzed by SDS-polyacrylamide gel electrophoresis under reducing conditions and autoradiography.

35 In the absence of glycosidase treatment, autoradiography revealed comparable bands (with molecular weights of 110 kDa) for each precipitation. When the P-selectin ligand protein

5 was treated with neuraminidase, anti-P-selectin ligand polyclonal antibody precipitation revealed a slight decrease in mobility, consistent with removal of sialic acid residues. The amount of P-selectin ligand protein precipitated by LEC-γ1 was significantly reduced after neuraminidase treatment, consistent with the role of sialic acid residues in the P-selectin/P-selectin ligand interaction. When the P-selectin ligand protein was treated with both neuraminidase and O-glycanase, a substantial increase in electrophoretic mobility was observed after precipitation with the anti-P-selectin ligand polyclonal antibody, indicating that a number of O-linked oligosaccharide chains had been removed. However, removal of O-linked oligosaccharides from the P-selectin ligand protein may not have been complete, since the electrophoretic mobility did not correspond to a protein with a molecular weight of 38 kD, as would be predicted from the amino acid sequence set forth in SEQ ID NO:1. The neuraminidase/O-glycanase digested P-selectin ligand protein bound to LEC-γ1 very poorly, further indicating the role of oligosaccharides in the P-selectin/P-selectin ligand interaction. Treatment of the purified P-selectin ligand with N-glycanase resulted in a slight increase in electrophoretic mobility, demonstrating that some of the consensus sites for N-linked glycosylation are occupied. The amount of P-selectin ligand protein precipitated by LEC-γ1 was slightly reduced, indicating that N-linked glycosylation also contributes to the P-selectin/P-selectin ligand interaction, though not as dramatically as sialylation and O-linked glycosylation.

EXAMPLE 7

25 POLYCLONAL ANTIBODIES SPECIFIC FOR P-SELECTIN LIGANDS

A. Polyclonal Rabbit anti-P-selectin Ligand Protein/Maltose Binding Protein Fusion Protein

30 The anti-P-selectin ligand polyclonal antibody was generated by immunizing rabbits with a fusion protein generated in *E. coli*. The fusion protein consisted of the amino terminal one-third of the P-selectin ligand (amino acids 1 to 110 of SEQ ID NO 1) fused in frame to the maltose binding protein (Maina, C. V. *et al.*, Gene 74, 365-373 (1988); Riggs, P., in Current Protocols in Molecular Biology, F. M. Ausubel *et al.*, Eds., Greene Associates/Wiley Interscience (New York, 1990) chapter 16.6). Under conditions employed herein, the fusion protein antibody recognizes the P-selectin ligand protein.

B. Polyclonal Rabbit Anti-sPSL.T7 Protein

A soluble form of the invention (sPSL.T7; see example 5(C)) was purified to apparent homogeneity according to the following scheme. COS cells were transfected with three plasmids,

5 one encoding each of the following: sPSL.T7 (Example 5(C)), 3/4FT (Example 2), and a soluble form of PACE (as set forth in SEQ ID NO 5). After 72 hours, the conditioned medium was collected and recombinant sPSL.T7 was purified as follows.

Conditioned medium was diluted two fold with 50 mM MOPS, 150 mM NaCl, 0.5 mM CaCl₂ and 0.5 mM MnCl₂, pH 7.2, and applied to a column of lentil lectin-Sepharose 4B
10 equilibrated in the same buffer. After loading, the column was washed with the same buffer until the optical absorbance at 280 nm dropped to a stable baseline. The column was then eluted with the same buffer which had been adjusted to 0.5 M α -methyl-mannoside and 0.3 M NaCl. Recombinant sPSL.T7 was collected over 5-15 column volumes of this elution buffer. The lentil lectin eluate was then subjected to a 0-70% ammonium sulfate precipitation by adding 472g of
15 ammonium sulfate per liter of column eluate at 4°C. After stirring for 30 minutes, the precipitate was resuspended in a minimal volume of TBS (20 mM Tris-HCl, 150 mM NaCl, pH 7.5) and applied to a TSK G4000SW_{XL} gel filtration column equilibrated in TBS. The flow rate on the column was 0.5 ml/min and a guard column was employed. In aliquots of < 250 μ l, the resuspended ammonium sulfate pellet was injected on the column and fractions analyzed by SDS-
20 PAGE with Western analysis. Fractions containing sPSL.T7 were pooled and then used for immunizing rabbits.

Antibodies to sPSL.T7 were generated in the standard fashion by antigen priming and subsequent boosting over a 3 month period. Specifically, primary immunization was performed by mixing 50 μ g of sPSL.T7 (denatured by mixing in 0.1% SDS and heating for 10 minutes at
25 100°C) with complete Freund's adjuvant and injected at five sites subcutaneously. The second (and all subsequent) boosts were performed by mixing 25 μ g of sPSL.T7 (denatured by mixing in 0.1% SDS and heating for 10 minutes at 100°C) [12.5 μ g for the third and subsequent boosts] with incomplete Freund's adjuvant and injecting at two sites subcutaneously (or later, intramuscularly) every two weeks. Test bleeds were performed every two weeks to monitor
30 antibody titer. When the antibody titer reached a suitable level, a larger scale bleed was performed and a total serum fraction prepared. This polyclonal antibody preparation was used to inhibit the specific binding of HL60 cells to CHO P-selectin cells in a manner similar to that described in Example 4.

This assay employed fluorescently-labeled HL60 cells (labelled with BCECFAM; 2',7'-bis-(2-carboxymethyl)-5-(and-6)-carboxyfluorescein, acetoxymethyl ester) binding to CHO cells
35 plated on the bottom of microtiter plates. The labelled HL60 cells were pre-incubated with either sera containing polyclonal antibody or with pre-immune sera for 30 minutes at 4°C. The cells were then washed and incubated with the CHO P-selectin cells for 10 minutes. The plates were

5 then washed and the fluorescence read with a fluorescence microtiter plate reader. Using this assay, a 1:15 dilution of the anti-sPSL.T7 polyclonal serum resulted in essentially complete inhibition of HL60 cell binding to CHO:P-selectin. Demonstrable inhibition of HL60 binding to CHO:P-selectin was still observed at antiserum dilutions of 1:150. Pre-immune serum had no effect on HL60 cell binding to CHO:P-selectin.

10 EXAMPLE 8

COTRANSFORMATION WITH CORE2

A. Isolation of the cDNA encoding Core2 GlcNAc Transferase

The cDNA encoding core2 GlcNAc transferase was isolated by standard molecular biology techniques. Two oligos were designed at the 5' and 3' end (including translational
15 initiation and termination codon, respectively) based on the published human core2 sequence (Bierhuizen, M.F.A., Fukuda, M., Proc. Natl. Acad. Sci. 89, 9326-9330 (1992)). The pools of an HL60 cDNA library (Sako, D., Cell 75, 1179-1186 (1993)) were used as template to amplify the core2 coding sequence by a standard PCR protocol. The PCR amplified fragment was purified and subcloned into pED vector. To isolate cDNA, the pools which gave a positive signal in the
20 PCR reaction were transformed into *E. coli* and plated. Transformants were transferred onto nitrocellulose filters and hybridized with a ³²P radiolabelled PCR fragment according to standard protocols. Positive clones were picked and purified by replating. The sequence of the cDNA and PCR clone was confirmed by dideoxy sequencing.

25 B. Generation of Stable PSGL-1 Chinese Hamster Ovary Cell Lines Expressing Core2 Enzyme

A cell line made in accordance with the methods of Example 3 expressing full-length P-selectin ligand protein and 3/4 fucosyltransferase was co-transfected with core2 cDNA and a neomycin resistance gene (pMT4Neo) by standard calcium phosphate methods. After about two weeks, stable G418-resistant transfectants were picked either as single isolates or in a pool. These
30 transfectants were grown in 1 mg/ml G418 complete DMEM media and analyzed for core2 enzyme activity (Higgins, E.A., *et al.*, J. Biol. Chem. 266, 6280-6290 (1991)). Positive clones or pools found positive for core2 activity were analyzed for P-selectin ligand binding to P-selectin by various methods. In a similar fashion, cell lines expressing either P-selectin ligand protein or soluble P-selectin ligand protein with both the 3/4 fucosyltransferase and PACE enzymes (see
35 Example 3) were used to isolate stable cotransfectants of core2 as described above.

C. Effects of Core2 on P-selectin Binding Activity

The effects of core2 on P-selectin binding activity was evaluated by three different methods:

5 1 Binding of mPSGL-1 transfectants to immobilized soluble P-selectin or P-selectin/IgG chimera

48- well plates were coated with 1 ug/ml anti human Fc antibody in 50 mM Tris pH 9.5 at 4° C for five to six hours. After washing twice with HBSS buffer, P-selectin/IgG chimera (0.1-1 ug/ml conc., Example 5) was plated in HBSS buffer overnight at 4° C. The plates were blocked with BSA (3 mg/ml) at 4° C for three to four hours. In the case of soluble P-selectin ligand protein, the protein was coated directly onto plates in the same buffer. The ³H labelled CHO cells were lifted with 2 mM EGTA, washed three times with PBS, and resuspended to a final density of 10⁶ cells/ml. A 300 ul aliquot of this suspension was added to each well (300,000 cell/well). After incubating for 12 minutes at room temperature, wells were washed four times with serum free DMEM to remove unbound cells. Bound cells were lifted with 5 mM EGTA and counted in scintillation counter. U937 cells, used as a positive control for native P-selectin ligand protein binding, were pretreated with gamma globulin (5 mg/ml) to block endogenous Fc receptor before binding to P-selectin IgG chimera. Comparative binding data are shown in Fig. 1.

20 2. Immunoprecipitation of PSGL-1 with P-selectin/IgG Chimera

Recombinant full-length or soluble P-selectin ligand protein prepared from transformants, with and without additional core2, was labelled with ³⁵S-methionine and subsequently immunoprecipitated with either the anti P-selectin ligand protein polyclonal antibody or P-selectin ligand/IgG chimera as described previously in Examples 7 and 5; Sako, D., Cell 175, 1179-1186 (1993). Data are depicted in Fig. 2.

25 3. Flow Cytometry

Stable murine P-selectin ligand protein transfectants (with and without core2) were analyzed by standard FACS techniques using either P-selectin/IgG chimera (LecY1) (Example 5) or anti P-selectin ligand protein monoclonal antibody (MAb 275, raised against a peptide having the sequence from amino acid 42 to amino acid 56 of SEQ ID NO:2). Both reagents were pre-conjugated to FITC labelled Protein A. Cells were analyzed by FACS after incubating with this conjugate for 30 minutes at 4° C in the presence of 2 mM CaCl₂. Data are depicted in Fig. 3.

EXAMPLE 9

35 **E-SELECTIN BINDING OF P-SELECTIN BINDING PROTEIN**

E-Selectin/IgG chimera was made as described in Example 5 for the P-selectin IgG chimera using an E-selectin encoding DNA including amino acids -21 to 536 of the sequence reported in Bevilacqua *et al.*, Science, 243:1160 (1989).

5 U937 cells (approximately 6.5×10^7) were recovered from tissue culture plates and divided equally into two 50 mL cultures (final concentration of 1.3×10^6 cells/mL) containing fresh complete RPMI medium and 50 μ Ci/mL of 3 H-glucosamine hydrochloride (labels the protein-linked carbohydrate of glycoproteins [Varki, FASEB 5:226-235 (1991)]). After 48 hours incubation, the cells from both cultures were recovered by centrifugation and washed three times
10 with PBS. The pelleted cells were suspended in 2.5 mL each of a lysis buffer containing 1% Triton X-100 and disrupted by probe sonication for two minutes. The detergent lysates were placed on ice for three hours and then resonicated for an additional two minutes. The lysates were centrifuged at 16,000 rpm for five minutes, the supernatants were recovered and each adjusted to 12 mL with lysis buffer containing no detergent. To one of the two diluted cell lysates was
15 added 100 μ L of protein A sepharose precoupled with P-selectin/IgG chimera (see Example 5) and to the other was added 100 μ L of protein A sepharose precoupled with E-selectin/IgG chimera. Both chimeric proteins were present at a density of approximately 2 mg protein/mL of resin. Binding reactions were allowed to proceed overnight at 4 degrees C with end-over-end mixing. On occasion, purified membranes from U937 cells served as the starting material for the
20 detergent extraction of labeled proteins. In these cases, the detergent extraction and affinity precipitation steps were essentially identical to the above.

Following incubation, the two parallel reaction mixtures were each centrifuged at 2,000 rpm and supernatants were discarded. The resin pellets were washed four times with buffer (10 mM MOPS, 100 mM NaCl, 1 mM CaCl_2 , 1 mM MgCl_2 , 0.02% NaN₃, pH 7.5 with
25 Triton X-100 [0.25% for the first and second washes, 0.1% for the third wash and 0.01% for the fourth wash]). A final 1 mL pre-elution wash of each resin pellet using buffer containing 0.01% Triton X-100 was conducted and these were retained for quantitation of radioactive counts by liquid scintillation counting (LSC). The resins were then eluted overnight at 4 degrees C with end-over-end mixing in 1 mL each of buffer containing 0.01% Triton X-100 and 10 mM EDTA.
30 The supernatants were recovered by centrifugation and then quantitated by LSC.

Autoradiography of the materials released from the resins by EDTA was performed by electrophoresis of samples (approximately 10,000 cpm samples concentrated by Centricon-10 units where needed) on 10% cross-linked SDS-PAGE gels, subsequent treatment of the gels with ENHANCE (Dupont) as per the manufacturer's instructions followed by drying
35 for two hours on a commercially available gel dryer (Bio-Rad). Exposure of the dried gels to X-ray film was conducted for a minimum of three days at -80 degrees C.

Elution of immobilized E- or P-selectin, previously exposed to detergent extracts of U937 cells and exhaustively washed, with EDTA yielded liberated, 3 H-glucosamine labeled

5 proteins. The amount of radiolabel recovered from the EDTA eluates was at least 10-fold higher than the counts observed in the final, pre-EDTA washes. This observation suggests that both P- and E-selectin chimeras affinity captured ligand(s) from U937 whole cell lysates in an EDTA-dependent manner and that captured ligands were subsequently released upon treatment of the resins with EDTA.

10 The evaluation of the proteins released by EDTA from the two chimeras was performed by SDS-PAGE and autoradiography under reducing and non-reducing conditions (commercially available ^{14}C -labeled molecular weight standards were employed). As shown by the autoradiograph depicted in Fig. 4, the released counts from the whole cell lysates treated with the P-selectin chimera (lanes 2 and 10) and the E-selectin chimera (lanes 4 and 8) correlated to
 15 a major species of 200 kD molecular weight, non-reduced (lanes 2 and 4), and 100 kD reduced (lanes 8 and 10). In different experiments depicted in Fig. 4, where purified membrane extracts were used as the starting material in place of whole cells, both the E-selectin chimera (lane 3, non-reduced and lane 9, reduced) and the P-selectin chimera (not shown) gave similar results. Other experiments have demonstrated that the major U937 glycoprotein which binds to P-selectin is
 20 immunoreactive with Rb3026, a polyclonal antibody raised against recombinant sPSGL1.T7. Therefore, P- and E-selectin specifically recognize a single major glycoprotein species with identical properties in each case.

Example 10

25 Production and Analysis of Deleted or Altered Forms of Soluble P-Selectin Ligand Protein

A. Generation of DNA Constructs

Truncated forms of the P-selectin ligand protein-IgG chimeras were generated as follows. Plasmid pED.PSL.Fc was restricted with PstI and NotI and the 6kb fragment comprising the Fc
 30 portion and vector, pEDFc6kb, was gel purified. Plasmid constructs pED.149 Fc, pED.47 Fc and pED.19 Fc were created by standard PCR technique, using the following pairs of oligonucleotide primers:

"Upstream" primer for all constructs:

5'-CCAGGTCCAACCTGCAGGTCGACTCTAGAGGGCACTTCTTCTGGGCCCCACG-3' (SEQ ID NO.20)

35 "Downstream" primer for 148Fc:

5'-TATTATCTGTGCGGCCGCCCTCCAGAACCCATGGCTGCTGGTTGCAGTGG-3' (SEQ ID NO.21)

"Downstream" primer for 47Fc:

5'-TATTATCTGTGCGGCCGCCAGCAGGCTCCACAGTGGTAG-3' (SEQ ID NO.22)

"Downstream" primer for 19Fc:

5 5'-TATTATCTGTGCGGCGCGGAGGCTCCGTTTCTGGCAG-3' (SEQ ID NO:23)

The template DNA for PCR reaction was pED.PSL.Fc. The PCR conditions were 94°C, 1 min., 42°C, 1 min., 72°C, 3 min., 25 cycles, using a Perkin-Elmer Thermocycler. After completion of the last cycle, the reaction was treated with Klenow enzyme at 25°C for 30 min., extracted with phenol chloroform, sodium acetate added to 0.3M, and the PCR product DNA was precipitated
10 with 2.5 volumes of ethanol. The DNA pellet was rinsed with 70% ethanol and residual ethanol was evaporated. The resuspended DNA was digested with PstI and NotI, gel purified and ligated with the pEDFc6kb fragment described above. Correct constructs were identified by restriction analysis and confirmed by DNA sequencing.

Plasmid pED.ΔY148.Fc, pED.H24.Q70.148.Fc were created by site directed mutagenesis
15 (Maniatis et al., 1989, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratories) using pED.148Fc as template and the following mutagenesis oligonucleotides:
for ΔY148: 5'-CGGAGACAGGCCACCGAATTCCTGCCAGAAACG-3' (SEQ ID NO:24)
for H24: 5'-CCTCCAGAAATGCTGAGGCACAGCACTGACACCACTCCTC-3' (SEQ ID NO:25)
for Q70: 5'-GAGCTGGCCAACATGGGGCAACTGTCCACGGATTCAGCAG-3' (SEQ ID NO:26)
20 Positive clones were identified by colony hybridization (Maniatis et al. supra).

pED.FFFE.148.Fc was constructed by restricting pED.ΔY148.Fc with EcoRI and ligating the following duplexed oligonucleotides:

5'-AATTCGAGTTCCTAGATTTTG-3' (SEQ ID NO:27) and

5'-AATTCAAAATCTAGGAACTCG-3' (SEQ ID NO:28).

25 Constructs of the series pED.FYYD.19.Fc, pED.FFYD.19.Fc and pED.FFFD.19.Fc, were made by restricting pED.ΔY148.Fc with EcoRI and NotI and ligating the following duplexed oligonucleotides:

for pED.FYYD.19.Fc:

5'-AATTCGAGTACCTAGATTATGATTTCTGCCAGAAACTGAGCCTCCGC-3' (SEQ ID NO:29)

30 and

5'-GGCCGCGGAGGCTCAGTTTCTGGCAGGAAATCATAATCTAGGTAAGTGG-3' (SEQ ID NO:30);

for pED.FFYD.19.Fc:

5'-AATTCGAGTTCCTAGATTATGATTTCTGCCAGAAACTGAGCCTCCGC-3' (SEQ ID NO:31)

and

35 5'-GGCCGCGGAGGCTCAGTTTCTGGCAGGAAATCATAATCTAGGAACTCG-3' (SEQ ID NO:32).

for pED.FFFD.19.Fc:

5'-AATTCGAGTTCCTAGATTTCGATTTCTGCCAGAAACTGAGCCTCCGC-3' (SEQ ID NO:33)

and

5 5'-GGCCGCGGAGGCTCAGTTTCTGGCAGGAAATCGAAATCTAGGAAGTCG-3' (SEQ ID NO 34)

B. Plate Binding Assay for Analysis of Deleted or Altered Forms of Soluble P-Selectin Ligand Protein

The individual plasmid DNAs encoding the various mutated forms of soluble PSGL-1/Fc chimeras were co-transfected with pEA.3/4FT and PACE cDNA in COS cells as described in
 10 Example 3(c). 50mls of serum free medium, collected 40-64 hours post transfection from approximately 10^7 COS cells, was purified on a column of 0.25ml of protein A sepharose (Pharmacia) equilibrated with TBS supplemented with 2mM CaCl_2 . After washing with 20mls of TBS/ CaCl_2 , the bound material was eluted with 0.5mls of 0.1M acetic acid, 0.15M NaCl, 2mM CaCl_2 . The eluted material was neutralized with 1/20th volume 3M Tris pH 9.0. The
 15 material was quantitated by measuring absorbance at 280nm and by comassie blue staining of PAGE/SDS/Laemmli gels.

In order to produce non-sulfated forms of soluble PSGL-1, COS cell transfections of the relevant Fc chimeras were performed as described above except that following transfection the cells were cultured in the presence of 50mM Chlorate (Sigma).

20 Quantitative adhesion of CHO:P-selectin, CHO:E-selectin and CHO:L-selectin expressing cells was performed as described in Example 4(c), with the following modifications: COS cell and antibodies were omitted. Instead, 48-well microtiter plates (Costar) were coated for 16 hours at 4°C with varying quantities of protein A-purified soluble PSGL-1/Fc chimeras. The unbound material was removed and the coated wells were treated with Hank's buffered saline (HBS) with
 25 1mg/ml BSA and 2mM CaCl_2 for 1 hour at 4°C. Tritium labeled CHO selectin expressing cells were added and binding quantitated as described in Example 4(c).

C. Effects of Alteration of N-Linked Glycosylation Sites

30 Constructs expressing three P-selectin ligand-IgG chimeras were constructed to examine the effects of N-linked glycosylation sites on selectin binding. These constructs had the following characteristics:

- | | |
|----------------|---|
| 148.Fc | amino acids 42-189 of SEQ ID NO:2 |
| Q70.148.Fc | amino acids 42-189 of SEQ ID NO:2, with the asparagine residue at position 111 of SEQ ID NO:2 replaced with a glutamine residue |
| H24.Q70.148.Fc | amino acids 42-189 of SEQ ID NO:2, with the asparagine residue at position 65 of SEQ ID NO:2 with a histidine residue and the asparagine residue at position 111 of SEQ ID NO:2 replaced with glutamine residue |
- 35

These constructs are schematically represented in Fig. 6

The binding of these constructs to protein A and P-selectin-IgG chimera (LEC-γ1) was compared. The results of these experiments are shown in Fig. 7. Comparison of lanes 4, 5 and 6 in the autoradiograph demonstrates that removal of one or both of the first two N-linked glycosylation sites in soluble P-selectin ligand protein does not significantly effect its binding to P-selectin.

D. Effects of Tyrosines

Constructs were made to examine the role of tyrosine in P-selectin ligand protein binding to selectins by alteration of the anionic region of the soluble protein. The following constructs were made:

ΔY148.Fc amino acids 42-189 of SEQ ID NO:2, with amino acids 46-52 deleted

FFFE.148.Fc amino acids 42-189 of SEQ ID NO:2, with the tyrosine residues at positions 46, 48 and 51 replaced with phenylalanine residues and the aspartic acid residue at position 52 replaced with a glutamic acid residue

These constructs are schematically represented in Fig. 8.

The degree and sites of sulfation of P-selectin ligand protein were examined by expressing relevant constructs in the presence of radioactively labelled sulfate. The degree of sulfation of 148.Fc and ΔY.148.Fc were compared to that of a P-selectin-IgG chimera, which was not sulfated. Results are depicted in Fig. 9. These data demonstrate that the majority of sulfate incorporation is into the anionic region of the P-selectin ligand protein.

Additional constructs were made to determine whether the sulfation of the anionic region occurred at the tyrosine residues. The following additional constructs were made:

FYYD.19.Fc amino acids 42-60 of SEQ ID NO:2, with the tyrosine residue at position 46 of SEQ ID NO:2 replaced with a phenylalanine residue

FFYD.19.Fc amino acids 42-60 of SEQ ID NO 2, with the tyrosine residues at positions 46 and 48 of SEQ ID NO 2 replaced with a phenylalanine residues

FFFD.19.Fc amino acids 42-60 of SEQ ID NO 2, with the tyrosine residues at positions 46, 48 and 51 of SEQ ID NO 2 replaced with phenylalanine residues

These constructs are schematically represented in Fig. 9.

The degree of sulfation of these constructs was compared to 19.Fc ("FYYD.19.Fc"). Results are shown in Fig. 10. FYYD.19.Fc showed significant sulfation while FFFD.19.Fc was

5 substantially less sulfated. Thus, the tyrosine residues of the anionic region of P-selectin ligand proteins are the major site of sulfation.

Removal of sulfate from P-selectin ligand protein substantially reduces its binding to P-selectin. The binding of 148.Fc treated with chlorate to P-selectin was examined. As shown in Fig. 11, inhibition of sulfation by chlorate treatment substantially reduced the amount of P-selectin ligand protein binding to P-selectin.

E. Effects of C-terminal Deletions

Several additional C-terminal deleted constructs were made as follows:

254.Fc amino acids 42-295 of SEQ ID NO:2

47.Fc amino acids 42-88 of SEQ ID NO:2

15 19.Fc amino acids 42-60 of SEQ ID NO:2

These constructs are schematically represented in Fig. 12.

The binding of 254.Fc, 148.Fc, 47.Fc and 19.Fc to P-selectin, E-selectin and L-selectin was tested. Figs. 23 and 24 compare the binding of these deletion chimeras to selectins and controls. Results are also summarized in Fig. 12.

F. Binding to P-selectin and E-selectin Expressing Cells

Binding of various constructs described above to cells expressing P-selectin and E-selectin was compared using a quantitative plate binding assay of Example 4(c) (which is schematically described in Fig. 13).

Fig. 14 compares the binding of 148.Fc, Δ Y.148.Fc, FFFE.148.Fc and human IgG1 to P-selectin expressing CHO cells. Deletion of all of the tyrosine residues in the anionic region in Δ Y.148.Fc eliminated binding. Changing the tyrosine residues to phenylalanine residues in FFFE.148.Fc substantially reduced binding as compared to 148.Fc. Thus, it was demonstrated that the presence of the full length anionic region is essential to P-selectin binding and that P-selectin binding is enhanced by sulfation in this region. Fig. 14 also reports control experiments demonstrating that 148.Fc, Δ Y.148.Fc and FFFE.148.Fc do not bind to CHO cells which do not express selectin.

Fig. 15 compares the binding of 148.Fc, Δ Y.148.Fc, FFFE.148.Fc and human IgG1 to E-selectin expressing CHO cells. E-selectin binding was unaffected by the deletions or alterations of the native sequence. Thus, it was demonstrated that the anionic region is not required for E-selectin binding.

Fig. 16 summarizes the results of Figs. 14 and 15

5 Fig. 17 compares the binding of 47.Fc to P- and E-selectin expressing CHO cells. 47.Fc demonstrated substantial binding to both selectins despite deletion of the N-linked glycosylation sites at positions 111 and 292 of SEQ ID NO:2.

 Fig. 19 compares the binding of FYYD.19.Fc, FFFD.19.Fc, H24.Q70.148.Fc, 148.Fc, and human IgG1 to P-selectin expressing CHO cells. Replacement of all of the tyrosine residues
10 in the anionic region in FFFD.19.Fc eliminated binding. Changing the tyrosine residue at position 46 to a phenylalanine residue in FYYD.19.Fc substantially reduced binding as compared to 148.Fc. Alteration of the N-linked glycosylation sites in H24.Q70.148.Fc did not affect binding. Thus, it was demonstrated that P-selectin binding is enhanced by sulfation in the anionic region and that N-linked glycosylation is not required for P-selectin binding. Fig. 19 also reports control
15 experiments demonstrating that FYYD.19.Fc, FFFD.19.Fc, H24.Q70.148.Fc and 148.Fc do not bind to CHO cells which do not express selectin more than human IgG1 alone.

 Fig. 20 compares the binding of FYYD.19.Fc, FFFD.19.Fc, H24.Q70.148.Fc, 148.Fc, and human IgG1 to E-selectin expressing CHO cells. Truncation of the ligand protein to the degree of FYYD.19.Fc and FFFD.19.Fc substantially reduced E-selectin binding. Alteration of
20 the N-linked glycosylation sites in H24.Q70.148.Fc did not significantly affect E-selectin binding. Thus, it was demonstrated that P-selectin ligand proteins comprising amino acids 42 to 60 of SEQ ID NO:2 can selectively bind P-selectin, and, to a substantially less, extent E-selectin.

 Fig. 21 summarizes the results of Figs. 19 and 20.

G. Conclusions Regarding P- and E-Selectin Binding

25 Although applicants do not wish to be bound by any theory, these data allow several conclusion regarding the relationship between P-selectin binding and E-selectin binding by P-selectin ligand proteins. N-linked carbohydrates are not required for binding of a P-selectin ligand protein to either P- or E-selectin. P-selectin ligand proteins as small comprising as little as amino acids 42-60 of SEQ ID NO:2 are capable of binding to P-selectin, and, to a substantially less,
30 extent E-selectin.

 Fig. 22 depicts a proposed schematic model for binding of P-selectin ligand proteins to P- and E-selectin. O-linked sLe^x carbohydrate has been demonstrated to be required for both P- and E-selectin binding. Data presented herein demonstrate that sulfated tyrosine residues are implicated in P-selectin binding, but not E-selectin binding. Applicants' data also suggests that
35 no N-linked glycosylation binding site is required.

5

Example 11

Examination of Aggregation Phenomena and Dimer Formation in Forms of PSGL-1

A panel of PSGL-1 mutants were constructed by site-directed mutagenesis and/or PCR
amplification with primers that introduced a stopcodon. The template for all mutagenesis
10 experiments was pPL85.R16 (ATCC 75577, deposited by applicants).

The first group of mutants (C310S and C327S) encode full-length PSGL-1.R16 with only
one amino acid change compared to wild-type PSGL-1.R16 (Cys to Ser at position 310 or 327,
respectively). COS cells, co-transfected with pEA.3/4FT and the mutants C310S or C327S, were
labeled with ³⁵S-methionine. Cell lysates were prepared and the mutant proteins were
15 immunoprecipitated with the P-selectin ligand polyclonal antibody of Example 7(A) and analyzed
by SDS-polyacrylamide gel electrophoresis under non-reducing and reducing conditions.

The mutant C327S as well as wild-type PSGL-1.R16 migrated as a homodimer under
non-reducing conditions and as a monomer under reducing conditions. In contrast, the mutant
C310S migrated as a monomer both under non-reducing and reducing conditions, indicating that
20 the cysteine at position 310 is required for dimer formation of PSGL-1.

Both mutants were also analyzed for their ability to bind to P-selectin. Detergent extracts
of co-transfected COS cells were precipitated with the LEC-γ1 chimera of Example 4(A). The
precipitates were analyzed by SDS-PAGE under non-reducing and reducing conditions and by
autoradiography. Both PSGL-1.R16 and C327S were efficiently precipitated by LEC-γ1, whereas
25 C310S binding to LEC-γ1 was greatly reduced, indicating that the dimeric form of PSGL-1 binds
P-selectin more tightly than the monomeric form.

The second set of mutants encode soluble forms of PSGL-1.R16 and are listed in Table
1. The mutant ΔTM was generated by site-directed mutagenesis and has a deletion of the
transmembrane domain (amino acids 313-333) followed by RLSRKA. The mutants L311, L312,
30 A313, I314, L315, A318 and T322 were generated by site-directed mutagenesis or PCR
amplification with PCR primers that introduced a stop codon in the desired position. The name of
the mutant refers to the C-terminal amino acid of each truncated soluble form of PSGL-1.R16.
The mutants were analyzed according to the following criteria:

1. Expression and secretion from transfected COS cells
- 35 2. Dimer versus monomer formation
3. Lack of aggregate formation
4. P-selectin binding (LEC-γ1 chimera)

5 The mutants Δ TM and I316 fulfilled all four criteria. The shorter soluble forms of PSGL-1, such as sPSL.QC of Example 5(A), L311, L312, A313, I314 and L315 did not form dimers as well and the longer soluble forms of PSGL-1, such as sPSL.T7 of Example 5(C), A318 and T322 formed high molecular weight aggregates which were less desirable.

10 CHO cells, already expressing 3/4 fucosyltransferase and Core2 transferase, were transfected with psPSL.T7, Δ TM, I316 or psPSL.QC and amplified using methotrexate. Stable clones were isolated and labeled with 35 S-methionine. Conditioned media was either analyzed directly or first precipitated with LEC- γ 1 and then analyzed by SDS-PAGE under non-reducing and reducing conditions (Figure 25). The results indicated that Δ TM and I316 were most efficient in dimer formation and P-selectin binding.

15

Table I

20	Mutant	Dimer Formation	High MW Aggregates	P-selectin binding
	PSL.QC	+	-	+
	L311	+	-	+
25	L312	-	-	-
	A313	-	-	-
30	I314	-	-	-
	L315	+	-	+
	I316	++	-	++
35	A318	++	+	++
	T322	++	+	++
40	Δ TM	++	-	++

Example 12

Specificity of PSGL-1 Binding to P- and E-Selectins

45 Materials. A chimeric protein comprising the extracellular domain of human E-selectin and the Fc portion of human IgG₁ was constructed analogously to the P-selectin chimera, LEC- γ 1, described earlier. The soluble E-selectin chimera was expressed in baculovirus-infected

- 5 *Trichoplusia ni* high five cells (Invitrogen) and purified to homogeneity by Protein A Sepharose chromatography. Plasmid vectors pEA 3/4FT, pPL85, pFCD43, and pEA.sPACE, for COS expression of $\alpha(1,3/1,4)$ -fucosyltransferase (Fuc-TIII), PSGL-1, CD43 (leukosialin), and soluble paired basic amino acid converting enzyme (PACE), respectively, have been described herein and in the literature (Sako et al. (1993) *Cell* 75, 1179-1186; Rehemtulla, A. & Kaufman, R. J. (1992) *Curr. Opin. Biotechnol.* 3, 560-565; Wasley et al. (1993) *J. Biol. Chem.* 268, 8458-8465).
- 10 Fuc-TVII cDNA (plasmid pMT.FT7) was cloned from an HL60 cDNA expression library using oligonucleotide probes derived from the published sequence (Natsuka et al. (1994) *Journal of Biological Chemistry* 269, 16789-16794; Sasaki et al. (1994) *J. Biol. Chem.* 269, 14730-14737). A polyclonal neutralizing rabbit antibody, Rb3443, was raised against a peptide comprising the
- 15 first 15 amino acids of the mature (PACE-cleaved) N-terminus of PSGL-1. Monoclonal anti-CD43 antibodies from either Becton Dickinson or Biodesign International and isotype control antibodies were coupled to a solid support consisting of Sepharose TM-4B with a covalently attached goat affinity-purified antibody to mouse IgG (Cappel, Organon Teknika Corporation). Affinity coupling of selectin chimeras and murine antibodies to Protein A Sepharose 4 Fast Flow
- 20 (Pharmacia) and to the anti-mouse IgG resin, respectively, was carried out at a ratio of 2 mg protein/ml of resin. Antiserum Rb3443 was coupled to Protein A Sepharose at 1 ml/ml resin. Coupling efficiencies, indicated by micro-BCA assay (Pierce) of the post-reacted supernatants, were at least 95%. Aprotinin and pepstatin were from Boehringer Mannheim and benzamidine, leupeptin, and phenylmethylsulfonyl fluoride (PMSF) were from Sigma.
- 25 Labeling and Membrane Extraction of Myeloid Cells. U937 or HL60 cells grown in suspension to a density of $\sim 1.3 \times 10^6$ cells/ml were labeled in 50 ml of RPMI1640 medium supplemented with 10% fetal bovine serum and 2.5 mCi of ^3H -glucosamineHCl (Dupont/NEN) for 48 hr. Activities of greater than 1 cpm/cell were routinely obtained by this technique. The
- 30 labeled cells were washed with PBS, resuspended in cell lysis buffer (10 mM MOPS, 150 mM NaCl, 4 mM CaCl_2 , and 4 mM MgCl_2 , pH 7.5 containing protease inhibitors 20 mg/ml aprotinin, 10 mM benzamidine, 20 mg/ml leupeptin, 8 mg/ml pepstatin, and 10 mM PMSF) and subjected to several cycles of probe sonication on ice. Nuclei and cell debris were removed by low speed centrifugation and the cell membranes recovered from the supernatant by centrifugation at 100,000g RCF for 1 hr, washed by resuspension and high speed centrifugation in cell lysis buffer
- 35 containing 1 M NaCl, and finally resuspended in 3 ml membrane solubilization buffer (cell lysis buffer containing 1% Triton X-100). Several cycles of sonication and incubation on ice were employed to solubilize the membrane fraction. Finally, a low speed centrifugation step was employed to remove insoluble membrane residue.

5 Labeling and Membrane Extraction of Transfected COS Cells. COS M6 cells were transfected using DEAE-dextran and chloroquine (25) employing 8 µg of plasmids pPL85 or pFCD43 and 4 µg of pEA.sPACE, as well as 4 µg of pEA.3/4FT or pMT.FT7. After 40-45 hr recovery the transfected cells were starved in serum- and methionine-free DME medium for 30 min and then fed [³⁵S]-methionine in serum-free DME for 5 hr. The labeled cells were washed,
10 incubated with EGTA to loosen them from the dish surface, scraped from the dish, pelleted, and suspended in cold 10mM PIPES buffer, pH 7.5, containing 100 mM KCl, 3 mM NaCl, 3.5 mM MgCl₂, and protease inhibitors (see above). Membrane extraction then was carried out by sonication, low speed centrifugation, high speed centrifugation, and solubilization in membrane lysis buffer as above, for labeled myeloid cells.

15 Affinity Precipitations. Membrane extracts were diluted 1:4 or 1:5 with cell lysis buffer or with TBSC buffer (20 mM TrisHCl, 150 mM NaCl, 2 mM CaCl₂, pH 7.5) supplemented with 5 mg/ml bovine serum albumin (approximately 99%, Sigma). Extracts thus diluted to 0.2-0.25% Triton X-100 were incubated with human IgG₁-Protein A Sepharose with end-over-end mixing at 4°C overnight. The precleared supernatants then were reacted for 6-12 hrs at 4°C with Protein
20 A Sepharose precoupled with E- or P-selectin chimeras, control human IgG₁, Rb3443 or with rabbit pre-immune serum or with anti-CD43 antibody or isotype control precoupled to goat anti-mouse IgG Sepharose. The resins were washed 5 or more times in buffer containing 0.1-0.5% Triton X-100 until the radioactivity of the wash supernatants was reduced to background level. Elution of proteins bound specifically to P- or E-selectin resins was accomplished with 10 mM
25 EDTA or 5 mM EDTA/5 mM EGTA at room temperature or by boiling in SDS-PAGE sample buffer (Laemmli, U. K. (1970) *Nature* **227**, 680-685), whereas elution of proteins bound to antibody resins was achieved exclusively by the latter means. For resolution under reducing conditions, dithiothreitol was added to the sample buffer to a final concentration of 100 mM. Samples thus prepared were resolved by SDS-PAGE on 7.5% gels, treated with Enhance
30 (Dupont), dried, and exposed to autoradiography film.

For sequential affinity capture experiments, membrane extracts were precleared, affinity precipitated with P- or E-selectin or human IgG₁, and washed as above. Samples then were eluted twice from the resins with 5 mM EDTA in 10 mM MOPS, 150 mM NaCl, pH 7.5 for 1 hr at 4°C with tumbling. The first and second eluates were combined and then immunoprecipitated with
35 immobilized Rb3443 according to the protocols outlined above.

RESULTS

Soluble E- and P-selectin chimeras were used, in parallel with control human IgG₁ to probe detergent-solubilized membrane extracts of ³H-glucosamine-labeled U937 cells as described

5 under "Methods". Examination of eluates from the immobilized selectins by SDS-PAGE/autoradiography (Fig. 26) revealed the presence in both P-and E-selectin eluates of a major protein species with identical electrophoretic properties: Mr 200-kDa non-reduced with conversion to a species of Mr 120-kDa following reduction (Fig. 26, lanes 2 and 3, respectively). Occasionally, additional bands were observed in both E-and P-selectin eluates presumably
10 corresponding to this major band and reflecting the presence of naturally reduced material (the 120-kDa species in non-reduced samples) and incomplete reduction (the 200-kDa species in reduced samples). Additionally, a trace band of Mr 150-kDa in the E-selectin eluate which was unaffected by reduction with DTT was occasionally observed. No bands were observed in control experiments using immobilized human IgG₁ (Fig. 26, lane 1) or where elution of selectin resins
15 was performed in the absence of EDTA or SDS (data not shown). Essentially identical results were obtained using HL-60 cells (data not shown). Hence, the nature of these recognition events is interpreted to be specific metal-dependant interactions of these proteins with the respective selectins, presumably via the lectin domains (Lasky, L. A. (1992) *Science* **258**, 964-969; Drickamer, K. (1988) *J. Biol. Chem.* **263**, 9557).

20 The metal-dependant recognition and electrophoretic behavior of the major band precipitated with E-selectin was consistent with the properties of the previously identified P-selectin counterreceptor, P-selectin glycoprotein ligand or PSGL-1 (Moore et al. (1994) *J. Biol. Chem.* **269**, 23318-23327; Moore et al. (1992) *J. Cell Biol.* **118**, 445-456; Sako et al.).

To assess whether this species was indeed PSGL-1, EDTA eluates of the both E-and
25 P-selectin precipitates were subsequently reacted with the PSGL-1 specific polyclonal antiserum Rb3443. As shown in Fig. 27, the major band isolated by affinity capture with either selectin was immunoprecipitated using this antiserum (lanes 3 and 4, respectively). No species were detected after immunoprecipitation of the control IgG₁ EDTA eluate (Fig. 27, lane 5). Direct immunoprecipitations using fresh ³H-labeled U937 membrane extracts confirmed the specificity
30 of Rb3443: precipitation with Rb3443 results in the recovery of a single band with the electrophoretic properties of PSGL-1 (Mr 200-kDa non-reduced, Mr 120-kDa reduced, Fig. 27, lane 2) whereas precipitation with pre-immune antiserum fails to capture any material (Fig. 27, lane 1). These results indicate that the major protein species specifically captured from myeloid cells by both E-and P-selectins is PSGL-1.

35 To further assess the specificity of E-selectin for PSGL-1, U937 membrane lysates were probed directly for the presence of CD43 (or leukosialin), an abundant cell surface sialoglycoprotein known to bear the major portion of myeloid cell SLe^x residues (Maemura, K. & Fukuda, M. (1992) *J. Biol. Chem.* **267**, 24379-24386). Thus, membrane extracts of

5 ³H-glucosamine-labeled U937 cells were probed with an anti-CD43 antibody in parallel with the
PSGL-1 specific antiserum Rb3443 and control antibodies as described under "Methods". From
identical quantities of membrane lysate, the CD43 antibody precipitated in excess of 30-fold
greater radioactive counts than did the PSGL-1 antiserum. Evaluation of the immunoprecipitates
by SDS-PAGE/autoradiography (Fig. 28) revealed a single specific band for each antibody.
10 Rb3443 captured a single species with the electrophoretic characteristics of PSGL-1 (Fig. 28,
Lane 2). In contrast, the CD43 antibody precipitated a species with an Mr 120-kDa which was
insensitive to reduction (Fig. 28, Lane 4), consistent with the absence of cysteine in CD43.
Immunoprecipitations with control antibodies (Fig. 28, lanes 1 and 3) proved negative as
expected. There appears to be considerably greater quantities of CD43 than PSGL-1 in U937
15 cells, consistent with the quantitation of these proteins in HL-60 cells (Ushiyama et al. (1993) *J.*
Biol. Chem. **268**, 15229-15237). Thus, the inability of E-selectin to precipitate CD43 from
myeloid cells does not appear to be due to its absence in these cell lines. While we cannot exclude
the possibility that E-selectin captures trace quantities of CD43 (i.e., the low-intensity Mr
120-kDa band in Fig. 26, non-reduced lane 3 which is also consistent with monomeric PSGL-1),
20 PSGL-1 appears to be the major protein precipitated from myeloid membrane extracts.

Recombinant PSGL-1 expressed in COS cells is best achieved with cotransfection of the
PSGL-1 cDNA with a cDNA encoding an $\alpha(1,3/1,4)$ fucosyltransferase (Fuc-TIII) for P-selectin
binding (Sako et al.). Interestingly, initial efforts to demonstrate E-selectin recognition of
recombinant PSGL-1 failed: E-selectin was unable to capture the counterreceptor from
25 cotransfected COS cell membrane lysates under conditions where P-selectin capture was
successful. One interpretation of this result is that Fuc-TIII was able to modify recombinant
PSGL-1 for recognition by P-selectin but was unable to replicate the appropriate modification(s)
found in myeloid PSGL-1 necessary for E-selectin recognition. The recent cloning of a myeloid
fucosyltransferase, Fuc-TVII (Natsuka et al. (1994) *Journal of Biological Chemistry*, **269**, 16789-
30 16794; Sasaki et al. (1994) *J. Biol. Chem.* **269**, 14730-14737), that is also capable of generating
SLe^x carbohydrate structures, allowed evaluation of this interpretation.

COS cells were cotransfected with cDNAs encoding either PSGL-1 or CD43 and either
Fuc-TIII or Fuc-TVII. Membrane lysates were prepared from the transfected COS cells and these
were precipitated with either immobilized E- or P-selectin chimeras or with antibodies to either
35 PSGL-1 or to CD43. The precipitated products were evaluated by SDS-PAGE/autoradiography
following their release by EDTA/EGTA (for selectin mediated binding) or by boiling in SDS (for
immunoprecipitations). The results are shown in Fig. 29.

5 As observed in Fig. 29A, E-selectin capture of COS-expressed PSGL-1 was dependant upon the nature of the fucosyltransferase used in the transfection. In three separate experiments, Fuc-TVII, but not Fuc-TIII, supported PSGL-1 precipitation by E-selectin. The inability of Fuc-TIII to confer E-selectin reactivity to PSGL-1 cannot be attributed to a lack of PSGL-1
10 expression as the specific antiserum Rb3443 immunoprecipitated significant and comparable quantities of PSGL-1 from both Fuc-TIII and Fuc-TVII transfections (Fig. 29C). Furthermore, P-selectin was capable of precipitating equivalent quantities of PSGL-1 with either fucosyltransferase (Fig. 29B), demonstrating that Fuc-TIII and Fuc-TVII are expressed and active in these cotransfections.

Within the COS recombinant expression system as in myeloid cells, high-affinity
15 E-selectin recognition was also dependent upon the presence of an appropriate polypeptide. Although the polypeptide length, apparent molecular weight, and high frequency and specific types of posttranslational modifications are similar in CD43 and PSGL-1 (Maemura, K. & Fukuda, M. (1992) *J. Biol. Chem.* **267**, 24379-24386), neither fucosyltransferase was able to confer high-affinity E-selectin (or P-selectin) recognition to recombinant leukosialin in
20 cotransfected COS cells (Fig. 29A). Immunoprecipitations with the anti-CD43 antibody indicate that comparable quantities of leukosialin were expressed in both Fuc-TIII and Fuc-TVII cotransfections (Fig. 29C). The failure of E-selectin to capture CD43 was not due to lack of fucosyltransferase activity within the cotransfected COS cells. FACS analysis of COS cells transfected with either PSGL-1 or CD43 and either Fuc-TIII or Fuc-TVII all show high levels of
25 reactivity with the SLeX specific antibody CSLEX-1. Therefore, these results suggest that high-affinity E-selectin recognition requires the presence of a specific polypeptide(s) that is appropriately modified by a specific fucosyltransferase.

Example 13

Inhibition of P-Selectin/PSGL-1 Binding 30 by PSGL-1 Derived Peptides

A number of peptides derived from the sequence of PSGL-1 (SEQ ID NO 2) were tested for their ability to inhibit P-selectin/PSGL-1 binding. The tested peptides are listed in Fig. 30

Inhibition was tested according to the following protocol. The wells of a 96 well plate
35 were coated overnight at 4°C with PSGL-1 in 50 µl of 10 mM MOPS, 150 mM NaCl, 1 mM CaCl₂, 1 mM MgCl₂ at pH 7.5. After removal of the liquid from the wells, 150 µl of 10 mM MOPS, 150 mM NaCl, 1 mM CaCl₂, 1 mM MgCl₂, 0.05% tween-20, 0.05% gelatin at pH 7.5 was added per well to block the unoccupied sites. After 1/2 hr. to 2 hr. the block buffer was removed

5 from the wells and 100 μ l of a complex of Lec- γ 1 (P-selectin-human IgG Fc chimera) (2 μ g/ml),
biotinylated goat anti-human antibody, and streptavidin-conjugated alkaline phosphatase (which
had been allowed to tumble at room temperature for 30 minutes to 1 hour) plus any potential
inhibitors were added per well. Each plate was shaken and rapped to remove the block from plate.
The incubation proceeded for 1 hr at room temperature rotating in the dark. The unbound
10 complex was washed off the plate with two 150 μ l portions of 10 mM MOPS, 150 mM NaCl,
1 mM CaCl_2 , 1 mM MgCl_2 , 0.05% tween-20 followed by 150 μ l of 1M diethanolamine, 0.5 mM
 MgCl_2 . The chromogenic substrate for alkaline phosphatase, PNPP, in 10 mM DEA/0.5 mM
 MgCl_2 was added and the plate is then read at 405 nm.

The results of these assays are reported in Fig. 30. The peptides comprising amino acids
15 48-51 (in which the tyrosine residues have been phosphorylated) and amino acids 42-56 of SEQ
ID NO 2 provided particularly desirable results.

Example 14

Purification of a Soluble Form of PSGL-1

20 Substantial purification of a soluble form of P-selectin ligand protein has been achieved
according to the protocol described below.

A soluble P-selectin ligand protein, I316 (amino acid 42 to amino acid 316 of SEQ ID
NO:2) was expressed in CHO cells as described herein. CHO cell conditioned media was
concentrated with a Pellicon ultrafiltration membrane unit (Millipore) with either 10,000
25 molecular weight cutoff (MWCO) or 30,000 MWCO to about 10 times the original concentration.
The buffer was then exchanged into 25 mM Tris, 1mM CaCl_2 , pH 7.4.

The buffer-exchanged concentrate was loaded onto a Toyopearl QAE 550C (ToyoHaas)
column. Alternatively, the buffer exchange step can be omitted and the concentrate can be diluted
one part concentrate to three parts 25 mM Tris, 1mM CaCl_2 , pH 7.4, and then loaded onto the
30 column. The column was washed with 5-10 column volumes (CV) of 25 mM Tris, 1mM CaCl_2 ,
pH 7.4 at 4°C.

The P-selectin ligand protein eluted from the column with a linear NaCl gradient (0 M
NaCl to 1.0 M NaCl) in the 25 mM Tris, 1mM CaCl_2 , pH 7.4 buffer in approximately five column
volumes. Two peaks were eluted from the column. The second peak contained the P-selectin
35 ligand protein and was collected in bulk.

The peak from the QAE column was concentrated with a tangential flow ultrafiltration
membrane (Millipore) with a 30,000 MWCO and was then buffer exchanged into 25 mM Tris,
150 mM NaCl, 1mM CaCl_2 , pH 7.4 at 4°C.

5 The buffer exchanged concentrate was loaded onto a Jacalin Agarose column overnight at 4°C. The column was washed with the diafiltration buffer and the P-selectin ligand protein was eluted with a gradient of methyl α -D-galactopyranoside (0-100 mM or)-50 mM methyl α -D-galactopyranoside) at 20°C. Fractions from the Jacalin column were analyzed by SDS-PAGE and the purest fractions were pooled.

10

Example 15

P-Selectin Ligand Protein Fusions

Four fusions of a P-selectin ligand protein with a different amino acid sequence were constructed: 47.Fc, 47.AGP, 47.BMP and 47.IL11.

15 47.Fc: A cDNA was constructed encoding the signal peptide, PACE cleavage site and first 47 amino acids of the mature P-selectin ligand sequence fused to a mutated Fc region of human IgG1 at His224 of the native Fc sequence. The sequence of the cDNA construct is reported as SEQ ID NO:35. The fusion point is a novel NotI site at nucleotide 261. The amino acid sequence encoded by the cDNA construct is reported as SEQ ID NO:36. The mature amino acid sequence of the encoded fusion protein begins at amino acid 42 of SEQ ID NO:36. The mutations in the Fc portion were a change of Leu 234 and Gly237 of the native Fc sequence to Ala.

20 47.AGP: A cDNA was constructed encoding the signal peptide, PACE cleavage site and first 47 amino acids of the mature P-selectin ligand sequence fused to the first leucine residue of mature human AGP. The sequence of the cDNA construct is reported as SEQ ID NO:37. The fusion point is a novel NotI site at nucleotide 261. The amino acid sequence encoded by the cDNA construct is reported as SEQ ID NO:38. The mature amino acid sequence of the encoded fusion protein begins at amino acid 42 of SEQ ID NO:38.

25 47.BMP: A cDNA was constructed encoding the signal peptide, PACE cleavage site and first 47 amino acids of the mature P-selectin ligand sequence fused to the sequence of mature human BMP-2 (with its first 8 amino acids deleted). The sequence of the cDNA construct is reported as SEQ ID NO:39. The fusion point is a novel NotI site at nucleotide 261. The amino acid sequence encoded by the cDNA construct is reported as SEQ ID NO:40. The mature amino acid sequence of the encoded fusion protein begins at amino acid 42 of SEQ ID NO:40.

30 47.IL11: A cDNA was constructed encoding the signal peptide, PACE cleavage site and first 47 amino acids of the mature P-selectin ligand sequence fused to mature human IL-11. The sequence of the cDNA construct is reported as SEQ ID NO:41. The fusion point is a novel NotI site at nucleotide 261. The amino acid sequence encoded by the cDNA construct is reported as

- 5 SEQ ID NO 42. The mature amino acid sequence of the encoded fusion protein begins at amino acid 42 of SEQ ID NO 42.

Patent and literature references cited herein are incorporated as if fully set forth

SEQUENCE LISTING

1. GENERAL INFORMATION:

(i) APPLICANT: Larsen, Glenn
Sako, Dianne
Chang, Xiao Jia
Veldman, Geertruida M.
Cumming, Dale
Kumar, Ravindra
Shaw, Gray

(ii) TITLE OF INVENTION: NOVEL P-SELECTIN LIGAND PROTEIN

(iii) NUMBER OF SEQUENCES: 42

(iv) CORRESPONDENCE ADDRESS:

(A) ADDRESSEE: LEGAL AFFAIRS
(B) STREET: 87 CAMBRIDGE PARK DRIVE
(C) CITY: CAMBRIDGE
(D) STATE: MA
(E) COUNTRY: USA
(F) ZIP: 02140

(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.25

(vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER:
(B) FILING DATE:
(C) CLASSIFICATION:

(vii) PRIORITY APPLICATION DATA:

(A) APPLICATION NUMBER: US 07 965,662
(B) FILING DATE: 23-OCT-1992

(viii) PRIOR APPLICATION DATA:

(A) APPLICATION NUMBER: US 08-112,608
(B) FILING DATE: 26-AUG-1993

(ix) PRIORITY APPLICATION DATA:

(A) APPLICATION NUMBER: PCT/US93/10168
(B) FILING DATE: 20-OCT-1993

(x) PRIOR APPLICATION DATA:

(A) APPLICATION NUMBER: US 08/235,398
(B) FILING DATE: 28-APR-1994

(xi) PRIORITY APPLICATION DATA:

(A) APPLICATION NUMBER: US 08 318,305
(B) FILING DATE: 30-SEP-1994

(vii) PRIOR APPLICATION DATA:

- (A) APPLICATION NUMBER: US 09/423,734
 (B) FILING DATE: 25-APR-1995

(viii) ATTORNEY/AGENT INFORMATION:

- (A) NAME: BROWN, SCOTT A.
 (B) REGISTRATION NUMBER: 32,724
 (C) REFERENCE/DOCKET NUMBER: GI 5213F

(ix) TELECOMMUNICATION INFORMATION:

- (A) TELEPHONE: (617) 498-8224
 (B) TELEFAX: (617) 876-5851

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1649 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Homo sapiens
 (G) CELL TYPE: Promyelocyte
 (H) CELL LINE: HL60

(vii) IMMEDIATE SOURCE:

- (B) CLONE: PMT21:PL85

(ix) FEATURE:

- (A) NAME/KEY: 5'UTR
 (B) LOCATION: 1..59

(ix) FEATURE:

- (A) NAME/KEY: CDS
 (B) LOCATION: 60..1268

(ix) FEATURE:

- (A) NAME/KEY: 3'UTR
 (B) LOCATION: 1269..1649

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

```

GCCACTTCTT CTGGGCCCCAC GAGGCAGCTG TCCCATGCTC TGCTGAGCAC GGTGGTGCC      59
ATG CCT CTG CAA CTC CTC CTG TTG CTG ATC CTA CTG GGC CCT GGC AAC      107
Met Pro Leu Gln Leu Leu Leu Leu Ile Leu Leu Gly Pro Gly Asn
   1             5             10             15

AGC TTG CAG CTG TGG GAG ACT TGG GCA GAT GAA GCG GAG AAA GCG TTG      155
Ser Leu Gln Leu Trp Asp Thr Trp Ala Asp Glu Ala Glu Lys Ala Leu
   20             25             30

```

GGT GCG CTG CTT GCG GCG GAC GCG AGA CAG GCG ACC GAA TAT GAG TAC Gly Pro Leu Leu Ala Arg Asp Arg Arg Gln Ala Thr Glu Tyr Glu Tyr 35 40 45	203
CTA GAT TAT GAT TTC CTG CCA GAA ACG GAG CCT CCA GAA ATG CTG AGG Leu Asp Tyr Asp Phe Leu Pro Glu Thr Glu Pro Pro Glu Met Leu Arg 50 55 60	251
AAC AGC ACT GAC ACC ACT CCT CTG ACT GGG CCT GGA ACC CCT GAG TCT Asn Ser Thr Asp Thr Thr Pro Leu Thr Gly Pro Gly Thr Pro Glu Ser 65 70 75 80	299
ACC ACT GTG GAG CCT GCT GCA AGG CGT TCT ACT GGC CTG GAT GCA GGA Thr Thr Val Glu Pro Ala Ala Arg Arg Ser Thr Gly Leu Asp Ala Gly 85 90 95	347
GGG GCA GTC ACA GAG CTG ACC ACG GAG CTG GCC AAC ATG GGG AAC CTG Gly Ala Val Thr Glu Leu Thr Thr Glu Leu Ala Asn Met Gly Asn Leu 100 105 110	395
TCC ACG GAT TCA GCA GCT ATG GAG ATA CAG ACC ACT CAA CCA GCA GCC Ser Thr Asp Ser Ala Ala Met Glu Ile Gln Thr Thr Gln Pro Ala Ala 115 120 125	443
ACG GAG GCA CAG ACC ACT CCA CTG GCA GCC ACA GAG GCA CAG ACA ACT Thr Glu Ala Gln Thr Thr Pro Leu Ala Ala Thr Glu Ala Gln Thr Thr 130 135 140	491
CGA CTG ACG GCC ACG GAG GCA CAG ACC ACT CCA CTG GCA GCC ACA GAG Arg Leu Thr Ala Thr Glu Ala Gln Thr Thr Pro Leu Ala Ala Thr Glu 145 150 155 160	539
GCA CAG ACC ACT CCA CCA GCA GCC ACG GAA GCA CAG ACC ACT CAA CCC Ala Gln Thr Thr Pro Pro Ala Ala Thr Glu Ala Gln Thr Thr Gln Pro 165 170 175	587
ACA GCG CTG GAG GCA CAG ACC ACT GCA CCA GCA GCG ATG GAG GCA CAG Thr Gly Leu Glu Ala Gln Thr Thr Ala Pro Ala Ala Met Glu Ala Gln 180 185 190	635
ACC ACT GCA CCA GCA GCC ATG GAA GCA CAG ACC ACT CCA CCA GCA GCC Thr Thr Ala Pro Ala Ala Met Glu Ala Gln Thr Thr Pro Pro Ala Ala 195 200 205	683
ATG GAG GCA CAG ACC ACT CAA ACC ACA GCG ATG GAG GCA CAG ACC ACT Met Glu Ala Gln Thr Thr Gln Thr Thr Ala Met Glu Ala Gln Thr Thr 210 215 220	731
GCA CCA GAA GCC ACG GAG GCA CAG ACC ACT CAA CCC ACA GCG ACC GAG Ala Pro Glu Ala Thr Glu Ala Gln Thr Thr Gln Pro Thr Ala Thr Glu 225 230 235 240	779
GCA CAG ACC ACT CCA CTG GCA GCC ATG GAG GCG CTG TCC ACA GAA CCC Ala Gln Thr Thr Pro Leu Ala Ala Met Glu Ala Leu Ser Thr Glu Pro 245 250 255	827

AGT GGC ACA GAG GCC CTG TCC ATG GAA CCT ACT ACC AAA AGA GGT CTG Ser Ala Thr Glu Ala Leu Ser Met Glu Pro Thr Thr Lys Arg Gly Leu 260 265 270	878
TTC ATA GCC TTT TCT GTG TCC TCT GTT ACT CAC AAG GGC ATT CCC ATG Phe Ile Pro Phe Ser Val Ser Ser Val Thr His Lys Gly Ile Pro Met 275 280 285	923
GCA GCC AGC AAT TTG TCC GTC AAC TAC CCA GTG GGG GCC CCA GAC CAC Ala Ala Ser Asn Leu Ser Val Asn Tyr Pro Val Gly Ala Pro Asp His 290 295 300	971
ATC TCT GTG AAG CAG TGC CTG CTG GCC ATC CTA ATC TTG GCG CTG GTG Ile Ser Val Lys Gln Cys Leu Leu Ala Ile Leu Ile Leu Ala Leu Val 305 310 315 320	1019
GCC ACT ATC TTC TTC GTG TGC ACT GTG GTG CTG GCG GTC CGC CTC TCC Ala Thr Ile Phe Phe Val Cys Thr Val Val Leu Ala Val Arg Leu Ser 325 330 335	1067
CGC AAG GGC CAC ATG TAC CCC GTG CGT AAT TAC TCC CCC ACC GAG ATG Arg Lys Gly His Met Tyr Pro Val Arg Asn Tyr Ser Pro Thr Glu Met 340 345 350	1115
GTC TGC ATC TCA TCC CTG TTG CCT GAT GGG GGT GAG GGG CCC TCT GCC Val Cys Ile Ser Ser Leu Leu Pro Asp Gly Gly Glu Gly Pro Ser Ala 355 360 365	1163
ACA GCC AAT GGG GGC CTG TCC AAG GCC AAG AGC CCG GGC CTG ACG CCA Thr Ala Asn Gly Gly Leu Ser Lys Ala Lys Ser Pro Gly Leu Thr Pro 370 375 380	1211
GAG CCC AGG GAG GAC CGT GAG GGG GAT GAC CTC ACC CTG CAC AGC TTC Glu Pro Arg Glu Asp Arg Glu Gly Asp Asp Leu Thr Leu His Ser Phe 385 390 395 400	1259
CTC CCT TAGCTCACTC TGCCATCTGT TTTGGCAAGA CCCCACCTCC ACGGGCTCTC Leu Pro	1315
CTGGGGCCACC CTTGAGTGGC CAGACCCCAA TCCACAGCTC TGGGCTTCTT CCGAGACCCC	1375
TGGGGATGGG CATCTTCAGG GAAGGAACTC TGGCCACCCA AACAGGAGAA GAGCAGCCTC	1435
GGGCAAGCA CACGGGCAAG TGGAGCCACC TCTTTCTCTC CTCTCTGGAT GAAGCCAGC	1495
CACATTTGAG CCGAGGTCCA AGGCAGGAGG CCATTTACTT GAGACAGATT CTCTCTTTT	1555
TCTGTCCCC CATCTTCTCT GGGTCCCTCT AACATCTCCC ATGGCTCTCC CCGCTTCTCC	1615
TGGTCACTGG AGTCTCTCTC CCACTACCC AAGG	1649

(2) INFORMATION FOR SEQ ID NO. 1:

(11) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 400 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(12) MOLECULE TYPE: protein

(x1) SEQUENCE DESCRIPTION: SEQ ID NO:2:

```

Met Pro Leu Gln Leu Leu Leu Leu Ile Leu Leu Gly Pro Gly Asn
 1             5             10             15

Ser Leu Gln Leu Trp Asp Thr Trp Ala Asp Glu Ala Glu Lys Ala Leu
 20             25             30

Gly Pro Leu Leu Ala Arg Asp Arg Arg Gln Ala Thr Glu Tyr Glu Tyr
 35             40             45

Leu Asp Tyr Asp Phe Leu Pro Glu Thr Glu Pro Pro Glu Met Leu Arg
 50             55             60

Asn Ser Thr Asp Thr Thr Pro Leu Thr Gly Pro Gly Thr Pro Glu Ser
 65             70             75             80

Thr Thr Val Glu Pro Ala Ala Arg Arg Ser Thr Gly Leu Asp Ala Gly
 85             90             95

Gly Ala Val Thr Glu Leu Thr Thr Glu Leu Ala Asn Met Gly Asn Leu
100             105             110

Ser Thr Asp Ser Ala Ala Met Glu Ile Gln Thr Thr Gln Pro Ala Ala
115             120             125

Thr Glu Ala Gln Thr Thr Pro Leu Ala Ala Thr Glu Ala Gln Thr Thr
130             135             140

Arg Leu Thr Ala Thr Glu Ala Gln Thr Thr Pro Leu Ala Ala Thr Glu
145             150             155             160

Ala Gln Thr Thr Pro Pro Ala Ala Thr Glu Ala Gln Thr Thr Gln Pro
165             170             175

Thr Gly Leu Glu Ala Gln Thr Thr Ala Pro Ala Ala Met Glu Ala Gln
180             185             190

Thr Thr Ala Pro Ala Ala Met Glu Ala Gln Thr Thr Pro Pro Ala Ala
195             200             205

Met Glu Ala Gln Thr Thr Gln Thr Thr Ala Met Glu Ala Gln Thr Thr
210             215             220

Ala Pro Glu Ala Thr Glu Ala Gln Thr Thr Gln Pro Thr Ala Thr Glu
225             230             235             240

Ala Gln Thr Thr Pro Leu Ala Ala Met Glu Ala Leu Ser Thr Glu Pro

```

```

                245                250                255
Ser Ala Thr Glu Ala Leu Ser Met Glu Pro Thr Thr Lys Arg Gly Leu
      260                265                270
Phe Ile Pro Phe Ser Val Ser Ser Val Thr His Lys Gly Ile Pro Met
      275                280                285
Ala Ala Ser Asn Leu Ser Val Asn Tyr Pro Val Gly Ala Pro Asp His
      290                295                300
Ile Ser Val Lys Gln Cys Leu Leu Ala Ile Leu Ile Leu Ala Leu Val
      305                310                315                320
Ala Thr Ile Phe Phe Val Cys Thr Val Val Leu Ala Val Arg Leu Ser
      325                330                335
Arg Lys Gly His Met Tyr Pro Val Arg Asn Tyr Ser Pro Thr Glu Met
      340                345                350
Val Cys Ile Ser Ser Leu Leu Pro Asp Gly Gly Glu Gly Pro Ser Ala
      355                360                365
Thr Ala Asn Gly Gly Leu Ser Lys Ala Lys Ser Pro Gly Leu Thr Pro
      370                375                380
Glu Pro Arg Glu Asp Arg Glu Gly Asp Asp Leu Thr Leu His Ser Phe
      385                390                395                400
Leu Pro

```

(ii) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1239 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA (synthetic)

(iii) ORIGINAL SOURCE:

- (A) ORGANISM: Homo sapiens
- (G) CELL TYPE: placenta

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..1239

(xii) SEQUENCE DESCRIPTION: SEQ ID NO:3:

ATG GCT CTC CAA CTC CTC CTC TTG CTC ATC CTA CTC GCG GCT GCG AAC

42

Met	Pro	Leu	Gln	Leu	Leu	Leu	Leu	Ile	Leu	Leu	Gly	Pro	Gly	Asn	
1				5				10						15	
AGC	TTG	CAG	CTG	TGG	GAC	ACC	TGG	GCA	GAT	GAA	GCC	GAG	AAA	GCC	TTG
Ser	Leu	Gln	Leu	Trp	Asp	Thr	Trp	Ala	Asp	Glu	Ala	Glu	Lys	Ala	Leu
			20					25					30		
GGT	CCC	CTG	CTT	GCC	CGG	GAC	CGG	AGA	CAG	GCC	ACC	GAA	TAT	GAG	TAC
Gly	Pro	Leu	Leu	Ala	Arg	Asp	Arg	Arg	Gln	Ala	Thr	Glu	Tyr	Glu	Tyr
		35					40					45			
CTA	GAT	TAT	GAT	TTC	CTG	CCA	GAA	ACG	GAG	CCT	CCA	GAA	ATG	CTG	AGG
Leu	Asp	Tyr	Asp	Phe	Leu	Pro	Glu	Thr	Glu	Pro	Pro	Glu	Met	Leu	Arg
	50					55					60				
AAC	AGC	ACT	GAC	ACC	ACT	CCT	CTG	ACT	GGG	CCT	GGA	ACC	CCT	GAG	TCT
Asn	Ser	Thr	Asp	Thr	Thr	Pro	Leu	Thr	Gly	Pro	Gly	Thr	Pro	Glu	Ser
	65				70				75					80	
ACC	ACT	GTG	GAG	CCT	GCT	GCA	AGG	CCT	TCT	ACT	GGC	CTG	GAT	GCA	GGA
Thr	Thr	Val	Glu	Pro	Ala	Ala	Arg	Arg	Ser	Thr	Gly	Leu	Asp	Ala	Gly
			85					90					95		
GGG	GCA	GTC	ACA	GAG	CTG	ACC	ACG	GAG	CTG	GCC	AAC	ATG	GGG	AAC	CTG
Gly	Ala	Val	Thr	Glu	Leu	Thr	Thr	Glu	Leu	Ala	Asn	Met	Gly	Asn	Leu
		100						105					110		
TCC	ACG	GAT	TCA	GCA	GCT	ATG	GAG	ATA	CAG	ACC	ACT	CAA	CCA	GCA	GCC
Ser	Thr	Asp	Ser	Ala	Ala	Met	Glu	Ile	Gln	Thr	Thr	Gln	Pro	Ala	Ala
		115					120					125			
ACG	GAG	GCA	CAG	ACC	ACT	CAA	CCA	GTG	CCC	ACG	GAG	GCA	CAG	ACC	ACT
Thr	Glu	Ala	Gln	Thr	Thr	Gln	Pro	Val	Pro	Thr	Glu	Ala	Gln	Thr	Thr
	130					135					140				
CCA	CTG	GCA	GCC	ACA	GAG	GCA	CAG	ACA	ACT	CGA	CTG	ACC	GCC	ACC	GAG
Pro	Leu	Ala	Ala	Thr	Glu	Ala	Gln	Thr	Thr	Arg	Leu	Thr	Ala	Thr	Glu
	145				150					155				160	
GCA	CAG	ACC	ACT	CCA	CTG	GCA	GCC	ACA	GAG	GCA	CAG	ACC	ACT	CCA	CCA
Ala	Gln	Thr	Thr	Pro	Leu	Ala	Ala	Thr	Glu	Ala	Gln	Thr	Thr	Pro	Pro
				165					170					175	
GCA	GCC	ACG	GAA	GCA	CAG	ACC	ACT	CAA	GCC	ACA	GCC	CTG	GAG	GCA	CAG
Ala	Ala	Thr	Glu	Ala	Gln	Thr	Thr	Gln	Pro	Thr	Gly	Leu	Glu	Ala	Gln
		180					185					190			
ACC	ACT	GCA	CCA	GCA	GCC	ATG	GAG	GCA	CAG	ACC	ACT	GCA	CCA	GCA	GCC
Thr	Thr	Ala	Pro	Ala	Ala	Met	Glu	Ala	Gln	Thr	Thr	Ala	Pro	Ala	Ala
		195					200					205			
ATG	GAA	GCA	CAG	ACC	ACT	CCA	CCA	GCA	GCC	ATG	GAG	CCA	CAG	ACC	ACT
Met	Glu	Ala	Gln	Thr	Thr	Pro	Pro	Ala	Ala	Met	Glu	Ala	Gln	Thr	Thr
	210					215					220				

CAA ACC ACA GCC ATG GAG GCA CAG ACC ACT GCA CCA GAA GCC ACC GAG Gln Thr Thr Ala Met Glu Ala Gln Thr Thr Ala Pro Gln Ala Thr Glu 225 230 235 240	720
GCA CAG ACC ACT CAA CCC ACA GCC ACC GAG GCA CAG ACC ACT CCA CTG Ala Gln Thr Thr Gln Pro Thr Ala Thr Glu Ala Gln Thr Thr Pro Leu 245 250 255	768
GCA GCC ATG GAG GCC CTG TCC ACA GAA CCC AGT GCC ACA GAG GCC CTG Ala Ala Met Glu Ala Leu Ser Thr Glu Pro Ser Ala Thr Glu Ala Leu 260 265 270	816
TCC ATG GAA CCT ACT ACC AAA AGA GGT CTG TTC ATA CCC TTT TCT GTG Ser Met Glu Pro Thr Thr Lys Arg Gly Leu Phe Ile Pro Phe Ser Val 275 280 285	864
TCC TCT GTT ACT CAC AAG GGC ATT CCC ATG GCA GCC AGC AAT TTG TCC Ser Ser Val Thr His Lys Gly Ile Pro Met Ala Ala Ser Asn Leu Ser 290 295 300	912
GTC AAC TAC CCA GTG GGG GCC CCA GAC CAC ATC TCT GTG AAG CAG TGC Val Asn Tyr Pro Val Gly Ala Pro Asp His Ile Ser Val Lys Gln Cys 305 310 315 320	960
CTG CTG GCC ATC CTA ATC TTG GCG CTG GTG GCC ACT ATC TTC TTC GTG Leu Leu Ala Ile Leu Ile Leu Ala Leu Val Ala Thr Ile Phe Phe Val 325 330 335	1008
TGC ACT GTG GTG CTG GCG GTC CGC CTC TCC CGC AAG GGC CAC ATG TAC Cys Thr Val Val Leu Ala Val Arg Leu Ser Arg Lys Gly His Met Tyr 340 345 350	1056
CCC GTG CCT AAT TAC TCC CCC ACC GAG ATG GTC TGC ATC TCA TCC CTG Pro Val Arg Asn Tyr Ser Pro Thr Glu Met Val Cys Ile Ser Ser Leu 355 360 365	1104
TTG CCT GAT GGG GGT GAG GGG CCC TCT GCG ACA GCG AAT GGG GGC CTG Leu Pro Asp Gly Gly Glu Gly Pro Ser Ala Thr Ala Asn Gly Gly Leu 370 375 380	1152
TCC AAG GCC AAG AGC CCG GGC CTG ACC CCA GAG CCC AAG GAG GAG CCT Ser Lys Ala Lys Ser Pro Gly Leu Thr Pro Glu Pro Arg Gln Asp Arg 385 390 395 400	1200
GAG GGG GAT GAC CTC ACC CTG GAC AGC TTC CTC GGT TAG Glu Gly Asp Asp Leu Thr Leu His Ser Phe Leu Pro 405 410	1236

(2) INFORMATION FOR SEQ ID NO:4:

(A) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 410 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(x) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

```

Met Pro Leu Gln Leu Leu Leu Leu Ile Leu Leu Gly Pro Gly Asn
 1             5             10             15
Ser Leu Gln Leu Trp Asp Thr Trp Ala Asp Glu Ala Glu Lys Ala Leu
      20             25             30
Gly Pro Leu Leu Ala Arg Asp Arg Arg Gln Ala Thr Glu Tyr Glu Tyr
      35             40             45
Leu Asp Tyr Asp Phe Leu Pro Glu Thr Glu Pro Pro Glu Met Leu Arg
      50             55             60
Asn Ser Thr Asp Thr Thr Pro Leu Thr Gly Pro Gly Thr Pro Glu Ser
      65             70             75             80
Thr Thr Val Glu Pro Ala Ala Arg Arg Ser Thr Gly Leu Asp Ala Gly
      85             90             95
Gly Ala Val Thr Glu Leu Thr Thr Glu Leu Ala Asn Met Gly Asn Leu
      100            105            110
Ser Thr Asp Ser Ala Ala Met Glu Ile Gln Thr Thr Gln Pro Ala Ala
      115            120            125
Thr Glu Ala Gln Thr Thr Gln Pro Val Pro Thr Glu Ala Gln Thr Thr
      130            135            140
Pro Leu Ala Ala Thr Glu Ala Gln Thr Thr Arg Leu Thr Ala Thr Glu
      145            150            155            160
Ala Gln Thr Thr Pro Leu Ala Ala Thr Glu Ala Gln Thr Thr Pro Pro
      165            170            175
Ala Ala Thr Glu Ala Gln Thr Thr Gln Pro Thr Gly Leu Glu Ala Gln
      180            185            190
Thr Thr Ala Pro Ala Ala Met Glu Ala Gln Thr Thr Ala Pro Ala Ala
      195            200            205
Met Glu Ala Gln Thr Thr Pro Pro Ala Ala Met Glu Ala Gln Thr Thr
      210            215            220
Gln Thr Thr Ala Met Glu Ala Gln Thr Thr Ala Pro Glu Ala Thr Glu
      225            230            235            240
Ala Gln Thr Thr Gln Pro Thr Ala Thr Glu Ala Gln Thr Thr Pro Leu
      245            250            255
Ala Ala Met Glu Ala Leu Ser Thr Glu Pro Ser Ala Thr Glu Ala Leu
      260            265            270

```

Ser Met Glu Pro Thr Thr Lys Arg Gly Leu Phe Ile Pro Phe Ser Val
 275 280 285

Ser Ser Val Thr His Lys Gly Ile Pro Met Ala Ala Ser Asn Leu Ser
 290 295 300

Val Asn Tyr Pro Val Gly Ala Pro Asp His Ile Ser Val Lys Gln Cys
 305 310 315 320

Leu Leu Ala Ile Leu Ile Leu Ala Leu Val Ala Thr Ile Phe Phe Val
 325 330 335

Cys Thr Val Val Leu Ala Val Arg Leu Ser Arg Lys Gly His Met Tyr
 340 345 350

Pro Val Arg Asn Tyr Ser Pro Thr Glu Met Val Cys Ile Ser Ser Leu
 355 360 365

Leu Pro Asp Gly Gly Glu Gly Pro Ser Ala Thr Ala Asn Gly Gly Leu
 370 375 380

Ser Lys Ala Lys Ser Pro Gly Leu Thr Pro Glu Pro Arg Glu Asp Arg
 385 390 395 400

Glu Gly Asp Asp Leu Thr Leu His Ser Phe Leu Pro
 405 410

(2) INFORMATION FOR SEQ ID NO:5:

(1) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2151 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(12) IMMEDIATE SOURCE:

- (B) CLONE: pacesol

(13) SEQUENCE DESCRIPTION: SEQ ID NO:5:

ATGGAGCTGA GGCCTGTGTT GGTATGGGTG GTAGCAGCAA CAGGAACCTT GGTCTGTGTA 60

GCAGCTGATG CTCAGGGCCA GAAGGTCTTC ACCAACACGT GGGCTGTGCG CATCCCTGGA 120

GGCCACGGG TGGCCAACAG TGTGGCACGG AAGCATGGGT TCTCAACCT GGGCCAGATC 180

TTGGGGGACT ATTACCACTT CTGGCATCGA GGAATGACGA AGTGGTCTCT GTGGCTCTAC 240

GGCTGGCGCG ACAGCCGGCT GCAGAGGGAG CTTCAAGTAC AGTGGCTCGA ACAGCAGGTG 300

CCAAAGCCAC	GGACTAAACG	GGACGTGTAC	CAGSAGCCCA	CAGACCCCA	GTTCCTCAG	360
CAGTGGTACC	TGTCTGGTGT	CACTCAGCGG	GACCTGAATG	TGAAGGGGGC	CTGGCGGCAG	420
GGCTACACAG	GGCACGGCAT	TGTGGTCTCC	ATTCTGGACG	ATGGCATCGA	GAAGAACCCAC	480
CCGGACTTGG	CAGGCAATTA	TGATCCTGGG	GCCAGTTTTG	ATGTCAATGA	CCAGSACCCT	540
GACCCCCAGC	CTCGGTACAC	ACAGATGAAT	GACAACAGGC	ACGGCACACG	GTGTGCGGGG	600
GAAGTGGCTG	CGGTGGCCAA	CAACGGTGTC	TGTGGTGTAG	GTGTGGCCTA	CAACGCCCGC	660
ATTGGAGGGG	TGCGCATGCT	GGATGGCGAG	GTGACAGATG	CAGTGGAGGC	ACGCTCGCTG	720
GGCCTGAACC	CCAACCACAT	CCACATCTAC	AGTGCCAGCT	GGGGCCCCGA	GGATGACGGC	780
AAGACAGTGG	ATGGGGCCAGC	CCGCCTCGCC	GAGGAGGCCT	TCTTCGGTGG	GGTTAGCCAG	840
GGCCGAGGGG	GGCTGGGCTC	CATCTTTGTC	TGGGCCTCGG	GGAACGGGGG	CCGGGAACAT	900
GACAGCTGCA	ACTGCGACGG	CTACACCAAC	AGTATCTACA	CGCTGTCCAT	CAGCAGCGCC	960
ACGCAGTTTG	GCAACGTGCC	GTGGTACAGC	GAGGCCTGCT	CSTCCACACT	GGCCACGACC	1020
TACAGCAGTG	GCAACCAGAA	TGAGAAGCAG	ATCGTGACGA	CTGACTTGCG	GCAGAAGTGC	1080
ACGGAGTCTC	ACACGGGCAC	CTCAGCCTCT	GCCCCCTTAG	CAGCCGGCAT	CATTGCTCTC	1140
ACCCTGGAGG	CCAATAAGAA	CCTCACATGG	CGGGACATGC	AACACCTGGT	GGTACAGACC	1200
TGGAAGCCAG	CCCACCTCAA	TGCCAACGAC	TGGGCCACCA	ATGGTGTGGG	CCGGAAAGTG	1260
AGCCACTCAT	ATGGCTACGG	GCTTTTGGAC	GCAGGCGCCA	TGGTGGCCCT	GGCCGAGAAT	1320
TGGACCACAG	TGGCCCCCCA	GGGGAAGTGC	ATCATCGACA	TGCTCACCGA	GGCCAAAGAC	1380
ATCGGGAAAC	GGCTCGAGGT	GGGGAAGACT	GTGACCGGCT	GGCTGGGCGA	GGCCAAACAC	1440
ATCACTCGGC	TGGAGCACGC	TCAGGCGGGG	CTCACCTGT	CCTATAATGG	CGGTGGCGAC	1500
CTGGCCATCC	ACCTGGTCAG	CCCCATGGGC	ACCCGCTCCA	CCCTGCTGCG	AGCCAGGCCA	1560
CATGACTACT	CGGCAGATGG	GTTTAATBAC	TGGGCCTTCA	TGACAACTCA	TTCTTGGGAT	1620
GAGGATCCCT	CTGGCGAGTG	GGTCTTAGAG	ATTGAAAACA	CCAGCGAAGC	CAACAACTAT	1680
GGGACGCTGA	CCAAGTTCAC	CCTCGTACTC	TATGGCAACG	CCCCTGAGGG	GCTGCCCCGA	1740
CCTCCAGAAA	GCAGTGGCTG	CAAGACCTCT	ACGTCCAGTC	AGGCCTGTGT	GGTGTGCGAG	1800
GAAGGCTTCT	CCCTGCACCA	GAAGAGCTGT	GTCCAGCACT	GGCCTCCAGG	CTTGGCCCCC	1860
CAAGTCTCTG	ATACCCACTA	TAGCACCAGG	AATGACGTGG	AGACCATCCG	GGTCAGCCTC	1920
TGGCCCCCTT	GGCACGGCTC	ATGTGCCACA	TGCCAGGGGC	TGGCTGTGAC	AGATTGCTCT	1980

AGGTGCTGCA GGCACGCTC GTTGGACCTT GTGGAGCAGA GTTGGTCTGG GCAAAGCCAG 2040
 AGCAGGCGAG AGTCCCCCGC ACAGCAGCAG CCACCTGGGC TGCCCCCGGA GGTGGAGGCG 2100
 GGGCAACGGC TGCGGGCAGG GCTGCTGCCC TCACACCTGC CTGAGTGATG A 2151

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1587 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Homo sapiens

(vii) IMMEDIATE SOURCE:

- (B) CLONE: sPSL.F2

(x1) SEQUENCE DESCRIPTION: SEQ ID NO:6:

ATGCCTCTGC AACTCCTCCT GTTGCTGATC CTACTGGGCG CTGGCAACAG CTTGCAGCTG 60
 TGGGACACCT GGGCAGATGA AGCCGAGAAA GCCTTGGGTC CCCTGCTTGC CCGGGACCGG 120
 AGACAGGCCA CCGAATATGA GTACCTAGAT TATGATTTCC TGCCAGAAAC GGAGCCTCCA 180
 GAAATGCTGA GGAACAGCAC TGACACCACT CTTCTGACTG GGCCTGGAAC CCTTGAGTCT 240
 ACCACTCTGG AGCCTGCTGC AAGGCGTTCT ACTGGCCTGG ATGCAGGAGG GGCAGTCACA 300
 GAGCTGACCA CGGAGCTGGC CAACATGGGG AACCTGTCCA TGGATTGAGC AGCTATGGAG 360
 ATACAGACCA CTCACCCAGC AGCCACGGAG GCACAGACCA CTCGACTGGC AGCCACAGAG 420
 GCACACACAA CTCGACTGAC GGCACCGGAG GCACAGACCA CTCGACTGGC AGCCACAGAG 480
 GCACAGACCA CTCGACTGAG AGCCACGGAA GCACAGACCA CTCGACTGAG AGCCCTGGAG 540
 GCACAGACCA CTCGACTGAG AGCCATGGAG GCACAGACCA CTCGACTGAG AGCCATGGAA 600
 GCACAGACCA CTCGACTGAG AGCCATGGAG GCACAGACCA CTCGACTGAG AGCCATGGAG 660
 GCACAGACCA CTCGACTGAG AGCCACGGAG GCACAGACCA CTCGACTGAG AGCCACGGAG 720
 GCACAGACCA CTCGACTGAG AGCCATGGAG GGCCTGTCCA CAGAAAGGAG TGGCAGAGAG 780
 GGCCTGTCCA TGGAACTTAC TACCAAAAGA GGTCTGTTCA TACCCCTTTT TGTGTCTCTT 840
 GTTACTTADA AGGGCATTCT CATGGCAGCC AGCAATTTGT CCGTCTCTGG GCGGCAGTCT 900

ADAGACAAAA CTCACACATG CCCACCGTGC CCAGCACCTG AACTCCTGGG GGGACCGTCA 1080
 GTCTTCTCTT TCCCCCAAAA ACCCAAGGAC ACCCTCATGA TCTCCCGGAC CCCTGAGGTC 1090
 ACATGCGTGG TGSTGGACGT GAGCCACGAA GACCCTGAGG TCAAGTTCAA CTGGTACGTG 1098
 GACGGCGTGG AGGTGCATAA TSCCAAGACA AAGCCGCGGG AGGAGCAGTA CAACAGCACG 1140
 TACCGTGTGG TCAGCGTCCT CACCGTCCTG CACCAGGACT GGCTGAATGG CAAGGAGTAC 1200
 AAGTGCAAGG TCTCCAACAA AGCCCTCCCA GTCCCCATCG AGAAAACCAT CTCCAAAGCC 1260
 AAAGGGCAGC CCCGAGAACC ACAGGTGTAC ACCCTGCCCC CATCCCGGGA GGAGATGACC 1320
 AAGAACCAGG TCAGCCTGAC CTGCCTGGTC AAAGGCTTCT ATCCCAGCGA CATCGCCGTG 1380
 GAGTGGGAGA GCAATGGGCA GCCGGAGAAC AACTACAAGA CCACGCCTCC CGTGCTGGAC 1440
 TCCGACGGCT CCTTCTTCCT CTATAGCAAG CTCACCGTGG ACAAGAGCAG GTGGCAGCAG 1500
 GGGAACGTCT TCTCATGCTC CGTGATGCAT GAGGCTCTGC ACAACCACTA CACGCAGAAG 1560
 AGCCTCTCCC TGTCCCCGGG TAAATAG 1587

(2) INFORMATION FOR SEQ ID NO:7:

- (1) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 19 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(X1) SEQUENCE DESCRIPTION: SEQ ID NO:7:

AATTCCTCG ACTCTAGAG 13

(2) INFORMATION FOR SEQ ID NO:8:

- (1) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 15 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(X1) SEQUENCE DESCRIPTION: SEQ ID NO:8:

CTCTAGACTC GACGG 15

(2) INFORMATION FOR SEQ ID NO:9:

- (1) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 43 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(x1) SEQUENCE DESCRIPTION: SEQ ID NO:9:

TAGCATACGC TCTAGAGCAT GGATCCCCCTG GGTGCAGCCA AGC

43

(2) INFORMATION FOR SEQ ID NO:10:

- (1) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 27 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(x1) SEQUENCE DESCRIPTION: SEQ ID NO:10:

CCGGAATTCT CAGGTGAACC AAGCCGC

27

(2) INFORMATION FOR SEQ ID NO:11:

- (1) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 25 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(x1) SEQUENCE DESCRIPTION: SEQ ID NO:11:

AAGTATCTGT CCAGGGCTTC CAGGT

25

(2) INFORMATION FOR SEQ ID NO:12:

- (1) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 48 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(X1) SEQUENCE DESCRIPTION: SEQ ID NO:12:

AAC TACCCAG TGGGAGCACC AGACCACATC TCTGTGAACC AGTGCTAG
48

(2) INFORMATION FOR SEQ ID NO:13:

- (1) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 52 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(X1) SEQUENCE DESCRIPTION: SEQ ID NO:13:

AAT TCTAGCA CTGCTTCACA GAGATGTGGT CTGGTGCTCC CACTGGGTAG TT 52

(2) INFORMATION FOR SEQ ID NO:14:

- (1) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 45 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(X1) SEQUENCE DESCRIPTION: SEQ ID NO:14:

AAC TACCCAG TGGGAGCACC AGACCACATC TCTGTGAACC AGTAG 45

(2) INFORMATION FOR SEQ ID NO:15:

- (1) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 49 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(X1) SEQUENCE DESCRIPTION: SEQ ID NO:15:

AATTCTACTG GTTCACAGAG ATGTGGTCTG GTGTCCTCCAC TGGGTAGTT

49

(2) INFORMATION FOR SEQ ID NO:16:

- (1) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 52 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(X1) SEQUENCE DESCRIPTION: SEQ ID NO:16:

CTAGACCCCG GATGGCATCC ATGACAGGAG GACAACAAAT GGTAGGCCGT AG

50

(2) INFORMATION FOR SEQ ID NO:17:

- (1) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 53 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(X1) SEQUENCE DESCRIPTION: SEQ ID NO:17:

AATTCTACGG CCTACCCATT TGTGTCTCTC CTGTCATGGA TGCCATCCTG GGT

51

(2) INFORMATION FOR SEQ ID NO:18:

- (1) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 13 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(X1) SEQUENCE DESCRIPTION: SEQ ID NO:18:

CTGCGGCCCG AGT

13

(2) INFORMATION FOR SEQ ID NO:19:

- (1) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 17 base pairs
 - (B) TYPE: nucleic acid

(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(x1) SEQUENCE DESCRIPTION: SEQ ID NO:19:

CTAGACTGCG GCCGCAG

17

(2) INFORMATION FOR SEQ ID NO:20:

(1) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 50 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(x1) SEQUENCE DESCRIPTION: SEQ ID NO:20:

CCAGGTCCAA CTGCAGGTCG ACTCTAGAGG GCACTTCTTC TGGGCCCCAG

50

(2) INFORMATION FOR SEQ ID NO:21:

(1) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 50 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(x1) SEQUENCE DESCRIPTION: SEQ ID NO:21:

TATTATCTGT GCGGCCGCCG TCCAGAACCC ATGGCTGCTG GTTGCACTCG

50

(2) INFORMATION FOR SEQ ID NO:22:

(1) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 40 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(x1) SEQUENCE DESCRIPTION: SEQ ID NO:22:

TATTATCTGT GCGGCCGCGC AGCAGGCTCC ACAGTGGTAG

40

(2) INFORMATION FOR SEQ ID NO:23:

(1) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 38 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(x1) SEQUENCE DESCRIPTION: SEQ ID NO:23:

TATTATCTGT GCGGCCGCGG AGGCTCCGTT TCTGGCAG

38

(2) INFORMATION FOR SEQ ID NO:24:

(1) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 33 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(x1) SEQUENCE DESCRIPTION: SEQ ID NO:24:

CGGAGACAGG CCACCGAATT CCTGCCAGAA ACG

33

(2) INFORMATION FOR SEQ ID NO:25:

(1) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 40 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(x1) SEQUENCE DESCRIPTION: SEQ ID NO:25:

CCTCCAGAAA TGGTGAGGCA CAGCACTGAC ACCACTGCTC

40

(2) INFORMATION FOR SEQ ID NO:26:

- (1) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 40 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(x1) SEQUENCE DESCRIPTION: SEQ ID NO:26:

GAGCTGGCCA ACATGGGGCA ACTGTCCACG GATTCAGCAG

40

(2) INFORMATION FOR SEQ ID NO:27:

- (1) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 21 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(x1) SEQUENCE DESCRIPTION: SEQ ID NO:27:

AATTCGAGTT CCTAGATTTT G

21

(2) INFORMATION FOR SEQ ID NO:28:

- (1) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 21 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(x1) SEQUENCE DESCRIPTION: SEQ ID NO:28:

AATTCAAAT CTAGGAACTC G

21

(2) INFORMATION FOR SEQ ID NO:29:

- (1) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 48 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(X1) SEQUENCE DESCRIPTION: SEQ ID NO:29:

AATTCGAGTA CCTAGATTAT GATTTCCTGC CAGAAACTGA GCCTCCGC

48

(2) INFORMATION FOR SEQ ID NO:30:

(1) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 48 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(X1) SEQUENCE DESCRIPTION: SEQ ID NO:30:

GGCCGCGGAG GCTCAGTTTC TGGCAGGAAA TCATAATCTA GGTACTCG

48

(2) INFORMATION FOR SEQ ID NO:31:

(1) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 48 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(X1) SEQUENCE DESCRIPTION: SEQ ID NO:31:

AATTCGAGTA CCTAGATTAT GATTTCCTGC CAGAAACTGA GCCTCCGC

48

(2) INFORMATION FOR SEQ ID NO:32:

(1) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 48 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(X1) SEQUENCE DESCRIPTION: SEQ ID NO:32:

GGCCGCGGAG GCTCAGTTTC TGGCAGGAAA TCATAATCTA GGAAGTCC

48

(2) INFORMATION FOR SEQ ID NO:33:

- (1) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 48 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

AATTCGAGTT CCTAGATTTC GATTTCCTGC CAGAAACTGA GCCTCCGC

48

(2) INFORMATION FOR SEQ ID NO:34:

- (1) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 48 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:

GGCCCGCGAG GCTCAGTTTC TGGCAGGAAA TCGAAATCTA GGAACTCS

48

(2) INFORMATION FOR SEQ ID NO:35:

- (1) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 942 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

1X FEATURE:

- (A) NAME/KEY: CDS
(B) LOCATION: 1..939

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:

ATGCCTGTGC AACTCCTCCT GTTGCTGATC CTACTGGGCC CTGGCAACAG GTTGCAGCTG
TGGGACACCT GGGCAGATGA AGCCGAGAAA GCCTTGGGTC CCCTGCTTGC CCGGGACCGG

50

120

```

AGACAGGCCA CGGAATATGA GTACCTAGAT TATGATTTCC TGCCAGAAAC GGAGCCTCCA      120
GAAATGCTGA GGAACAGCAC TGACACCACT CCTCTGACTG GGCCTGGAAC CCTTGAGTCT      240
ACCACTGTGG AGCCTGCTGC GCGGCCGCAC ACATGCCCCAC CGTGCCCAGC ACCTGAAGCC      300
CTGGGGGGCAC CGTCAGTCTT CCTCTTCCCC CCAAAACCCA AGGACACCCCT CATGATCTCC      360
CGGACCCCTG AGGTCACATG CGTGGTGGTG GACGTGAGCC ACGAAGACCC TGAGGTCAAG      420
TTCAACTGGT ACGTGGACGG CGTGGAGGTG CATAATGCCA AGACAAAGCC GCGGGAGGAG      480
CAGTACAACA GCACGTACCG TGTGGTCAGC GTCCTCACCG TCCTGCACCA GGA CTGGCTG      540
AATGGCAAGG AGTACAAGTG CAAGGTCTCC AACAAAGCCC TCCCAGTCCC CATCGAGAAA      600
ACCATCTCCA AAGCCAAAGG GCAGCCCCGA GAACCACAGG TGTACACCCT GCGCCCATCC      660
CGGGAGGAGA TGACCAAGAA CCAGGTCAGC CTGACCTGCC TGGTCAAAGG CTTCTATCCC      720
AGCGACATCG CCGTGGAGTG GGAGAGCAAT GGGCAGCCGG AGAACAATA CAAGACCACG      780
CCTCCCCTGC TGGACTCCGA CGGCTCCTTC TTCCTCTATA GCAAGCTCAC CGTGGACAAG      840
AGCAGGTGGC AGCAGGGGAA CGTCTTCTCA TGCTCCGTGA TGCATGAGGC TCTGCACAAC      900
CACTACACGC AGAAGAGCCT CTCCTGTCC CCGGGTAAAT GA                          942

```

(2) INFORMATION FOR SEQ ID NO:36:

(1) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 313 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS:
- (D) TOPOLOGY: linear

(2) MOLECULE TYPE: protein

(3) SEQUENCE DESCRIPTION: SEQ ID NO:36:

```

Met Pro Leu Gln Leu Leu Leu Leu Ile Leu Leu Gly Pro Gly Asn
1           5           10           15
Ser Leu Gln Leu Trp Asp Thr Trp Ala Asp Glu Ala Glu Cys Ala Leu
20           25           30
Gly Pro Leu Leu Ala Arg Asp Arg Arg Gln Ala Thr Glu Tyr Glu Tyr
35           40           45
Leu Asp Tyr Asp Phe Leu Pro Glu Thr Glu Pro Pro Glu Met Leu Arg
50           55           60

```

Asn Ser Thr Asp Thr Thr Pro Leu Thr Gly Pro Gly Thr Pro Glu Ser
 65 70 75 80
 Thr Thr Val Glu Pro Ala Ala Arg Pro His Thr Cys Pro Pro Cys Pro
 85 90 95
 Ala Pro Glu Ala Leu Gly Ala Pro Ser Val Phe Leu Phe Pro Pro Lys
 100 105 110
 Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val
 115 120 125
 Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr
 130 135 140
 Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu
 145 150 155 160
 Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His
 165 170 175
 Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys
 180 185 190
 Ala Leu Pro Val Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln
 195 200 205
 Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Glu Glu Met
 210 215 220
 Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro
 225 230 235 240
 Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn
 245 250 255
 Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu
 260 265 270
 Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val
 275 280 285
 Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr Gln
 290 295 300
 Lys Ser Leu Ser Leu Ser Pro Gly Lys
 305 310

(2) INFORMATION FOR SEQ ID NO:27:

(1) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 310 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(1x) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 1..807

(x1) SEQUENCE DESCRIPTION: SEQ ID NO:37:

```

ATGCTCTGTC AACTCCTCCT GTTGCTGATC CTACTGGGCG CTGGCAACAG CTTGCAGCTG      60
TGGGACACCT GGGCAGATGA AGCCGAGAAA GCCTTGGGTC CCCTGCTTGC CCGGGACCGG      120
AGACAGGCCA CCGAATATGA GTACCTAGAT TATGATTTCC TGCCAGAAAC GGAGCCTCCA      180
GAAATGCTGA GGAACAGCAC TGACACCACT CCTCTGACTG GGCTTGAAC CCCTGAGTCT      240
ACCACTGTGG AGCCTGCTGC GCGGCGGCTG TGTGCCAACC TAGTACCGGT GCCCATCACC      300
AACGCCACCC TGGACCAGAT CACTGGCAAG TGGTTTTATA TGGCATCGGC CTTTCGAAAC      360
GAGGAGTACA ATAAGTCGGT TCAGGAGATC CAAGCAACCT TCTTTTACTT CACCCCCAAC      420
AAGACAGAGG ACACGATCTT TCTCAGAGAG TACCAGACCC GACAGGACCA GTGCATCTAT      480
AACACCACCT ACCTGAATGT CCAGCGGGAA AATGGGACCA TCTCCAGATA CGTGGGAGGC      540
CAAGAGCATT TCGCTCACTT GCTGATCCTC AGGGACACCA AGACCTACAT GCTTGCTTTT      600
GACGTGAACG ATGAGAAGAA CTGGGGGCTG TCTGTCTATG CTGACAAGCC AGAGACGACC      660
AAGGAGCAAC TGGGAGAGTT CTACGAAGCT CTCGACTGCT TCGGCATTCC CAACTCAGAT      720
GTCGTGTACA CCGATTGGAA AAAGGATAAG TGTGAGCCAC TGGAGAAGCA GCACGAGAAG      780
GAGAGGAAAC AGGAGGAGGG GGAATCCTAG                                     840

```

(2) INFORMATION FOR SEQ ID NO:38:

(1) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 269 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS:

(D) TOPOLOGY: linear

(11) MOLECULE TYPE: protein

(x1) SEQUENCE DESCRIPTION: SEQ ID NO:38:

```

Met Pro Leu Gln Leu Leu Leu Leu Ile Leu Leu Gly Pro Gly Asp
1           5           10           15

```

Ser Leu Gln Leu Trp Asp Thr Trp Ala Asp Gln Ala Glu Lys Ala Leu
 20 25 30
 Gly Pro Leu Leu Ala Arg Asp Arg Arg Gln Ala Thr Glu Tyr Glu Tyr
 35 40 45
 Leu Asp Tyr Asp Phe Leu Pro Glu Thr Glu Pro Pro Glu Met Leu Arg
 50 55 60
 Asn Ser Thr Asp Thr Thr Pro Leu Thr Gly Pro Gly Thr Pro Glu Ser
 65 70 75 80
 Thr Thr Val Glu Pro Ala Ala Arg Pro Leu Cys Ala Asn Leu Val Pro
 85 90 95
 Val Pro Ile Thr Asn Ala Thr Leu Asp Gln Ile Thr Gly Lys Trp Phe
 100 105 110
 Tyr Ile Ala Ser Ala Phe Arg Asn Glu Glu Tyr Asn Lys Ser Val Gln
 115 120 125
 Glu Ile Gln Ala Thr Phe Phe Tyr Phe Thr Pro Asn Lys Thr Glu Asp
 130 135 140
 Thr Ile Phe Leu Arg Glu Tyr Gln Thr Arg Gln Asp Gln Cys Ile Tyr
 145 150 155 160
 Asn Thr Thr Tyr Leu Asn Val Gln Arg Glu Asn Gly Thr Ile Ser Arg
 165 170 175
 Tyr Val Gly Gly Gln Glu His Phe Ala His Leu Leu Ile Leu Arg Asp
 180 185 190
 Thr Lys Thr Tyr Met Leu Ala Phe Asp Val Asn Asp Glu Lys Asn Trp
 195 200 205
 Gly Leu Ser Val Tyr Ala Asp Lys Pro Glu Thr Thr Lys Glu Gln Leu
 210 215 220
 Gly Glu Phe Tyr Glu Ala Leu Asp Cys Leu Arg Ile Pro Lys Ser Asp
 225 230 235 240
 Val Val Tyr Thr Asp Trp Lys Lys Asp Lys Cys Glu Pro Leu Glu Lys
 245 250 255
 Gln His Glu Lys Glu Arg Lys Gln Glu Glu Gly Glu Ser
 260 265

(2) INFORMATION FOR SEQ ID NO:39:

(1) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1314 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(1x) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 1..1311

(x1) SEQUENCE DESCRIPTION: SEQ ID NO:39:

ATGGTGGCCG GGACCCGCTG TCTTCTAGCG TTGCTGCTTC CCCAGGTCCT CCTGGGCGGC	60
GCGGCTGGCC TCGTTCCGGA GCTGGGCGCG AGGAAGTTCTG CGGCGGCGTC GTGGGCGCGC	120
CCCTCATCCC AGCCCTCTGA CGAGGTCCTG AGCGAGTTCTG AGTTGCGGCT GCTCAGCATG	180
TTGGGCTGA AACAGAGACC CACCCCCAGC AGGGACGCGG TGGTGCCCCC CTACATGCTA	240
GACCTGTATC GCAGGCACTC AGGTCAGCCG GGCTCACCCG CCCCAGACCA CCGGTTGGAG	300
AGGGCAGCCA GCCGAGCCAA CACTGTGCGC AGCTTCCACC ATGAAGAATC TTTGGAAGAA	360
CTACCAGAAA CGAGTGGGAA AACAAACCCG AGATTCTTCT TTAATTTAAG TTCTATCCCC	420
ACGGAGGAGT TTATCACCTC AGCAGAGCTT CAGGTTTTTC GAGAACAGAT GCAAGATGCT	480
TTAGGAAACA ATAGCAGTTT CCATCACCGA ATTAATATTT ATGAAATCAT AAAACCTGCA	540
ACAGCCAACT CGAAATTCCC CGTGACCAGA CTTTGGGACA CCAGGTTGGT GAATCAGAAT	600
GCAAGCAGGT GGGAAAGTTT TGATGTCACC CCCGCTGTGA TGCGGTGGAC TGCACAGGGA	660
CACGCCAACC ATGGATTCGT GGTGGAAGTG GCCCACTTGG AGGAGAAACA AGGTGTCTCC	720
AAGAGACATG TTAGGATAAG CAGGTCTTTG CACCAAGATG AACACACCTG CTCACAGATA	780
AGGCCATTGC TAGTAACTTT TGGCCATGAT GGAAAAGGGC ATCCTCTCCA CAAAAGAGAA	840
AAACGTCAGG CCACCGAATA TGAGTACCTA GATTATGATT TCCTGCCAGA AACGGAGCCT	900
CCAGAAATGC TGAGGAACAG CACTGACACC AATCCTCTGA CTGGGCTCG AACCCCTGAG	960
TCTACCACTG TGGAGCCTGC TGCAAGGCGG AAACGCCTTA AGTCAGCTG TAAGAGACAG	1020
CCTTTGTACG TGGACTTCAG TGACGTGGGG TGGAAATGACT GGATTGTGGC TCCCGGGGG	1080
TATCAGCCT TTTACTGCCA CGGAGAATGC CCTTTTCCCTC TGGCTGATCA TGTGAAGTCC	1140
ACTAATCATG CCATTGTTCA GACGTTGGTC AACTCTGTTA ACTCTAAGAT TCCTAAGGCA	1200
TGCTGTGTCC CGACAGAACT CAGTGCTATC TCGATGCTGT ACCTTGACCA GAATGAAAAG	1260
GTTGTATTAA AGAACTATCA GGACATGGTT GTGGAGGGTT GTGGGTCTCG CTAG	1314

(2) INFORMATION FOR SEQ ID NO:40:

(ii) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 437 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS:
- (D) TOPOLOGY: linear

(iii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:

```

Met Val Ala Gly Thr Arg Cys Leu Leu Ala Leu Leu Leu Pro Gln Val
1           5           10           15

Leu Leu Gly Gly Ala Ala Gly Leu Val Pro Glu Leu Gly Arg Arg Lys
          20           25           30

Phe Ala Ala Ala Ser Ser Gly Arg Pro Ser Ser Gln Pro Ser Asp Glu
          35           40           45

Val Leu Ser Glu Phe Glu Leu Arg Leu Leu Ser Met Phe Gly Leu Lys
          50           55           60

Gln Arg Pro Thr Pro Ser Arg Asp Ala Val Val Pro Pro Tyr Met Leu
65           70           75           80

Asp Leu Tyr Arg Arg His Ser Gly Gln Pro Gly Ser Pro Ala Pro Asp
          85           90           95

His Arg Leu Glu Arg Ala Ala Ser Arg Ala Asn Thr Val Arg Ser Phe
          100          105          110

His His Glu Glu Ser Leu Glu Glu Leu Pro Glu Thr Ser Gly Lys Thr
          115          120          125

Thr Arg Arg Phe Phe Phe Asn Leu Ser Ser Ile Pro Thr Glu Glu Phe
          130          135          140

Ile Thr Ser Ala Glu Leu Gln Val Phe Arg Glu Gln Met Gln Asp Ala
145          150          155          160

Leu Gly Asn Asn Ser Ser Phe His His Arg Ile Asn Ile Tyr Glu Ile
          165          170          175

Ile Lys Pro Ala Thr Ala Asn Ser Lys Phe Pro Val Thr Arg Leu Leu
          180          185          190

Asp Thr Arg Leu Val Asn Gln Asn Ala Ser Arg Trp Glu Ser Phe Asp
          195          200          205

Val Thr Pro Ala Val Met Arg Trp Thr Ala Gln Gly His Ala Asn His
210          215          220

```

Gly Phe Val Val Glu Val Ala His Leu Glu Glu Lys Gln Gly Val Ser
 225 230 235 240
 Lys Arg His Val Arg Ile Ser Arg Ser Leu His Gln Asp Glu His Ser
 245 250 255
 Trp Ser Gln Ile Arg Pro Leu Leu Val Thr Phe Gly His Asp Gly Lys
 260 265 270
 Gly His Pro Leu His Lys Arg Glu Lys Arg Gln Ala Thr Glu Tyr Glu
 275 280 285
 Tyr Leu Asp Tyr Asp Phe Leu Pro Glu Thr Glu Pro Pro Glu Met Leu
 290 295 300
 Arg Asn Ser Thr Asp Thr Thr Pro Leu Thr Gly Pro Gly Thr Pro Glu
 305 310 315 320
 Ser Thr Thr Val Glu Pro Ala Ala Arg Arg Lys Arg Leu Lys Ser Ser
 325 330 335
 Cys Lys Arg His Pro Leu Tyr Val Asp Phe Ser Asp Val Gly Trp Asn
 340 345 350
 Asp Trp Ile Val Ala Pro Pro Gly Tyr His Ala Phe Tyr Cys His Gly
 355 360 365
 Glu Cys Pro Phe Pro Leu Ala Asp His Leu Asn Ser Thr Asn His Ala
 370 375 380
 Ile Val Gln Thr Leu Val Asn Ser Val Asn Ser Lys Ile Pro Lys Ala
 385 390 395 400
 Cys Cys Val Pro Thr Glu Leu Ser Ala Ile Ser Met Leu Tyr Leu Asp
 405 410 415
 Glu Asn Glu Lys Val Val Leu Lys Asn Tyr Gln Asp Met Val Val Glu
 420 425 430
 Gly Cys Gly Cys Arg
 435

(2) INFORMATION FOR SEQ ID NO:41:

(1) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 395 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(11) MOLECULE TYPE: cDNA

(16) FEATURES:

- (1) NAME KEY: CDS

(B) LOCATION: 1...792

(x1) SEQUENCE DESCRIPTION: SEQ ID NO:41:

```

ATGCCTCTGC AACTCCTCCT GTTGCTGATC CTA CTGGGGCC CTGGCAACAG CTTGCAGCTG      60
TGGGACACCT GGGCAGATGA AGCCGAGAAA GCCTTGGGTC CCCTGCTTGC CCGGGACCCG      120
AGACAGGCCA CCGAATATGA GTACCTAGAT TATGATTTCC TGCCAGAAAC GGAGCCTCCA      180
GAAATGCTGA GGAACAGCAC TGACACCACT CCTCTGACTG GGCCTGGAAC CCCTGAGTCT      240
ACCACTGTGG AGCCTGCTGC GCGGCCGCCA CCTGGCCCCC CTCGAGTTTC CCCAGACCCT      300
CGGGCCGAGC TGGACAGCAC CGTGCTCCTG ACCCGCTCTC TCCTGGCGGA CACGCGGCAG      360
CTGGCTGCAC AGCTGAGGGA CAAATTCCCA GCTGACGGGG ACCACAACCT GGATTCCCTG      420
CCCACCTGG CCATGAGTGC GGGGGCACTG GGAGCTCTAC AGCTCCCAGG TGTGCTGACA      480
AGGCTGCGAG CGGACCTACT GTCCTACCTG CGGCACGTGC AGTGGCTGCG CCGGGCAGGT      540
GGCTCTTCCC TGAAGACCCT GGAGCCCGAG CTGGGCACCC TGCAGGCCCC ACTGGACCGG      600
CTGCTGCGCC GGCTGCAGCT CCTGATGTCC CGCCTGGCCC TGCCCCAGCC ACCCCCGGAC      660
CCGCCGGCGC CCCCCTGGC GCCCCCTCC TCAGCCTGGG GGGGCATCAG GGCCGCCAC      720
GCCATCCTGG GGGGGCTGCA CCTGACACTT GACTGGGCCG TGAGGGGACT GCTGCTGCTG      780
AAGACTCGGC TGTGA
                                                                                   795

```

(2) INFORMATION FOR SEQ ID NO:42:

(1) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 264 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS:
- (D) TOPOLOGY: linear

(11) MOLECULE TYPE: protein

(x1) SEQUENCE DESCRIPTION: SEQ ID NO:42:

```

Met Pro Leu Gln Leu Leu Leu Leu Ile Leu Leu Gly Pro Gly Asn
1           5           10           15
Ser Leu Gln Leu Trp Asp Thr Trp Ala Asp Glu Ala Glu Lys Ala Leu
20           25           30

```

Gly Pro Leu Leu Ala Arg Asp Arg Arg Gln Ala Thr Glu Tyr Glu Tyr
 35 40 45
 Leu Asp Tyr Asp Phe Leu Pro Glu Thr Glu Pro Pro Glu Met Leu Arg
 50 55 60
 Asn Ser Thr Asp Thr Thr Pro Leu Thr Gly Pro Gly Thr Pro Glu Ser
 65 70 75 80
 Thr Thr Val Glu Pro Ala Ala Arg Pro Pro Pro Gly Pro Pro Arg Val
 85 90 95
 Ser Pro Asp Pro Arg Ala Glu Leu Asp Ser Thr Val Leu Leu Thr Arg
 100 105 110
 Ser Leu Leu Ala Asp Thr Arg Gln Leu Ala Ala Gln Leu Arg Asp Lys
 115 120 125
 Phe Pro Ala Asp Gly Asp His Asn Leu Asp Ser Leu Pro Thr Leu Ala
 130 135 140
 Met Ser Ala Gly Ala Leu Gly Ala Leu Gln Leu Pro Gly Val Leu Thr
 145 150 155 160
 Arg Leu Arg Ala Asp Leu Leu Ser Tyr Leu Arg His Val Gln Trp Leu
 165 170 175
 Arg Arg Ala Gly Gly Ser Ser Leu Lys Thr Leu Glu Pro Glu Leu Gly
 180 185 190
 Thr Leu Gln Ala Arg Leu Asp Arg Leu Leu Arg Arg Leu Gln Leu Leu
 195 200 205
 Met Ser Arg Leu Ala Leu Pro Gln Pro Pro Pro Asp Pro Pro Ala Pro
 210 215 220
 Pro Leu Ala Pro Pro Ser Ser Ala Trp Gly Gly Ile Arg Ala Ala His
 225 230 235 240
 Ala Ile Leu Gly Gly Leu His Leu Thr Leu Asp Trp Ala Val Arg Gly
 245 250 255
 Leu Leu Leu Leu Lys Thr Arg Leu
 260

Fig. 1

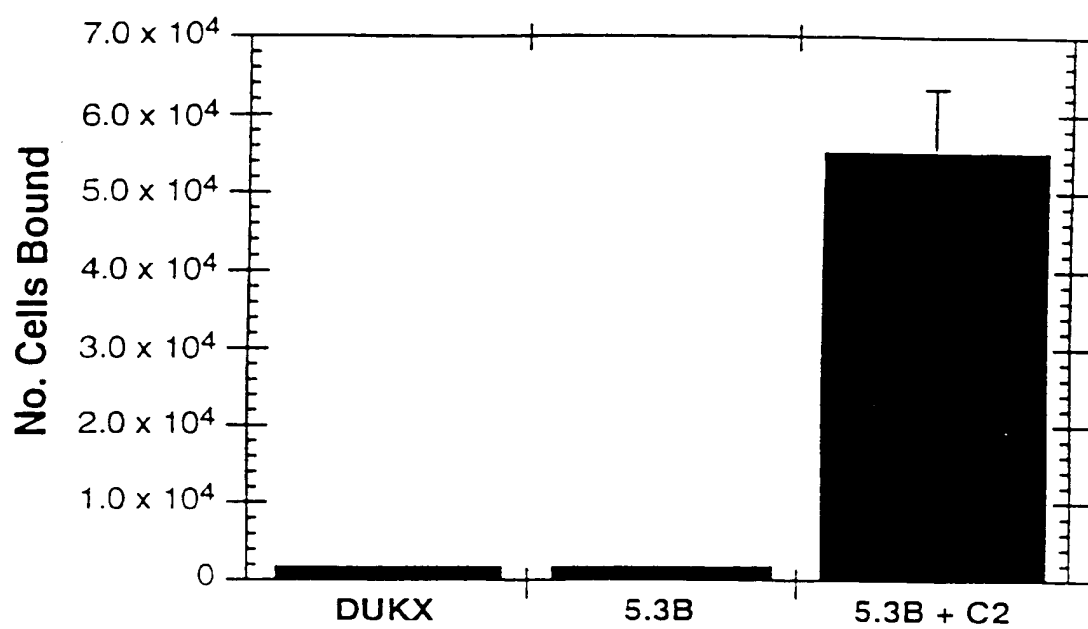
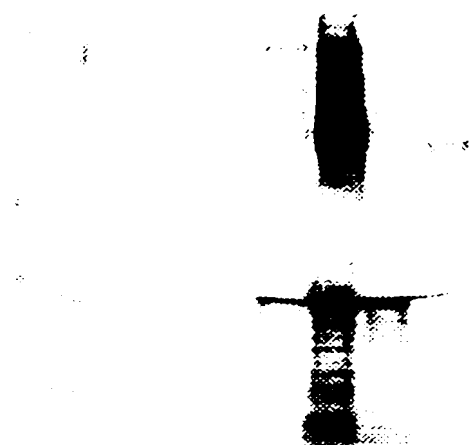


Fig. 2

poly PSL Ab		Lec γ 1	
-	+	-	+



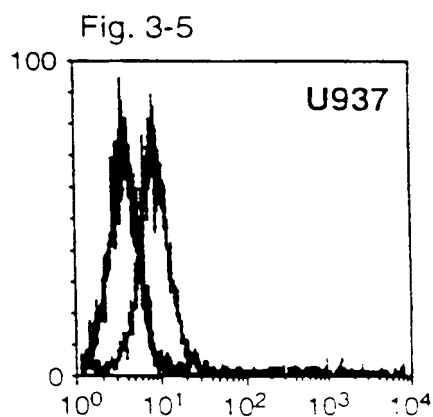
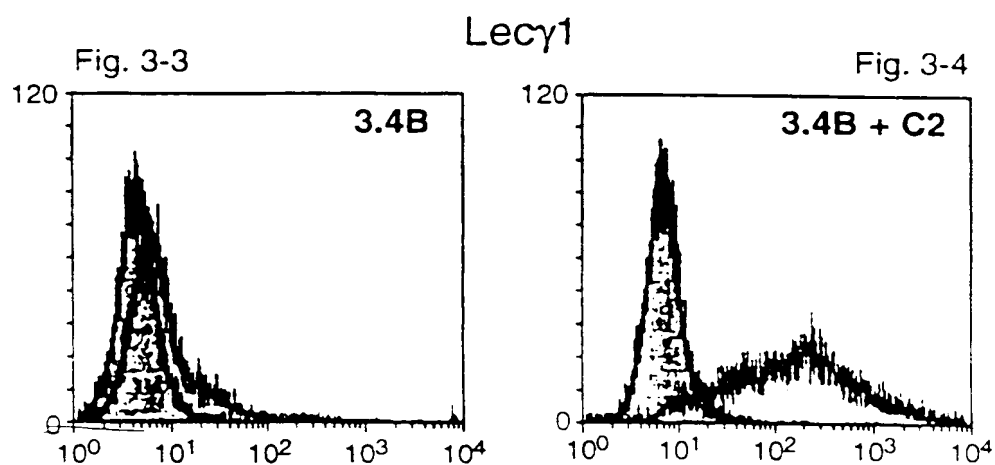
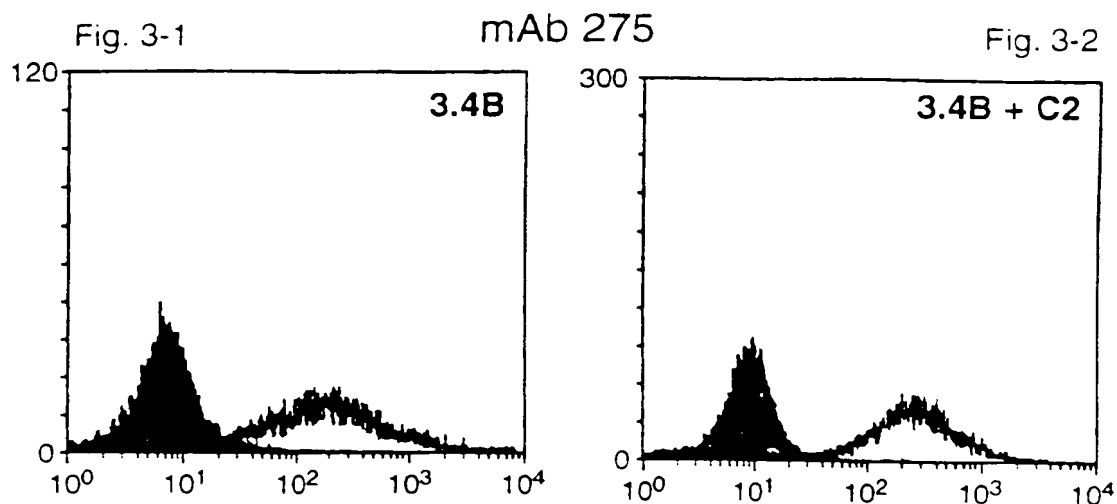


Fig. 4

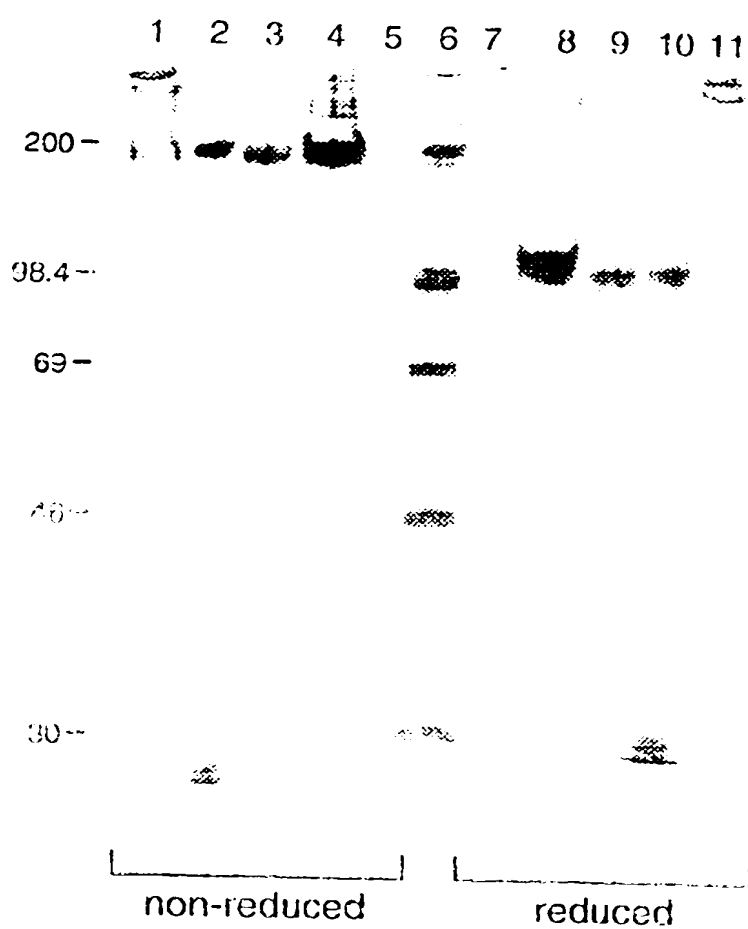
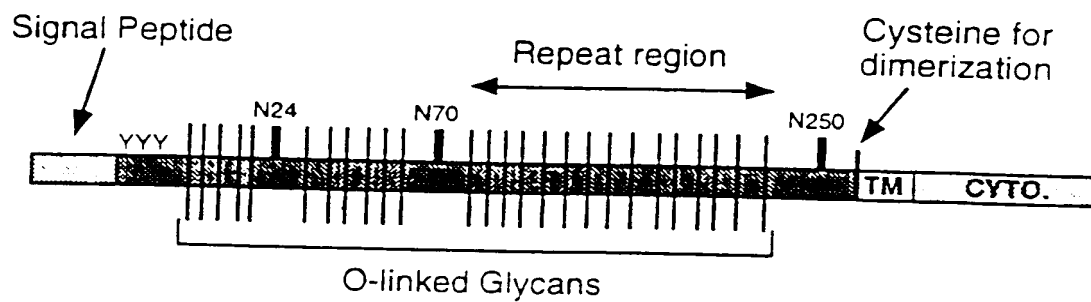


Fig. 5



Soluble "T7" Truncated Form:



Fig. 6

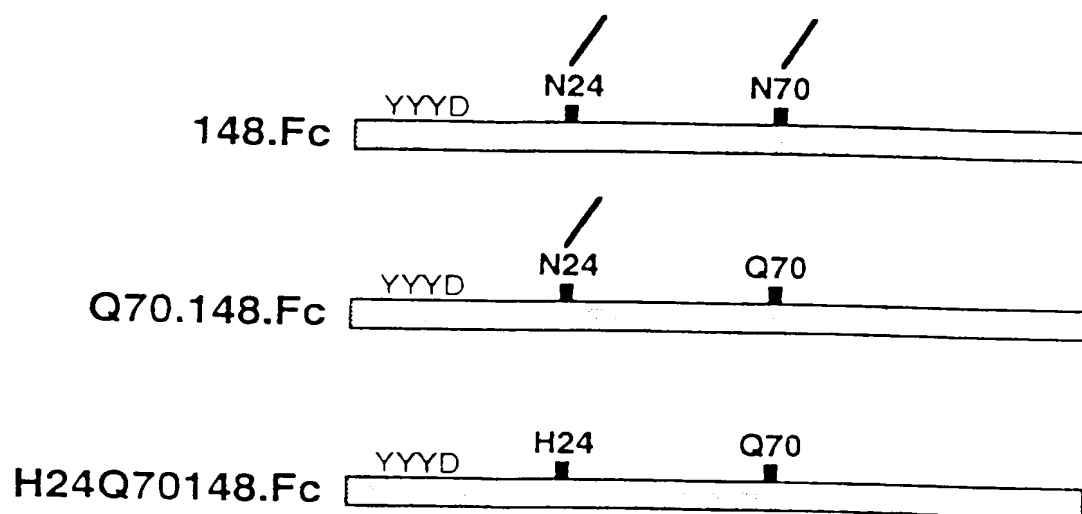


Fig. 7

1 2 3 4 5 6



³⁵S-Met Labeled

Captured with Protein A

1. 148.Fc
2. 148.Q70.Fc
3. 148.H24.Q70.Fc

Captured with P-Sel.Fc

4. 148.Fc
5. 18.Q70.Fc
6. 148.H24.Q70.Fc

Fig. 8

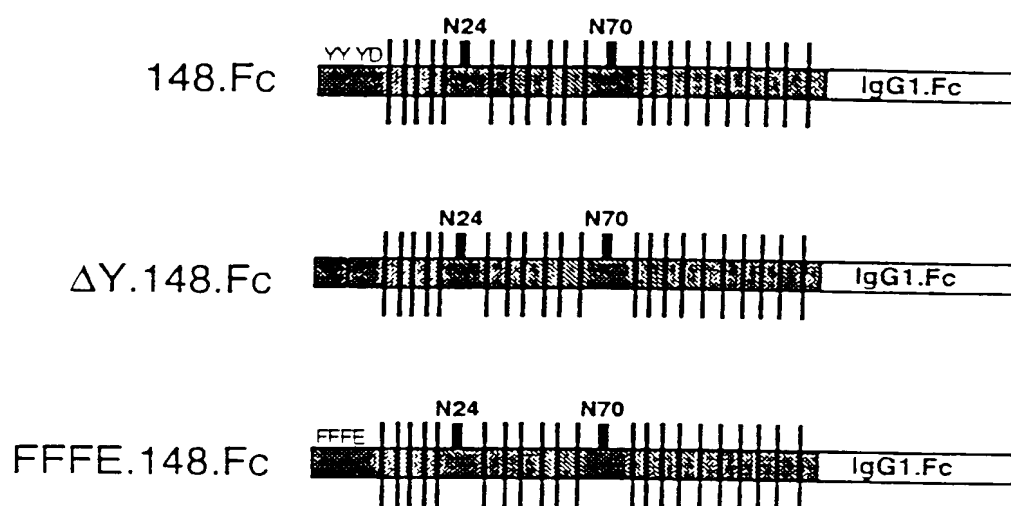
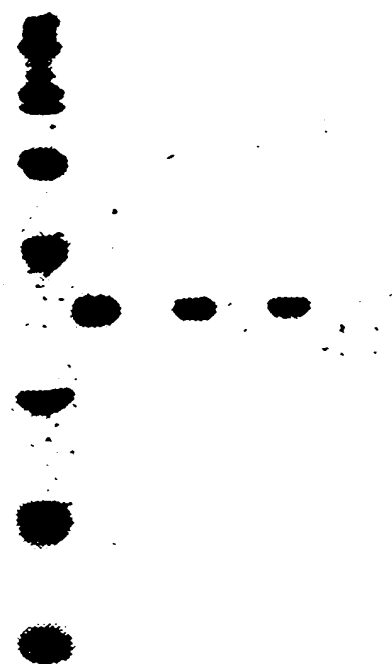


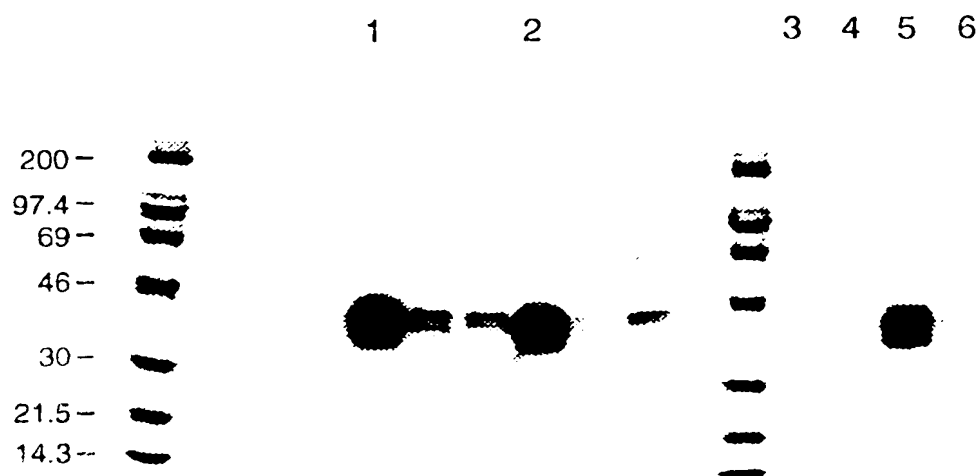
Fig. 9

M 1 2 3 4 5 6 7 8



Sample	Control
M. mw marker	
1. YYY.19Fc	+
2. YYY.19Fc	+
3. FYY.19Fc	+
4. FYY.19Fc	+
5. FFY.19Fc	+
6. FFY.19Fc	+
7. FFF.19Fc	+
8. FFF.19Fc	+

Fig. 10



^{35}S labeled

1. P-SEL.19.Fc

2. FFF.19.Fc

$^{35}\text{SO}_4$ Labeled

3. Mock

4. P-SEL.19.Fc

5. FYY.19.Fc

6. FFF.19.Fc

100

101

102

103

104

105

106

107

108

109

110

111

112

113

114

115

116

117

118

119

120

121

122

123

124

125

126

127

128

129

130

131

132

133

134

135

136

137

138

139

140

141

142

143

144

145

146

147

148

149

150

151

152

153

154

155

156

157

158

159

160

161

162

163

164

165

166

167

168

169

170

171

172

173

174

175

176

177

178

179

180

181

182

183

184

185

186

187

188

189

190

191

192

193

194

195

196

197

198

199

200